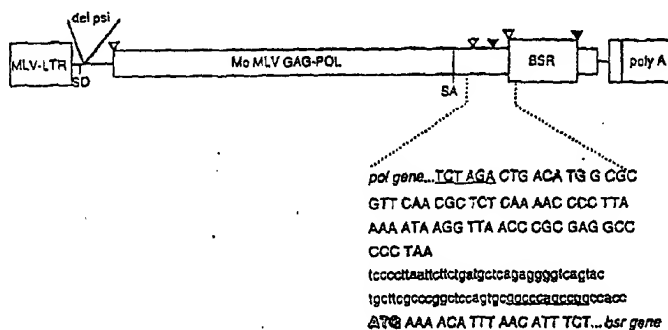




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(71) Applicant (for all designated States except US): CANCER RESEARCH CAMPAIGN TECHNOLOGY LIMITED [GB/GB]; Cambridge House, 6-10 Cambridge Terrace, Regent's Park, London NW1 4JL (GB).			
(72) Inventors; and (75) Inventors/Applicants (for US only): COLLINS, Mary, Katharine, Levinge [GB/GB]; Flat 4, Philips House, 52 Goodge Street, London W1P 1FP (GB). WEISS, Robin, Anthony [GB/GB]; 25 Cyprus Avenue, London N3 1SS (GB). TAKEUCHI, Yasuhiro [JP/GB]; 141 Elborough Street, London SW18 5DS (GB). COSSET, François-Lois [FR/FR]; 32, rue L.-Thévenet, F-69004 Lyon (FR).			
(74) Agents: CALDERBANK, T., R. et al.; Mewburn Ellis, York House, 23 Kingsway, London WC2B 6HP (GB).		Published With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.	

(54) Title: EXPRESSION SYSTEMS



Schematic structure of CaB expression vector

(57) Abstract

The invention relates to new expression systems and in particular to an expression system in which a gene of interest is expressed at an optimal level. The invention provides a recombinant expression vector comprising a gene of interest and a selectable marker gene, wherein the selectable marker gene is arranged downstream of the gene of interest and a stop codon associated with the gene of interest is spaced from a start codon of said selectable marker gene at a distance which is sufficient to ensure that translation reinitiation is required before said selectable marker protein is expressed from the corresponding mRNA. Examples of such expression systems are vector viral packaging cell lines and a number of preferred cell lines have been identified.

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Expression systems

The present invention relates to new expressions systems,
5 and in particular to expression systems in which a gene of
interest is expressed at an optimal level. Particular
examples of such expression systems are retroviral packaging
cell lines and a number of preferred cell lines have been
identified.

10 The ability of eukaryotic and prokaryotic ribosomes to
reinitiate translation at an internal start codon within an
mRNA sequence has previously been recognised. Studies have
been reported in which the efficiency of the process, which
15 is generally regarded as being low, has been connected with
the length of the intercistronic sequence (Kozak (1987)
Mol. Cell Biol. 7, 3438-3445). Selection of this sequence
or spacer as 70bp in length, and containing no other start
codons, has been previously reported as being optimal for
20 reinitiation in a eukaryotic cell line (Cosset F-L.,
Virology (1991) 185, 862).

The applicants have found a way in which the inefficiency
associated with the translation reinitiation process can be
25 used to good effect.

According to the present invention there is provided a
recombinant expression vector comprising a gene of interest
and a selectable marker gene, wherein the selectable marker
30 gene is arranged downstream of the gene of interest and a
stop codon associated with the gene of interest is spaced
from a start codon of said selectable marker gene at a
distance which is sufficient to ensure that translation re-
initiation is required before said selectable marker protein
35 is expressed from the corresponding mRNA.

The invention further provides a process for producing cell lines in which a gene of interest is expressed, which process comprises transforming host cells with an expression vector comprising said gene of interest and a selectable
5 marker gene, wherein the selectable marker gene is arranged downstream of the gene of interest and a stop codon associated with the gene of interest is spaced from a start codon of said selectable marker gene at a distance which is sufficient to ensure that translation re-initiation is
10 required before said selectable marker protein is expressed from the corresponding mRNA, and selecting those cells where expression of the selectable marker gene may be detected.

Since re-initiation of translation is a relatively
15 inefficient process, this means that the selectable marker protein will be expressed at lower levels than the product of the gene of interest. When the marker protein is expressed at detectable levels, the gene of interest will be expressed at higher levels. This will ensure that during
20 the subsequent selection procedure, only those cell clones which express the gene of interest at higher or optimal levels will survive. Low expressing clones will be eliminated by the selection process.

25 Cells transformed with the above-described expression vectors form a further aspect of the invention.

The host cells are suitably eukaryotic or prokaryotic host cells, preferably eukaryotic host cells.

30 The number of nucleotides in the space between the stop codon of the gene of interest and the start codon of the selectable marker will suitably be in the range of from 20-200 nucleotides, preferably from 60-80 nucleotides, even
35 more preferably 70-80 nucleotides.

The vectors used in the process of the invention may be any of the known types, for example expression plasmids or viral vectors.

5 Selected cells may be cultured and if required, the protein product of the gene of interest isolated from the culture using conventional techniques. Alternatively, expression of the gene of interest may result in other desired effects, for example, where the gene of interest is included as part
10 of a viral packaging construct.

Some experimental and clinical gene transfer protocols require the design of gene transfer vectors suitable for in vivo gene delivery (Miller, A.D. 1992. Nature 357:455-
15 460). Retroviral vectors are attractive candidates for such applications, because they can provide stable gene transfer and expression (Samarut J. et al., Meth. Enzymol. in press) and because packaging cells have been designed which produce non-replication competent viruses (Miller A.D (1990)
20 Hum Gene Ther. 1 5-14). However currently available recombinant retroviruses suffer from a number of drawbacks.

Packaging cell lines provide in trans the retroviral proteins encoded by the gag, pol, and env genes required to
25 obtain infectious retroviral particles. The gag and pol products are respectively the structural components of the virion cores and the replication machinery (enzymes) of the retroviral particles whereas the env products are envelope proteins responsible for the host-range of the virions and
30 for the initiation of infection and for sensitivity to humoral factors. An ideal packaging cell line should produce retroviruses that only contain the retroviral vector genome, and absolutely no replication-competent genomes or defective genomes encoding some of the viral structural
35 genes.

A number of packaging cell lines designed for human gene transfer have been designed in the past by introducing plasmid DNAs which contain "helper genomes" encoding gag, pol and/or env genes into cells.

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Retroviral packaging cell lines are cells that have been engineered to provide in trans all the functions required to express infectious retroviral vectors. A helper genome (or construct or unit), is herein also referred to as "retroviral packaging construct (or unit)," or "packaging-deficient construct (or genome unit)," or "gag-pol/env expression plasmids".

10

Much efforts has been made to design strategies to optimize the helper-genomes in order (i) to get the highest production of retroviral packaging functions (which correlates which infection titers of retroviral particles) and (ii) to minimise the chance that the helper genome can be transmitted via the viral particles (which may lead to emergence of unwanted retroviral forms).

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The first of these packaging cell lines used full length retroviral genomes as helper genomes that had been crippled for important cis-regulated replicative functions (reviewed in Miller, Hum. Gene. Ther. 1:5-14 1990). In order to reduce the possibility of occurrence of replication-competent viruses and of transfer of virus structural genes, a second generation of safer packaging cell lines has been designed by using two separate and complementary helper genomes which express either gag-pol or env and are packaging-deficient (Miller supra).

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The cells into which these helper genomes were introduced were isolated by cotransfecting them with plasmids encoding selectable markers. However, as no selection was applied on

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the packaging-deficient retroviral genome itself, the helper functions can be lost during the passages of the cells in culture and the current packaging systems provide limited titers of infectious retroviral vectors, usually only of the order of 10^5 - 10^6 infectious units i.u./ml. Indeed the cotransfection with a plasmid encoding a selectable marker does not directly select the best gag-pol-env-expressing cells.

The invention further provides a retroviral packaging cell line comprising a host cell transformed with (i) a packaging deficient construct which expresses a viral gag-pol gene and a first selectable marker gene, and/or (ii) a packaging-deficient construct which expresses a viral env gene and a second selectable marker gene; wherein a start codon of the first and second selectable markers are spaced from the stop codons of the viral gag-pol gene and the viral env gene respectively by a distance which ensures that reinitiation of mRNA translation is required for expression of marker protein product of said first and/or second selectable marker gene.

The retroviral packaging cell line may be obtained by the above described process which will involve selecting transfected cells which express said first and/or second marker genes.

By using helper constructs which are directly selectable and which provide for high expression of the viral gene, high titre retroviral vectors may be obtained.

Helper constructs for use in the process form a further aspect of the invention.

The retroviral vectors prepared from the conventional

packaging cell lines are usually not contaminated by replication-competent retroviruses (RCRs). However, recombinant amphotropic murine retroviruses have been shown to arise spontaneously from certain packaging cell lines.

5 The generation of such RCRs involves recombination at least between gag-pol/env packaging sequence and vector sequences (Cosset et al., Virology, (1993) 193:385-395).

Recombinant RCRs have been associated with the development of lymphomas in some severely immunosuppressed monkeys (Donahue et al., J. Exp Med (1992) 176: 1125-1135). In addition, retroviral vector preparations may also contain, at low frequencies, retroviruses coding for functional envelope glycoproteins (Kozak and Kabat, 1990, J. Virol. 64: 3500-3508) or for gag-pol proteins. Although the

15 pathogenicity of these gag-pol or env recombinant retroviruses is probably low, more evolved recombinant retroviruses with higher pathogenic potential may occur when injected in vivo, by recombination and/or complementation of

20 the initial recombinant viruses with some endogenous retroviruses.

In a preferred embodiment of the retroviral packaging cell lines of the invention, the overlapping sequences between the genomes of the retroviral vector and the helper

25 construct are reduced, for example as compared to constructs such as CRIPenv and CRIPAMgag (Danos et al., Proc. Natl. Acad. Sci USA 85: 6460-6464). In particular, the viral sequences in the helper construct are reduced, for example,

30 not only the packaging sequence but also the 3' Long Terminal Repeat (LTR), the 3' non-coding sequence and/or the 5' LTR may be eliminated.

The possibility of generation of such RCRs and recombinant retroviruses can be reduced by reducing the overlapping

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sequences between the genomes of both the retroviral vector and the helper construct.

Conventional retroviral vectors are strongly inactivated by human serum which makes them of limited or no use for in situ gene transfer in gene therapy applications. It has previously been shown that inactivation by complement in human serum is controlled by the cell line used to produce the virions and by viral envelope determinants (Takeuchi et al., J. Virol (1994) 68:8001-8007). In particular, inactivation is caused by some properties of the cell lines that have been used to construct the packaging cells (NIH-3T3) and also by viral determinants located in the retroviral envelope as shown (Takeuchi et al., J. Virol (1994) 68:8001-8007). In vivo gene delivery is an important goal for a number of human gene therapy strategies.

The applicants have found that certain cell lines form preferred packaging cell lines.

Particularly preferred packaging cell lines are the HT1080 line, the TE671 line, the 3T3 line, the 293 line and the Mv-1-Lu line. One example of retroviral packaging cells that will produce complement-resistant virus comprise human HT1080 cells and express RD114 envelope. Such cells form a preferred aspect of the invention.

Packaging cell lines according to the invention provide 50-100 fold increased titers of retroviral vectors as compared to conventional packaging cell lines. Retroviral vectors provided by these new cells are safe, in terms of generation of RCRs, and considerably more resistant to inactivation by human complement.

Packaging cell lines according to the invention may be able

to transduce helper-free, human complement-resistant retroviral vectors at titers consistently higher than 10^7 i.u./ml.

5 Suitable semi-packaging cell lines in accordance with the invention are those which express only the gag-pol genes. Such cell lines may suitably be derived from TE671, MINK Mv-1-Lu, HT1080, 293 or NIH-3T3 cells by introduction of plasmid CeB (the MoMLV gag-pol expression unit).

10 Particularly preferred expression vectors in accordance with the invention for use in retroviral packaging cell lines are those which include MLV gag and pol genes such as CeB. Other plasmids may include gag and pol genes from other
15 retroviruses or chimeric or mutated gag and pol genes.

Various viral and retroviral envelope genes may be included in the plasmids such as MLV-A envelope, GALV envelope, VSV-G protein, BaEV envelope, RD114 envelope and chimeric or
20 mutated envelopes. Plasmids which include the RD114 env gene such as FBdelPRDSAF as illustrated hereinafter, provide one example of suitable constructs.

25 The novel retroviral packaging cells described hereinafter, have been designated FLY cells, and may be designed for in vivo gene delivery.

30 Considerable variations were found between the various cell lines screened for their ability to release type C mammalian retroviruses. In addition, few cell lines were able to produce retroviruses completely resistant to human complement. Based on these two criteria, human fibrosarcoma HT1080 and rhabdomyosarcoma TE671 cells were selected for optimum construction of packaging cells.

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Other studies have shown the importance of endogenous retrovirus expression in the generation of recombinant retroviruses from retroviral packaging lines (Ronfort et al., Virology, (1995), 207, 271-275, Vanin, E.F. et al., J Virol (1994) 68:4241-4250.). The co-packaging of an endogenous genome and a vector can lead to emergence of recombinant retroviruses (Vanin et al., supra).

Recombination involves template switching during reverse transcription of such hybrid retroviruses (Hu et al., Science, (1990) 250:1227) and homologies between the two genomes considerably enhance the frequency of reverse transcriptase jumps (Zhang et al., J. Virol. (1994) 68: 2409-2414). Therefore an ideal packaging cell line should not express endogenous MLV-like (or type C retrovirus-like) retroviral genomes which can be packaged by type C gag proteins (Scadden et al., J. Virol. (1990) 64: 424-427, Torrent et al., J. Mol. Biol. (1994) 240 434-444).

Packaging of human endogenous retroviral RNA was not detected in TELCeB and FLY packaging cells when virion associated RNA was analysed by RT-PCR using generic primers.

HT1080- and TE671 derived packaging cell lines may be safer in this respect than those generated from NIH3T3 cells, such as GP+EAM12 cells, which are known to express and package sequences related to type C retroviruses (Scadden et al. supra).

To generate the FLY packaging cell lines, HT1080 cells were transfected with gag-pol and env expression plasmids designed to optimise viral protein expression. Direct selection for viral gene expression was achieved in accordance with the invention by expression of a selectable marker gene by re-initiation of translation of the mRNA expressing the viral proteins. This strategy resulted in packaging cell lines capable of producing extremely high

titer viruses. Furthermore, long-term expression of packaging functions can be maintained in these cells. Many unnecessary viral sequences were eliminated from the packaging constructs to reduce the risk of helper virus generation; indeed the final packaging cells did not produce helper virus, in that no replication competent virus (RCR) could be detected per 10^7 vector particles.

The FLY packaging cells described herein are safer than, for example, psiCRIP cells, at least for generation of env recombinant retroviruses as is illustrated in Table 4 hereinafter, probably because less retroviral sequences overlapping with the vector were present in the present env-expression plasmid. Few reports have addressed the question of the characterization of recombinant retroviruses (RVs) (Cosset, F.L., et al., Virology (1993) 193:385-395). It is possible that such RVs could not be detected in previous packaging cell lines due to lower overall titers. RVs are defective in normal cell culture conditions but are likely to evolve to replication competent viruses if they are allowed to replicate in cells complementing their expression like co-cultivated packaging cells (Bestwick et al., Proc. Natl Acad Sci USA, (1988) 85: 5404-5408, Cosset et al., (1993) supra).

In preferred retroviral packaging systems according to the invention, RVs are eradicated for example by removal of viral LTRs from the packaging construct.

Consistent with our previous studies (Takeuchi, Y., et al., J Virol (1994) 68:8001-8007), LacZ(RD114) and lacZ(MLV-A) pseudotypes produced from HT1080 and TE671 cells were more resistant to human complement than LacZ(RD114) or LacZ(MLV-A) pseudotypes produced by 3T3 or dog cells. It was therefore decided to use RD114 and MLV-A env genes to

generate recombinant virions with MoMLV cores.

The sequence of RD114 env gene was determined and is shown in Figure 4. It was found to be very close to BaEV (baboon endogenous virus) a type C retrovirus (Benveniste, R.E. et al., Proc. Natl. Acad. Sci. USA (1973) 70:3316-3320; Kato, S. et al., Japan. J. Genet. (1987) 62:127-137) with an envelope gene displaying similarities to the external part of type D simian retroviruses (SRVs). RD114 uses the SRV receptor on human cells (Sommerfelt & Weiss, Virology (1990) 176:58-69; Sommerfelt, M.A. et al., J Virol (1990) 64:6214-6220) making the FLY packaging cells with RD114 envelope capable of generating virions with different tropism. Retroviral vectors prepared so far for human gene therapy have used either MLV-A or GALV (gibbon ape leukemia virus) envelopes which display some similarities (Battini, J.L., et al., J Virol. (1992) 66:1468-1475) and which use two related cell surface receptors for infection (Miller, D.G. et al., J Virol (1994) 68:8270-8276). Differences in tissue-specific expression of MLV-A or GALV receptors have been reported (Kavanaugh et al., Proc Natl Acad Sci USA (1994) 91:7071-7075).

The invention will now be particularly described by way of example with reference to the accompanying drawings in which:

Figure 1. illustrates the structure and expression of CeB. The env gene (XbaI-Clal) of plasmid pCRIP was removed and was replaced by coininsertion of the two fragments XbaI-SfiI (restriction sites underlined) from pOXEnv and a SfiI-Clal PCR product containing the bsr selectable marker. This results in positioning the bsr start codon (shadowed) 74 bp downstream to the pol stop codon (bold).

Open triangle are start codons (gag and bsr), black triangles are stop codons (pol and bsr). The shadowed triangle is the start codon of env, in the same reading frame with that of bsr. SD and SA are the splice donor and splice acceptor sites.

Figure 2 illustrates the structure and expression of FbdelpASAF.

Immediately after the stop codon of env (bold) was inserted a non retroviral KasI-NcoI (restriction sites underlined) linker which positions the phleo start codon (shadowed) 76 bp downstream.

Open triangle are start codons (env and phleo), black triangles are stop codons (env and phleo). SD and SA are the splice donor and splice acceptor sites.

Figure 3 illustrates plasmids for expression of Amphi, Eco, RD114, Xeno, 10A1, GALV, VSV-G and FeLV envelopes.

All genes are expressed in the same backbone as detailed in fig. 2. The BglII sites for ecotropic (MoMLV strain), 10A1, xenotropic (NZB.1.V6 strain) and amphotropic (4070A strain), the NdeI site of RD114 (SC3C strain, the BamHI site for both FeLV and GALV were used as 5' ends, and linked to MscI site immediately after the splice donor site in the leader of FB29 LTR.

Figure 4 shows the sequence of the RD114 env gene (SEQ ID No 1).

Figure 5 shows the genetic structure of gag-pol constructs. Initiation (☉) and termination (▼) codons are shown. The thick dotted line below each construct shows MLV-derived sequences. Nucleotide positions of MLV-derived sequences are shown according to: Shinnick et al. (1981) (from nt 1 to nt 6000 with deletion of the packaging signal (DY) from BalI

(nt 215) to PstI (nt 568), and with some further MoMLV sequences in both CeB and CeB DS- from nt 7676 to nt 7938. gag-pol and bsr genes were expressed from the same transcription unit using the either a retroviral promoter (Mo LTR) or a non retroviral promoter (hCMV) and non retroviral polyadenylation sequence (polyA). Splice donor (SD) and acceptor (SA) sites are indicated. The thin line denotes retroviral non coding sequences. The thick line shows the rabbit beta-1 globin intron B. The position of some restriction sites is indicated.

The nucleic acid sequences of portions of constructs (as shown in Figure 5 (boxed areas)) are displayed for CeB (SEQ ID No 2, Figure 6), hCMV+intron (SEQ ID No 3, Figure 7) and hCMV+intronkaSD (SEQ ID No 4, Figure 8).

The nucleic acid sequences of portions of constructs (as shown in Figure 3 (boxed areas)) are displayed for FbdelpASAF (SEQ ID No 5, Figure 9), FbdelpMOSAF (SEQ ID No 6, Figure 10), FbdelpGASAF (SEQ ID No 7, Figure 11), FbdelpRDSAF (SEQ ID No 8, Figure 12) and CMV10A1 (SEQ ID No 9, Figure 13) are shown.

The components of the viral particles are produced by two independent expression plasmids (gag-pol or env) which also contain selectable markers (bsr or phleo) expressed from the same transcriptional units as gag-pol or env (figs. 1& 2). The selectable markers are located downstream to gag-pol or env genes and there is an optimal distance between the stop codon of the upstream reading frames and the start codon of the selectable genes that should allow re-initiation of translation (Kozak, Mol Cell Biol. (1987) 7, :3438-3445). Because there is no "Kozak" sequence (Kozak, Cell, (1986) 44: 283-292) required for a normal initiation of translation for

the marker gene, they can only be expressed by re-initiation of translation after the upstream viral gene has been successfully expressed. Consequently and also because re-initiation of translation is a poorly efficient process, after transfection of these plasmids, cells resistant to the drugs corresponding to those selectable genes express high levels of the viral proteins.

To avoid viral transmission of these "helper" genomes the constructs used suitably have the classical deletions of both the packaging sequence located in the leader region and of the 3'LTR, the latter being replaced by SV40 polyadenylation sequences (Figs 1 & 2).

Plasmid CeB is the MoMLV gag-pol-expression unit. It derives from pCRIP, a plasmid used to generate the constructs introduced in the CRIP and CRE packaging cell lines (Danos and Mulligan, 1988). As shown in fig. 1 for generation of plasmid CeB the env gene of pCRIP has been deleted mostly and the bsr selectable marker, -encoding a protein conferring resistance to blasticidin (Izumi et al., Experimental Cell Research (1991) 197, 229-233)- has been inserted downstream to pol gene. There are exactly 74 bp with no ATG triplets between the stop codon of pol and the start codon of bsr, this allows its expression by re-initiation of translation on the gag-pol mRNA, after translation of the gag-pol reading frame.

FbdelpASAF is a plasmid expressing the amphotropic env gene and the phleo selectable marker conferring resistance to phleomycin (Gatignol et al., FEBS Letters (1988) 230:171-175). By using a PCR-mediated mutagenesis strategy which modifies the end of env gene (see fig. 2), a 76 bp linker was inserted between the stop codon of env and the start codon of phleo. This allows expression of phleo from the

env mRNA by re-initiation of translation. In addition compared to known env-expressing constructs, this strategy of construction has reduced the length of sequences overlapping with the ends of conventional retroviral vectors. The env genes of Mo-MLV, FeLVB, NZB.1V6, 10A1, GALV and RD114 are expressed by plasmids FBdelPMoSAF, FBdelPBSAF, FBdelXSAF, FBdelpGSAF, FBdelp10A1SALF and FBdelPRDSAF, respectively, by using the same backbone as FBdelPASAF (fig. 3). Retroviral vectors produced with the RD114 envelope will be useful for in vivo gene delivery as comparatively to MLV ecotropic or amphotropic envelopes, virions pseudotyped with RD114 envelopes are not inactivated by human complement when they are produced by Mink Mv-1-Lu cells or by some human cells (Table 1).

The HT1080 cell line, isolated from a human fibrosarcoma (ATCC CCL121). The TE671 cell line isolated from a human rhabdomyosarcoma (ATCC CRL 8805) (purchased from ATCC, and tested for absence of usual cell culture contaminants by ECACC), has been used for the definitive construction of packaging cell lines. HT1080 line was chosen among a panel of primate and human lines because MLV-A and RD114 efficiently rescued retroviral vectors from these cells and also because RD114 pseudotypes produced by this cell line were stable when incubated in human serum. In a standard assay (Takeuchi et al., J Virol (1994), 68, 8001-8007), these latter viruses were found more than 500 fold more stable than similar pseudotypes produced in 3T3 cells.

Another advantage for the use of non murine cells to derive packaging lines is the absence of MLV-related endogenous retroviral-like sequences (like VL30 in 3T3 cells) that can cross-package with MLV-derived retroviral vectors (Torrent et al., 1994) and generate potentially harmful recombinant retroviruses.

The helper constructs were introduced into other cell lines (HT1080 (table 2) Mink Mv-1-Lu (table 2), 3T3 (not shown), TE671 (table 2)) for the purpose of comparisons of the efficiency of the constructs.

5 As illustrated hereinafter (Table 2), the reverse transcriptase (RT) activity (provided by expression of the pol gene) in cells transfected with CeB is significantly higher than that of the same cells transfected by the parental plasmid pCRIP or that of cells chronically infected by MLV. This enhancement of viral gene expression is correlated with the titers of lacZ retroviral vectors when an envelope is provided in CeB-lacZ cells after comparison with titers of lacZ pseudotypes of either replication-competent viruses or other helper-free packaging systems.

10 For the generation of final packaging cell lines, the best clonal env transfectants have been selected. Packaging systems obtained in this way will be able to produce helper-free retroviral vectors at titers greater than 10^8 infectious particles per ml, which would be 10-100 fold higher to helper-free preparations of others.

25 Because of the way the selectable markers are expressed (see above), growing the packaging cells in phleomycin and blasticidin selective pressure increase and stabilize the expression of the retroviral components and particularly the envelopes, as it is possible that env glycoproteins have toxic effects for the producer cells in the long term which may lead to a decrease of expression.

30 Such an enhancement of viral production observed with the packaging systems described herein might increase the emergence of unwanted retroviruses having recombined between the genomes of both the retroviral vector and either of the

two packaging-deficient constructs. However, the constructs have been designed in such a way that it reduces the probability of emergence of recombinant viruses compared to the parental constructs. To check their safety, attempts
5 have been made to detect the presence of replication-competent retroviruses by a mobilisation assay of a lacZ provirus. No RC viruses have been found in all retroviral vector preparations tested so far.

10 The following Examples illustrate the invention.

Example 1

Preparation of Cell lines and viruses.

15 The following cell lines were used:

A204 (ATCC HTB 82), HeLa (ATCC CCL2), HT1080 (ATCC CCL121), MRC5 (ATCC CCL171), T24 (ATCC HTB 4), VERO (ATCC CCL81) and D17 (ATCC CCL183) were purchased from ATCC.

20 HOS, TE671 and Mv-1-Lu cells and their clones harboring MFGnls lacZ retroviral vector as described by Takeuchi et al., J Virol (1994), 68, 8001-8007.

25 The above cell lines were grown in DMEM (Gibco-BRL, U.K.) supplemented with 10% fetal calf serum.

EB8 (Battini et al., J. Virol (1992) 66: 1468-1475);
psiCRE, psiCRELLZ and psiCRIP (Danos et al., Proc. Natl.
Acad. Sci USA (1988) 85: 6460-6464);
30 Cells GP+EAM12 (Markowitz et al., Virology (1988), 167, 400-406); and
NIH-3T3 murine fibroblasts.

These cell lines were grown in DMEM (GIBCO-BRL, U.K.)
35 supplemented with 10% new-born calf serum.

Mv-1-Lu, TE671 and HT1080 cells were transfected using calcium-phosphate precipitation method (Sambrook et al., "Molecular Cloning" 1989, Cold Spring Harbour Laboratory Press: New York) as described elsewhere (Battini et al., supra). CeB-transfected Mv-1-Lu, TE671 and HT1080 cells were selected with 3, 6-8 and 4 µg/ml of blasticidin S (ICN, UK), respectively, and blasticidin-resistant colonies were isolated 2-3 weeks later. Cells transfected with the various env-expression plasmids were selected with phleomycin (CAYLA, France): 50 µg/ml (for FBASALF-transfected cells) or 10 µg/ml (for FBASAF-, FbdelPASAF-, FbdelPMOSAF, FbdelPIOAISAF or FbdelPRDSAF-transfected cells). Phleomycin-resistant colonies were isolated 2-3 weeks later.

Production of lacZ pseudotypes using replication competent viruses, amphotropic murine leukemia virus (MLV-A) 1504 strain and cat endogenous virus RD114, was carried out as described previously (Takeuchi et al., J Virol (1994), 68, 8001-8007).

Example 2

Preparation of Plasmids.

The env gene of pCRIP (Danos et al., supra) was excised by HpaI/ClaI digestion. A 500 bp PCR-generated DNA fragment was obtained using pSV2-bsr (Izumi et al., Experimental Cell Research (1991), 197, 299-233) as template and a pair of oligonucleotides:

(5' > CGGAATTCGGATCCGAGCTCGGCCAGCCGGCCACCATGAAAACATTTAACATTTTC TC) (SEQ ID NO 2) at 5' end and (5' > GATCCATCGATAAGCTTGGTGGTAAACTTTT) (SEQ ID No 3) at 3' end, with SfiI and ClaI sites, respectively. This fragment was inserted in HpaI/ClaI sites of pCRIP by co-ligation with a 85 bp HpaI/SfiI DNA fragment isolated from pOXEnv (Russell et al., Nucleic Acids Research (1993), 21, 1081-1085) which

provides the end of the Moloney murine leukemia virus (MoMLV) pol gene. The resulting plasmid named CeB (Fig. 1) could express the MoMLV gag-pol gene as well as the bsr selectable marker conferring resistance to blasticidin S, both driven by the MoMLV 5'LTR promoter.

A series of env-expression plasmids was generated using the 4070A MLV (amphotropic) env gene (Ott et al., J Virol (1990), 64, 757-766) and the FB29 Friend MLV promoter (Perryman et al., Nucleic Acid Res (1991), 19, 6950). In FBASALF (Fig. 1) a BglII/ClaI fragment containing the env gene was cloned in BamHI/ClaI sites of plasmid FB3LPh which also contained the C57 Friend MLV LTR driving the expression of the phleo selection marker. A 136 bp env fragment was generated by PCR using plasmid FB3 (Heard et al., J Virol (1991), 65, 4026-4032) as template and a pair of oligonucleotides: (5'>GCTCTTCGGACCCTGCATTC) (SEQ ID NO 4) at 5' end (before ClaI site) and (5'>TAGCATGGCGCCCTATGGCTCGTACTCTATAGGC) (SEQ ID NO 5) at 3' end, providing a Kasi restriction site immediately after the env stop codon. This PCR fragment was digested using ClaI and Kasi. A DNA fragment containing the FB29 LTR and the MLV-A env gene was obtained by NdeI/ClaI digestion of FBASALF. The fragments were co-ligated in NdeI/Kasi digested pUT626 (kindly provided by Daniel Drocourt, CAYLA labs, France). In the resulting plasmid, named FBASAF (Fig. 1), the phleo selectable marker was expressed from the same mRNA as the env gene. A BglII restriction site was created after the MscI site at position 214 in the FB29 leader by using a commercial linker (Biolabs, France). A NdeI/BglII fragment containing the FB29 LTR was co-inserted with the BglII/ClaI env fragment in NdeI/ClaI-digested FBASAF plasmid DNA, resulting in plasmid FBdelPASAF (Fig. 1). Compared to FBASAF, FBdelPASAF has a 100bp larger deletion in the leader region.

Example 3

Cloning and Sequencing of the RD114 env gene

The RD114 env gene was first sub-cloned in plasmid
5 Bluescript KS+ (Stratagene) as a 3 Kb HindIII insert
isolated from SC3C, an RD114 infectious DNA clone (Reeves et
al., J. Virol (1984), 52, 164-171). A 2.7 kb Scal-Hind III
fragment of this subclone containing the RD114 env gene was
sequenced (Figure 4 (SEQ ID NO 1) - EMBL accession number;
10 X87829). The 5' non-coding sequence upstream of an NdeI site
was deleted by an EcoRI/NdeI digestion followed by filling-
in with Klenow enzyme and self-ligation. From this plasmid,
two DNA fragments were obtained: a BamHI/NcoI 2.5 Kb
fragment and a 63 bp PCR-generated DNA fragment using
15 (5'>CGCCTCATGGCCTTCATTAA) (SEQ ID NO 6) at 5' end (before
NotI site) and (5'>TAGCATGGCGCCTCAATCCTGAGCTTCTTCC) (SEQ ID
NO 7) at 3' end, providing a KsI restriction site just
after RD114 env gene stop codon. The PCR fragment was
digested with NcoI and KsI. Both fragments were co-
20 inserted between BglII and KsI sites of FBdelPASAF and the
resulting plasmid was named FBdelPRDSAF (Fig. 1).

Plasmid pCRIPAMgag- (Danos, O. et al., Proc Natl Acad
Sci USA (1988) 85:6460-6464) was used for transfection.

25 Example 4

Infection assays.

Target cells were seeded in 24-multiwell plates (4x10⁴ cells
per well) and were incubated overnight. Infections were then
30 carried out at 37°C by plating 1 ml dilutions of viral
supernatants in the presence of 4 µg/ml polybrene (Sigma) on
target cells. 3h later virus-containing medium was replaced
by fresh medium and infected cells were incubated for two
days before X-gal staining, performed as previously
35 described (Tailor et al., J Virol (1993), 67, 6737-6741,

Takeuchi et al., J Virol (1994), 68, 8001-8007). Viral titers were determined by counting lacZ-positive colonies as previously described (Cosset et al., J. Virol. (1990) 64: 1070-1078). Stability of lacZ pseudotypes in fresh human serum was examined by titrating surviving virus after incubation in 1:1 mixture of virus harvest in serum-free medium and fresh human serum for 1 h at 37°C as described before (Takeuchi et al. supra).

10 Example 5

Reverse transcriptase (RT) assay.

RT assays were performed either as described previously (Takeuchi et al. supra) or using an RT assay kit (Boehringer Mannheim, U.K.) following the manufacturer's instruction but using MnCl₂ (2 mM) instead of MgCl₂.

Example 6

20 Screening producer cell lines.

Viral particles generated with RD114 envelopes have been found to be more stable in human serum than virions with MLV-A envelopes and that the producer cell line also controls sensitivity (Takeuchi et al. supra). A panel of cell lines was screened for their ability to produce high titer viruses and for the sensitivity of these virions to human serum. To do this, cells were infected at high multiplicity with lacZ pseudotypes of either MLV-A or RD114 and cells producing helper-positive lacZ pseudotypes were established. Human HT1080 and TE671 and mink Mv-1-Lu cells were found to release high titer lacZ(RD114) and lacZ(MLV-A) viruses. LacZ(MLV-A) pseudotypes produced by HT1080 cells were more resistant to human serum than those produced by other cells. The titer of these viruses was only four-fold less following a 1 hr incubation with human serum than a

control incubation (Table 1). LacZ(RD114) pseudotypes produced by human cells or mink Mv-1-Lu cells were in general stable in human serum (Table 1). These results suggested that HT1080, TE671 and Mv-1-Lu cells provided the best combination of high lacZ titers and resistance to human serum and they were therefore used for the generation of retroviral packaging cells.

Table 1. Titer and stability of lacZ pseudotypes.

Producer cell	LacZ (MLV-A)		LacZ (RD114)	
	Titer ^a	Stability ^b	Titer ^a	Stability ^b
A204	650	<3	1,200	105
HeLa	9	nd	2,000	115
HOS	4,500	6	23,000	86
HT1080	2,000,000	26	400,000	129
MRC-5	450	10	1,000	nd
T24	350	nd	1,200	nd
TE671	15,000	2	90,000	38
VERO	260	nd	90	nd
D17	900	<1	200,000	1
Mv-1-Lu	80,000	1	200,000	120

a: titration on TE671 cells as lacZ i.u./ml

b: % of infectivity of human serum-treated viruses compared to fetal calf serum-treated viruses

Example 7

Construction of an improved gag-pol expression vector.

A MoMLV gag-pol expression plasmid, CeB (Fig. 1), was

derived from pCRIP (Danos et al., Proc. Natl. Acad. Sci. USA
 (1988) 85: 6460-6464). Approximately 2 Kb of env sequence
 were removed from pCRIP and the bsr selectable marker,
 conferring resistance to blasticidin S (Izumi et al.,
 5 Experimental Cell Research (1991) 197:229-233), was inserted
 74 nts downstream of the gag-pol gene. This 74 nts interval
 had no ATG triplets and was thought to provide an optimal
 distance between the stop codon of the pol reading frame and
 the start codon of the bsr gene to allow re-initiation of
 10 translation (Kozak Mol Cell Biol., 1987, 7: 3438-3445).
 There was no "Kozak" consensus sequence (Kozak Cell, (1986)
 44: 283-292) at the 5' end of the marker gene. Therefore,
 bsr could only be expressed by re-initiation of translation
 after the upstream gag-pol gene had been expressed.
 15 Consequently, after transfection of CeB in Mv-1-
 Lu/MFGnlsLacZ (ML), TE671/MFGnlsLacZ (TEL) or HT1080 cells,
 blasticidin S-resistant bulk populations and most cell
 clones expressed high levels of gag-pol proteins assessed by
 the reverse-transcriptase (RT) activity found in cell
 20 supernatants (Table 2). Considerably higher RT activities
 were found in bulk populations of CeB-transfected ML cells
 compared to bulk population of ML cells stably transfected
 with the parental pCRIP construct. Similarly the RT
 activities of two packaging cell lines generated using
 25 pCRIPenv- construct, psiCRE cells (Danos et al., supra) and
 EB8 cells (Battini supra.) were less than that of CeB
 transfected clones (Table 2). Finally, RT activity in CeB
 transfected cell supernatants was higher than that of cells
 30 chronically infected by replication-competent MLV-A (Table
 2).

Table 2. Secreted reverse transcriptase expression

Cell ^a	RT activity ^b	LacZ Titer ^c
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	ML/MLV-A	1	8×10^4
	MLSvB	0.1	<1
	MLCRIP (bulk)	0.15	nd
	MLCeB (bulk)	1.7	nd
5	MLCeB1	4.2	1×10^6
	MLCeB4	1.6	1×10^6
	TEL/MLV-A	3.6	2×10^6
	TELCeB6	5.2	4×10^7
	HT1080/MLV-A	1.1	1×10^6
10	HTCeB6	1.9	1×10^6
	HTCeB18	2.7	2×10^6
	HTCeB22 (FLY)	6.9	5×10^6
	HTCeB48	5.5	3×10^6
	EB8	0.22	1×10^4
15	psiCRE-LLZ	1.2	1×10^{5d}

a: ML, Mv-1-Lu cells harboring a MFGnslacZ provirus; TEL, TE671 cells harboring a MFGnslacZ provirus; /MLV-A, cells chronically infected with MLV-A 1504 strain; MLSvB, ML cells transfected with a plasmid pSV2bsr alone; MLCRIP, ML cells co-transfected with pCRIP and pSV2bsr.

b: Average of arbitrary units relative to ML/MLV-A RT activity of at least two independent experiments was shown. The standard errors did not exceed 20 % of the values.

c: titration on TE671 cells as lacZ i.u./ml. After polyclonal transfection of a plasmid which expresses MLV-A env in MLCeB clones, TELCeB clones, HTCeB clones and EB8 cells; nd, not done.

d: titration on NIH3T3 cells

To rescue infectious lacZ viruses, MLCeB and TELCeB clones were transfected with FBASALF DNA, a plasmid designed to express the MLV-A env gene (Fig. 1). Bulk populations of stable FBASALF transfectants were isolated and supernatants were titrated using TE671 cells as targets. Titers of lacZ viruses were higher than either MLV-A infected ML or TEL cells, or FBASALF-transfected EB8 cells (Table 2). These data suggested that CeB was an extremely efficient MLV gag-pol expression vector in mink Mv-1-Lu and TE671 cells. CeB

was therefore used to derive packaging cells by transfection of HT1080 cells. 41/49 blasticidin S-resistant colonies had detectable levels of RT; 9 had RT activity higher than that of control MLV-A-infected HT1080 cells (data not shown).

5 Expression of gag precursor was confirmed in cell lysates and supernatants of these 9 HTCeB clones by immunoblotting using antibodies against p30-CA (data not shown). The 4 clones with the highest expression of gag proteins (clones 6,18,22 and 48) were infected at high-multiplicity with
10 helper free, lacZ pseudotypes bearing MLV-A envelopes (MFGnslacZ(A)) produced by TELCeB6/FBASALF (Table 3) and then transfected with FBASALF. Supernatants of bulk, phleomycin-resistant transfectants were assessed for RT activity and lacZ titer (Table 2). Clone HTCeB22, named FLY,
15 was found to be the best gag-pol producer clone and was used to introduce env expression vectors for the generation of packaging cell lines.

Table 3. Titer following env construct transfection

	Producer cell	Env source	Titer ^a
5	psiCRIP lacZ 5	pCRIPAMgag-	6x10 ^{4b}
	GP+EAM12 lacZ 25	envAM	3x10 ^{5b}
10	TELCeB6	FBASALF ^c	5x10 ⁷
		FBASAF ^c	2x10 ⁷
		FbdelPASAF ^c	2x10 ⁷
15	TELCeB6	FBdelPASAF 1	3x10 ⁷
		FbdelPASAF 4	2x10 ⁷
		FbdelPASAF 6	1x10 ⁷
		FbdelPASAF 7	5x10 ⁷
		FbdelPASAF 8	1x10 ⁷
20		FbdelPRDSAF 2	1x10 ⁶
		FbdelPRDSAF 4	3x10 ⁵
		FbdelPRDSAF 7	1x10 ⁷
		FbdelPRDSAF 8	2x10 ⁶
25	FLY ^d	FBdelPASAF 1	1x10 ⁴
		FbdelPASAF 4	1.5x10 ⁶
		FbdelPASAF 5	1x10 ⁶
		FbdelPASAF 7	1x10 ⁶
		FbdelPASAF 13	7x10 ⁶
		FbdelPASAF 14	4x10 ⁶
30		FbdelPASAF 15	1x10 ⁶
		FbdelPASAF 16	5x10 ⁶
		FbdelPASAF 17	6x10 ⁶
35	FLYA4 lacZ 3	FBdelPASAF 4	2x10 ^{7b}
	FLY ^d	FBdelPRDSAF 1	2.5x10 ⁶
		FbdelPRDSAF 2	1x10 ⁷
		FbdelPRDSAF 6	5x10 ⁶
		FbdelPRDSAF 10	2x10 ⁶
40		FbdelPRDSAF 11	3x10 ⁶
		FbdelPRDSAF 13	1x10 ⁶
		FbdelPRDSAF 17	5x10 ⁶
		FbdelPRDSAF 18	3x10 ⁷
45		FbdelPRDSAF 19	6x10 ⁶

Average titers of at least three independent experiments were shown. The standard errors did not exceed 30 % of the titer values.

50 a: titrated on TE671 cells as lacZ i.u./ml

b: results of best MFGnslacZ producer clones.

c: bulk populations of env-transfectants in TELCeB6 cells.
d: titration after bulk infection with helper-free MFGnslacZ.

5 Example 8

Construction of env expression vectors.

A series of MLV-A env expression plasmids were then
generated (Fig. 1). In FBASALF, the env gene was inserted
10 between two Friend-MLV LTRs, its expression driven by the
FB29 MLV LTR (Perryman et al., supra). Most of the packaging
signal located in the leader region was deleted. This
plasmid also expressed the phleo selectable marker (Gatignol
et al., supra) driven by the 3' LTR. FBASAF and FBdelPASAF
15 were then designed following the same strategy used for CeB.
These two vectors differed only by the extent of deletion of
the packaging signal, FBdelPASAF having virtually no leader
sequence. Compared to pCRIPAMgag- and pCRIPgag-2 env
plasmids expressed in psiCRIP or psiCRE packaging cells
20 (Danos et al., supra) about 5 Kb of gag-pol sequences was
removed. In addition the 258 bp retroviral sequence
containing the end of env gene and the beginning of U3 found
in pCRIPAMgag- and pCRIPgag-2 was also removed. For both
FBASAF and FBdelPASAF plasmids, the phleo selectable marker
25 was inserted downstream of the env gene by positioning a 76
nts linker with no ATG codons between the two open-reading
frames. Phleo could therefore only be expressed by re-
initiation of translation by the same ribosomal unit that
had expressed the upstream env open reading frame.
30 FBdelPASAF was also used to generate FBdelPRDSAF, an RD114
envelope expression plasmid (Fig. 1).

After transfection of the env plasmids into TELCeB6 cells
(Table 2), bulk populations of phleomycin-resistant colonies
35 were isolated and their production of lacZ virus measured

(Table 3). FBASALF gave a titer of 5×10^7 lacZ-i.u./ml, whilst titers with either FBASAF or FBdelPASAF were 2×10^7 lacZ-i.u./ml (Table 3). Titers of 5×10^7 or 10^7 lacZ-i.u./ml could be obtained with some FBdelPASAF cell clones or FBdelPRDSAF clones, respectively.

As FBdelPASAF has minimal virus-derived sequences and was shown to be the safest construct (see below and Table 4), it and FBdelPRDSAF were used to generate packaging lines from FLY cells (clone HTCeB22, Table 2). Envelope expression of these clones was assayed by interference to challenge with MFGnslacZ(A) or MFGnslacZ(RD) pseudotypes produced by TELCeB6/FBdelPASAF-7 or TELCeB6/FBdelPRDSAF-7, respectively (Table 3). The cell lines showing most interference were cross-infected at high multiplicity with these pseudotypes to provide MFGnslacZ proviruses, and supernatants were then titrated on TE671 cells (Table 3). FLY-FBdelPASAF-13 (FLYA13 packaging line) and FLY-FBdelPRDSAF-18 (FLYRD18 packaging line) gave the highest productions of lacZ viruses, around 10^7 lacZ-i.u./ml. The best MFGnslacZ producer clones derived from either psiCRIP cells (Danos et al., supra) or GP+EAM12 cells (Markowitz et al., supra) gave approximately 50 fold lower titers (Table 3). The lacZ titers of the FLY-derived lines shown in Table 3 are lower than the best TELCeB6-derived lines after transfection of either FBdelPASAF or FBdelPRDSAF (Table 3). However it should be noted that the lacZ provirus expressed in TELCeB6 cells was obtained after clonal selection but was introduced polyclonally in FLY-derived env-transfected cell clones. When FLY-FBdelPASAF-4 cells (FLYA4 packaging line), infected with helper-free MFGnslacZ(RD), were cloned by limiting dilution the best clones (eg. FLYA4lacZ3) were found to produce 20 times more infectious viruses than the bulk population, reaching the range of titers obtained with the best TELCeB6-FBdelPASAF clones (Table 3).

Example 9

Assays for transfer of gag-pol or env functions.

To assay for replication-competent viruses, supernatants
5 were used to infect TEL cells (a clone of TE671 cells
harboring an MFGnslacZ provirus). Infected cells were
passaged for 6 days or longer and their supernatants were
used for infection of fresh TE671 cells. No transmission of
lacZ viruses could be detected (Table 4), demonstrating that
10 the supernatants of pCRIPAMgag--, FBASALF-, FBASAF-, or
FBdelpASAF-transfected TELCeB6 cells were helper-free.
Similar absence of replication competent recombinant
retroviruses was demonstrated using supernatant from a
clone of psiCRIP-MFGnslacZ cells or from two clones of
15 FLYA-MFGnslacZ cells (Table 4).

There have been reports that helper-free retroviral vector
stocks may nevertheless contain recombinant retroviruses
(replication incompetent) carrying either gag-pol or env
20 genes (Bestwick et al., Proc Natl Acad Sci USA (1988), 85,
5404-5408, Cosset et al., Virology (1993), 193, 385-395,
Girod et al., Virology (1995), in press). To assay for such
recombinant retroviruses, mobilisation of an MFGnslacZ
provirus from two indicator cell lines which could cross-
25 complement potential recombinant viruses carrying either
gag-pol or env functional genes was attempted. The TELCeB6
line (Table 2) expressing gag-pol proteins was used as
indicator cell line to test for the presence of env
recombinant (ER) viruses. The TELMOSAF indicator line
30 expressing MoMLV env glycoproteins (obtained by transfection
of FBMOSAF, a plasmid expressing the MoMLV env gene using
FBASAF backbone, in TEL cells) was used to detect the
presence of gag-pol recombinant retroviruses (GPR viruses).
After passaging 4-8 days, the supernatants of the infected
35 indicator cells were used to infect either human TE671 cells

or murine NIH3T3 cells.

TELCeB6 cells transfected with various env-expressing constructs, pCRIPAMgag-, FBASAF and FBdelPASAF were compared. Although the supernatants of TELCeB6-FBdelPASAF cells were devoid of replication-competent retroviruses, they were found sporadically to transfer gag-pol genomes (Table 4). No GPR viruses could be detected when less than 2×10^5 virions were used to infect the indicator cells. Similarly TELCeB6 indicator cells infected with various helper-free viruses were shown sporadically to release lacZ virions (Table 4). The number depended both on the env-expression vector used and on the virus input quantity. Compared to lacZ viruses generated using pCRIPAMgag-plasmid, the frequency of detection of the env-recombinant viruses was lower for supernatants generated by using FBASAF and FBdelPASAF constructs (Table 4). For FBdelPASAF construct when less than 5×10^5 MFGnslacZ(A) helper-free virions were used to infect the indicator cells, no ER retroviruses could be detected. From these experiments, it could be estimated that a supernatant, produced from TELCeB6-FBdelPASAF cells, containing 1×10^7 infectious units of MFGnslacZ retroviral vector contained no replication-competent virus, and about 100 gag-pol and 100 env recombinant retroviruses.

Table 4. Transfer of packaging function

Producer cell		Indicator cell	Input virus ^a (lacZ-i.u.)	Detection ^b		
				++	+	-
5	Replication competent virus					
	psiCRIP lacZ 5	TEL	2x10 ⁴	0/4	0/4	4/4
	TELCeB6-pCRIPAMgag-	TEL	5x10 ⁶	0/4	0/4	4/4
10	TELCeB6-FBASAF	TEL	5x10 ⁶	0/4	0/4	4/4
	TELCeB6-FBdelPASAF	TEL	5x10 ⁶	0/4	0/4	4/4
15	FLYA4 lacZ 3	TEL	1x10 ⁷	0/4	0/4	4/4
	FLYA4 lacZ 7	TEL	1x10 ⁷	0/4	0/4	4/4
Gag-pol recombinant						
	TELCeB6-FBdelPASAF 7	TELMOSAF	2x10 ⁷	0/4	1/4	3/4
20	TELCeB6-FBdelPASAF 7	TELMOSAF	2x10 ⁶	0/4	2/4	2/4
	TELCeB6-FBdelPASAF 7	TELMOSAF	2x10 ⁵	0/4	2/4	2/4
	TELCeB6-FBdelPASAF 7	TELMOSAF	2x10 ⁴	0/4	0/4	4/4
Env recombinant						
25	TELCeB6-pCRIPAMgag-	TELCeB6	5x10 ⁶	2/4	1/4	1/4
	TELCeB6-pCRIPAMgag-	TELCeB6	5x10 ⁵	1/4	1/4	2/4
	TELCeB6-pCRIPAMgag-	TELCeB6	5x10 ⁴	0/4	2/4	2/4
	TELCeB6-FBASAF	TELCeB6	5x10 ⁶	0/4	2/4	2/4
30	TELCeB6-FBASAF	TELCeB6	5x10 ⁵	0/4	1/4	3/4
	TELCeB6-FBASAF	TELCeB6	5x10 ⁴	0/4	1/4	3/4
	TELCeB6-FBdelPASAF	TELCeB6	5x10 ⁶	0/4	1/4	3/4
	TELCeB6-FBdelPASAF	TELCeB6	5x10 ⁵	1/4	3/4	0/4
35	TELCeB6-FBdelPASAF	TELCeB6	5x10 ⁴	0/4	0/4	4/4

a: number of lacZ i.u. used to infect indicator cells

b: number of incidence out of four experiments. The ranges of lacZ titers rescued from infected indicator cells are shown for each virus input: >100 lacZ i.u./ml (++) , 1-100 lacZ i.u./ml (+) and <1 lacZ i.u./ml (-).

Titers were determined on TE671 cells for replication competent virus and env recombinant and NIH3T3 cells for

gag-pol recombinant.

Example 10

In order to confirm resistance to complement and absence of replication competent virus in our best packaging lines, MFGnslacZ(A) and (RD) harvested from FLYA13 and FLYRD18, respectively, after polyclonal transduction of MFGnslacZ (Table 3 above) were tested for stability in fresh human serum and generation of replication competent virus. Titers of MFGnslacZ(RD) from FLYRD18 after 1 hr incubation with 3 independent samples of fresh human serum were 80 to 120 % of control incubations, while titers of MFGnslacZ(A) from FLYA13 were 50 to 90 % of controls (data not shown). No replication competent virus was detected in the same assay described above (Table 4) when 1×10^7 i.u. each of MFGnslacZ(A) and (RD) were tested.

EXAMPLE 11.

Generation of plasmids.
CeB plasmid (Fig. 5) expressing MoMLV gag-pol gene, was further modified to remove the splice donor site located in the leader region. A 272 bp fragment was PCR-generated by using OUSD- (5'-TCTCGCTTCTGTTCGCGCGC) and OLSD- (5'-TCGATCAAGCTTGCGGCCGCGGTGGTGGGTGGTGGTCC) as primers and further digested with BssHII and HindIII. A 1008 bp HindIII-XhoI fragment isolated from CeB (encompassing a part of leader sequence and beginning MoMLV gag) and the PCR fragment were co-inserted into pCeB from which the 1275 bp BssHII-XhoI fragment (encompassing R-U5-leader-gag) had been removed. The resulting plasmid, named pCeB DS- (Fig. 5), beared the deletion of splice donor (SD) site and a NotI restriction site created just downstream to the lost SD site.

A series of gag-pol expression plasmids in which the MoMLV LTR promoter was replaced by the human cytomegalovirus immediate early promoter (hCMV promoter) was derived from both CeB DS- and hCMV-G (Yee et al., 1994 PNAS, 91: 9564-9568), a plasmid used as a source for the hCMV promoter. A NotI-filled/EcoRI 7260 bp fragment was isolated from CeB DS- and cloned into hCMV-G which had been opened with SalI (further rendered blunt-ended) and EcoRI to remove the VSV-G gene. The resulting plasmid was cutted with ClaI and EcoRI to remove a 1155 bp fragment encompassing sequence derived from 3'-LTR and SV40 polyA sequence and self-ligated after filling both protruding DNA ends. The resulting plasmid, named phCMV-intron (Fig. 5), had gag-pol and bsr ORFs inserted between the CMV promoter and rabbit beta-globin polyA post-transcriptional regulatory sequences.

An intermediate plasmid was generated by sub-cloning a 7260 bp EcoRI fragment (isolated from CeB DS-) into hCMVG opened with EcoRI. A 1155 bp fragment (encompassing sequence derived from 3'-LTR and SV40 polyA sequence) was removed from this intermediate plasmid which was then re-circularized by self ligation after filling both ends. The resulting plasmid, named phCMV+intron 2P (Fig. 5), was digested with NotI and the vector was treated with klenow enzyme. A 1440 bp fragment (encompassing hCMV promoter and rabbit beta-1 globin intron B (Rohrbaugh et al., 1985 Mol. Cell Biol, 5: 147-160)) was isolated from phCMV+intron 2P by NotI/EcoRI digestion. This fragment was further treated with klenow enzyme and ligated back into the vector. The resulting plasmid, named hCMV+intron (Fig. 5), could express gag-pol and bsr genes driven by the hCMV promoter and beared an intron sequence derived from rabbit beta-1 globin intron B having both SD and SA (splice acceptable) sites.

A 2450 bp fragment was removed from phCMV+intron 2P by

NotI/XhoI digestion. The resulting vector fragment was then used to co-ligate a 1330 bp fragment (containing hCMV promoter + 5' end of rabbit beta-1 globin intron B (with SD site)) isolated from phCMVG by ApaI-filled/NotI digestion and a 1 kb fragment isolated from phCMV+intron 2P by NotI-filled/XhoI digestion. Compared to phCMV+intron 2P, the resulting plasmid, named hCMV+SD intron (Fig. 5), had the deletion of the 3' end of the rabbit beta-1 globin intron B and thus no SA site in the leader region.

Construct phCMV+leader (Fig. 5) has been described elsewhere (Savard et al., unpublished). This plasmid, in which gag-pol and bsr genes were driven by the hCMV promoter, had the MoMLV SD site in the leader region.

Gag-pol expression.

The different constructs, including the parental CeB plasmid, were analysed comparatively in a complementation assay after transfection in TEL-FBdelpASAF cells expressing 4070A-MLV (amphotropic) envelope and harboring a MFGnslacZ provirus. The transient production of lacZ retroviruses as well as the stable production of lacZ retroviral vectors after selection with blasticidin S were determined (Table 5). All the constructs were able to rescue infectious lacZ retroviruses indicating the expression of gag-pol proteins after transient transfection. Most likely due to the efficient hCMV and rabbit beta-1 globin intron B (post)-transcriptional regulatory sequences, hCMV+intron was particularly potent in transient retroviral vector production. However, 10 times less blasticidin-resistant colonies were obtained with hCMV+intron comparatively to CeB, and stable lacZ virus production from hCMV+intron was about 5-10 times lower than that of CeB. Clonal examination of lacZ retrovirus production from blasticidin-resistant colonies indicated that 80-90% of colonies could express

high levels of gag-pol proteins for both hCMV+intron and CeB plasmids. In contrast, despite variation in their ability to form blasticidin-resistant colonies after transfection and despite their ability to express gag-pol proteins from transient transfectants, all other constructs had a weak capacity for rescuing lacZ retroviral vectors from stable transfectants (Table 5).

Table 5. Comparative study of gag-pol-bsr plasmids.

gag-pol-bsr plasmid	Transient (lacZ i.u./ml)	no clones bsr ^r	Stable (lacZ i.u./ml)	% gag-pol /bsr
Ceb	300/ml	50	10 ⁷	90%
Ceb DS-	144/ml	5	10 ⁵	50%
hCMV+intron 2P	ND	20	10 ⁶	50%
hCMV-intron	812/ml	0	-	-
hCMV+SD intron	150/ml	1000	10 ²	nd
hCMV+leader	328/ml	1000	10 ² -10 ³	nd
hCMV+intron	12000/ml	5	10 ⁶ -10 ⁷	80%

Northern blot analyses were performed on stable transfectants (blasticidin-resistant) obtained with some of the gag-pol-bsr plasmids. As expected, the results (not shown) displayed a correlation between expression of gag-pol mRNAs and gag-pol protein expression detected by rescue analysis (Table 5). CeB construct was found to produce 2-3 fold more gag-pol mRNAs compared to hCMV+intron. Interestingly, an unexpected 2.45 kb RNA band was found for hCMV+intron construct at a ratio of 2:1 compared to the abundancy of the gag-pol mRNA band (at 5.95 kb). Further

investigations by using other probes revealed that a cryptic splice donor (SD) site located in the gag gene (right in the middle of the CA coding region at position 1596-1597 -numbering according to Shinnick et al., 1981 Nature (London) 293: 543-548) was activated in this latter construct. The 2.45 RNA species, lacking the 3' half of the gag gene and most of the pol gene, is unlikely to give rise to any useful translational product. It is therefore interesting to notice that hCMV+intron construct was able to give rise to slightly more transcripts (gag-pol 5.95 mRNA + 2.45 alternative RNA band) compared to gag-pol mRNA expressed from CeB construct. Therefore we decided to inactivate the cryptic SD site in the hCMV+intron construct in order to increase the ratio of gag-pol mRNAs.

Assays for transfer of gag-pol functions.

Although the supernatants of packaging cell lines generated with CeB gag-pol expression construct were devoid of replication-competent retroviruses, they were found sporadically to transfer gag-pol genomes (example 9, Table 4) (Cosset et al., 1995 J. Virol 69: 7430-7436). Because gag-pol-bsr constructs generated here by using the hCMV promoter had much less retroviral sequences homologous to the retroviral vector than the parental CeB construct (Fig. 5), they are less likely to give rise to gag-pol recombinant (GPR) viruses. Therefore, the most efficient gag-pol-bsr plasmids, hCMV+intron and CeB, were further analysed for emergence of GPR viruses. To assay for such recombinant retroviruses, we attempted to mobilise an lacZ provirus from an indicator cell lines which could cross-complement potential recombinant viruses carrying gag-pol functional genes. Results displayed in Table 6 showed that consistently with data reported previously (example 9, Table 4) (Cosset et al., 1995 Supra), lacZ retrovirus vectors generated by using CeB gag-pol construct were contaminated with GPR viruses. In

contrast lacZ retrovirus vectors generated by using hCMV+intron construct were completely devoid of such GPR viruses, suggesting that this construct was improved compared to CeB with respects with emergence of recombinant viruses.

Table 6. Comparative study of gag-pol-bsr plasmids.

plasmid	input virus (lacZ i.u.) ^a	no of experiments giving titres of ^b		
CeB	5x10 ⁶	5	3	0
	5x10 ⁵	2	4	2
	5x10 ⁴	0	1	7
hCMV+intron	5x10 ⁶	0	0	8
	5x10 ⁵	0	0	8
	5x10 ⁴	0	0	8

4x10E4 cells of TEL/MOSAF in 24 wells were challenged with lacZ(A) of i.u. indicated in the table (a), and incubated at 37°C for 3 days. Cells were trypsinized and transferred into small flasks. Cell sup was harvested on day 5 after lacZ(A) challenge and plated on either TE571 (not shown) and 3T3 cells (b). No lacZ was mobilized into TE671 at all. LacZ(A) from CMV-int 10 again did not rescue lacZ from TEL/MOSAF.

Example 12

Generic primers to detect D-type (Medstrand and Blomberg J.Virol. (1993) 67:6778-6787) , C-type (Shih et al., J Virol. (1989) 63:64-75), human endogenous virus RTVL-H (Wilkinson et al., J.Virol. (1993) 67:2981-2989), by RT-PCR were employed (Patience et al., supra). Primers to detect mouse endogenous VL30 element (Adams et al Mol.Cel.Biol. (1988) 8:2989-2998), and MFGn1slacZ RNA were designed and synthesized (TABLE X). Overnight supernatants (in 4ml of culture medium) from 106 cells of GP+EAM12lacZ25, FLYA4lacZ3

and TELCeB6FBASALF cells (Table 3) were harvested and centrifuged in sucrose gradient as described previously (Patience et al., J.Virol., 70:2654-2657). Fractions containing retrovirus particles were collected, and RNA extracted. One twentieth of the RNA preparation or dilution's thereof were applied to RT-PCR as described previously (Table X). A 1/200 of RNA harvested from GP+EAM12lacZ25 cells was positive for VL30 RNA. MFGnls lacZ RNA was found from 1/20 of RNA from GP+EAM12lacZ and TELCeB6FBASALF cells and 1/200 of RNA from FLYA4lacZ3 cells. The primer combinations for RTVL-H, C- and D-type RNA did not give detectable PCR product.

Table 7. RT-PCR detection of endogenous retrovirus RNA associated with virus particles.

RNA	primer (5'-3') forward(F)/reverse(R)	rt-pcr of virion associated RNA from ^a		
		GP+EAM12 lacZ25	FLYA4 lacZ3	TELCeB6F BASALF
MFGnls lacZ	F) CTCTGGCTCACAGTACGACGTAG R) CCATCAATCCGGTAGGTTTTCCG	+	++	+
C-type	F) CARRGKTTCAARAACWSYCCCAC R) AGYARVGTAGCNGGGTTHAGG	-	-	-
D-type	F) TCCCCTTGGAATACTCCTGTTTTYGT R) CATTCCTTGTGGTAAACTTTCCAYTG	-	-	-
RTVL-H	F) CCTCACCTGATCACRYTTG R) GAATTATGTCTGACAGAAGGG	NT	-	-
VL30	F) GTTGACATCTGCAGAGAAAGACC R) TCTGAGGTCTGTACACACAATGG	++	NT	NT

a: -, not detected; + detected in 1/20 RNA preparation; ++ detected in 1/200 RNA preparation; NT, not tested because the cells do not possess the corresponding genes.

5

EXAMPLE 13.**Generation of gag-pol pre-packaging cells by using TE671 cells.**

10 CeB, a plasmid designed to over-express MoMLV gag and pol proteins was introduced in TE671 human rhabdomyosarcoma cells (ATCC CRL8805). After selection with blasticidin, 50 bsr-positive colonies were isolated and the RT (reverse transcriptase) activity was analysed in their supernatants.

15 12 TE671-CeB (TECeB) clones with high RT activity were selected for further analysis. The best TECeB clone, clone #15, had a RT activity roughly equivalent to that TELCeB6 cells (Cosset et al., J. Virol. 69:7430-7436 (1995); see also Example 7, Table 6 in this patent application) but

20 displayed 2-3 fold more gag-precursors into cells as demonstrated in immunoblots by using anti-CA antibodies. The biological activity of gag-pol proteins expressed in the six best TECeB clones was further confirmed by their ability to produce infectious retroviruses in a complementation assay.

25 A lacZ provirus was introduced into each of the TECeB clones by polyclonal cross-infection by using lacZ(RD114) helper-free retrovirus vectors. FBMOSALF, a MoMLV env expression plasmid (Cosset et al., J. Virol. 69:6314-6322), was then transfected in each of the TECeB-lacZ lines and in the

30 TELCeB6 cell line for comparison. After selection with phleomycin, the titer of lacZ retrovirus vectors was determined in the supernatant of pools of phleomycin-resistant colonies for each TECeB-lacZ-FBMOSALF lines. A

good correlation was found between gag-pol expression into the TE-CeB clones (as determined by RT-assays and anti-gag immunoblots) and their ability to release infectious lacZ particles. TE-CeB15 cells could release approximately the same number of lacZ particles when compared to TELCeB6 cells although TELCeB6 cells had the advantage of being selected for lacZ expression (Cosset et al., J. Virol. 69:7430-7436 (1995)). TE-CeB15 cells were therefore used to derive retroviral packaging cell lines.

Construction of env-expression plasmids.

A series of plasmid (Fig. 3) was designed to allow expression of different retroviral envelope genes (isolated from MoMLV, GALV -Gibbon Ape Leukemia Virus-, and MLV-10A1). FBdelPMOSAF (Fig. 3, nucleotide sequence in Fig. 10) and FBdelP10A1SAF, expressing ecotropic MoMLV or MLV-10A1 envelopes, were generated by replacing the BglII/ClaI fragment from FBdelPASAF (Cosset et al., J. Virol. 69:7430-7436 (1995); see also Example 7, Fig. 2 and nucleotide sequence in Fig. 9) encompassing most of the env gene and splice acceptor site with that of MoMLV (position 5407 to 7679, Shinnik et al., 1981) or with that of MLV-10A1 (Ott et al., J. Virol. 64:757-766 (1990)).

Nucleotides 7514-7516 of GALV (Delassus et al., Virology 173:205-213 (1989)) were mutated by PCR-mediated mutagenesis to create a ClaI site (AAG to CGA), thereby introducing a conservative modification (a lysine (amino-acid 665 of GALV env precursor) to an arginine). The BamHI/ClaI fragment (nts 4994 (Delassus et al. Virology 173:205-213 (1989)) to 7517) was then sub-cloned into FBdelPASAF in which the BglII/ClaI encompassing most of the env gene and splice acceptor site had been removed. The resulting plasmid, expressing GALV

envelope glycoproteins, was named FBdelPGASAF (Fig. 3, nucleotide sequence in Fig. 11).

CMV10A1 was generated by inserting a Klenow enzyme-filled EagI/SalI fragment from FBdelP10A1SAF (encompassing 10A1 MLV env gene and phleo selectable marker) into hCMV-G digested with BamHI and filled with Klenow enzyme. The resulting plasmid, CMV10A1 (Fig. 3 and nucleotide sequence in Fig. 13) could express 10A1 envelopes under control of the hCMV promoter and the phleo selectable marker by translation re-initiation.

Generation of a multi-tropic set of TE671-based retroviral packaging lines.

FBdelPRDSAF (Fig. 3, nucleotide sequence in Fig. 12), FBdelPASAF, FBdelPGASAF, FBdelPMOSAF and FBdelP10A1SAF were independently introduced into cells of the TE-CeB15 pre-packaging line, expressing MoMLV gag-pol proteins.

Transfected cells were phleomycin-selected and 15-20 phleo-resistant colonies were isolated for each env-expression plasmid transfected.

Individual colonies were then analysed for expression of envelope glycoproteins by immunoblots on cell lysates by using antibodies against RD114 SU glycoproteins or against Rausher leukemia virus SU (to screen MoMLV, MLV-4070A and MLV-10A1 env-producer clones) or against GALV. The best env-producer colonies as determined in this assay were further analysed by a complementation assay after introducing a lacZ retroviral vector. LacZ pseudotypes released from the different packaging cell lines were titrated by using NIH 3T3 cells or TE671 cells as target. Titers higher than 1×10^7 lacZ i.u./ml were obtained for the best clones. Depending on the envelope specificities expressed in these cells, the new

TE671-based retroviral packaging cell lines were named TE-FLYE, TE-FLYA, TE-FLYRD, TE-FLY10A1, and TE-FLYGA and could express the MoMLV, MLV-4070A, RD114, MLV-10A1, and GALV env genes, respectively.

- 5 Assays for detecting replication-competent retroviruses (RCRs) were performed in the supernatants of these cells and were negative (less than 1/ml).

10 TE671 cells are very potent for transient expression resulting in more than 95% of cells expressing transgene three days after plasmid transfection (Hatzilioannou and Cosset, unpublished data, (1996)). The ability of retroviral packaging cell lines to transiently produce retroviral vectors is of crucial importance for gene therapy where
15 vectors carrying toxic gene have to be prepared. Transient expression of retroviral vectors was comparatively determined from cells of the TE-FLYA line and from the BING line (Pear et al., Proc Natl Acad Sci U S A 90, 8392-6 (1993)), a retroviral packaging cell line designed to
20 transiently express retroviral vectors. Results (Table 8) showed that TE-FLYA cells were more efficient for transient expression of a lacZ retroviral vector hence resulting in higher titers.

25 Table 8. Comparative study of transient production of lacZ vectors.

packaging cell line	cell number ^a	% transfected cells ^b	transient titer ^c
BING	281	5.3	2x10 ²
TE-FLYA	117	35	1.3x10 ³

30

Cells were transfected by MFGnslacZ retroviral vectors with calcium phosphate precipitation method and titers of of lacZ vectors (c) released in cell

supernatant were determined as lacZ i.u./ml at day 3 following transfection. The relative number of cells (a) (average per microscope field) and the % of transfected cells (b) determined after X-gal staining are shown.

5 Retroviral vectors prepared from TE671-based packaging cell lines were analysed for their sensitivity to human-complement mediated inactivation. Experiments were conducted as previously described (Cosset et al., J. Virol. 69:7430-7436 (1995); see also Example 10 in this patent application)

10 by using three human sera of individual donors (Table 9). As expected MLV-A prepared from mouse 3T3 cells were highly sensitive to inactivation after 1 hr incubation with sera. In contrast, titers of lacZ vectors produced from TE-FLYRD

15 lacZ vectors from TE-FLYA cells were 1 to 30% of controls.

Table 9. Human serum sensitivity of viruses produced from TE671-based packaging cell lines.

Virus from:	hu56 ^a	hu57 ^a	BTS ^a
3T3/A	<0.2, <0.2	<0.2, <0.2	<0.2, <0.2
TE-FLYE	15, 7.8	16, 11	48, 60
TE-FLYA	1, 0.6	2.2, 7.1	28, 19
TE-FLYRD	17, 22	30, 44	54, 63

Three human fresh serum samples were tested in duplicate; hu56 (A+), hu57 (AB+), BTS (AB+). (a) % control (average for FCS and opti-MEM treatment) is shown.

CLAIMS:

1. A recombinant expression vector comprising a gene of interest and a selectable marker gene, wherein the selectable marker gene is arranged downstream of the gene of interest and a stop codon associated with the gene of interest is spaced from a start codon of said selectable marker gene at a distance which is sufficient to ensure that said selectable marker protein is expressed from the corresponding mRNA as a result of translation reinitiation.
2. A recombinant expression vector according to claim 1 wherein the vector is a viral vector.
3. A recombinant expression vector according to claim 2 wherein the vector is a retroviral vector.
4. A recombinant expression vector according to any one of claims 1 to 3 wherein the gene of interest is included as part of a viral packaging construct.
5. A recombinant expression vector according to any one of the preceding claims wherein the number of nucleotides in the space between the stop codon of the gene of interest and the start codon of the selectable marker is in the range of from 20 to 200 nucleotides.
6. A recombinant expression vector according to claim 5 wherein the number of nucleotides in the space between the stop codon of the gene of interest and the start codon of the selectable marker is in the range of from 60 to 80 nucleotides.
7. A process for producing a cell line in which a gene of interest is expressed, which process comprises:
transforming host cells with an expression vector

according to any one of the claims 1 to 6; and
selectable those cells where expression of the
selection marker gene may be detected.

8. A process according to claim 7 wherein the host cell is a eukaryotic cell.
9. A host cell transformed with a recombinant expression vector according to any one of the claims 1 to 6.
10. A retroviral packaging cell line comprising a host cell transformed with a first and a second recombinant expression vector, said first recombinant expression vector having a packaging-deficient construct comprising a viral gag-pol gene and a first selectable marker gene downstream thereof, and said second recombinant expression vector having a packaging-deficient construct comprising a viral env gene and a second selectable marker gene downstream thereof; wherein the start codon of the first and second selectable markers are spaced from the stop codons of the viral gag-pol gene and the viral env gene respectively by a distance which ensures that said selectable marker protein is expressed from the corresponding mRNA as a result of translation reinitiation.
11. A retroviral packaging cell line according to claim 10 wherein the first selectable marker is a bsr selectable marker and the second selectable marker is a phleo selectable marker.
12. A retroviral packaging cell line according to any one of claims 10 or 11 wherein the packaging-deficient construct comprising the viral gag-pol gene and first selectable marker is the CeB (SEQ ID No 2) expression construct.

13. A retroviral packaging cell line according to any one of claims 10 or 11 wherein the packaging-deficient construct comprising the viral env gene and second selectable marker is the FbdelPASAF (SEQ ID No 5), the FbdelPMOSAF (SEQ ID No 6), the FbdelPGASAF (SEQ ID No 7), the FbdelPRDSAF (SEQ ID No 8), the FbdelPXSAF (Fig. 3), the FbdelP10A1SAF (Fig. 3), or the FbdelPVSVGSAF (Fig. 3) expression construct.
14. A retroviral packaging cell line according to any one of claims 10 or 11 wherein the recombinant expression vector is a packaging-deficient retroviral helper construct.
15. A retroviral packaging cell line according to claim 14 wherein the overlapping sequences between the genomes of the retroviral vector and the packaging-deficient construct is reduced by minimizing the extent of non-coding retroviral sequences in the packaging-deficient genome.
16. A retroviral packaging cell line according to any one of claims 10 to 15 wherein the viral gag-pol gene and the selectable marker are expressed under the control of a non-retroviral promoter.
17. A retroviral packaging cell line according to claim 16 wherein the promoter is fused to rabbit beta-1 globin intron.
18. A retroviral packaging cell line according to claim 16 or claim 17 wherein the promoter is a hCMV promoter.
19. A retroviral packaging cell line according to any one of claims 16 to claim 18 wherein the viral gag-pol gene and the selectable marker is a hCMV+intron (SEQ

- ID No3) or a hCMV+intronkaSD (SEQ ID No 4) expression construct.
20. A retroviral packaging cell line according to anyone of claims 10 to 15 wherein the viral env gene and the selectable marker are under the control of a non-retroviral promoter.
 21. A retroviral packaging cell line according to claim 20 wherein the promoter is fused to rabbit beta-1 globin intron.
 22. A retroviral packaging cell line according to claim 20 or claim 21 wherein the promoter is a hCMV promoter.
 23. A retroviral packaging cell line according any one of claims 20 to 22 wherein the viral env gene and the selectable marker is a CMV10A1 (SEQ ID No 9) expression construct.
 24. A retroviral packaging cell line according to any one of claims 10 to 23 wherein the cell line is the HT1080 line, the TE671 line, the 3T3 line, the 293 line or the MV-1-LU line.
 25. A retroviral packaging cell line according to anyone of claims 10 to 24 wherein the retroviral packaging cells comprises human HT1080 cells and express RD114 envelopes.
 26. A retroviral packaging cell line according to anyone of claims 10 to 24 wherein the retroviral packaging cells comprises human TE671 cells and express RD114 envelopes.

27. A process for producing a retroviral packaging cell line in which a gene of interest is expressed, which process comprises:

transforming host cells with a first and a second recombinant expression vector, said first recombinant expression vector having a packaging-deficient construct comprising a viral gag-pol gene and a first selectable marker gene downstream thereof, and said second recombinant expression vector having a packaging-deficient construct comprising a viral env gene and a second selectable marker gene downstream thereof; wherein the start codon of the first and second selectable markers are spaced from the stop codons of the viral gag-pol gene and the viral env gene respectively by a distance which ensures that said selectable marker protein is expressed from the corresponding mRNA as a result of translation reinitiation; and

selecting transformed cells which express said first and/or second marker genes.

28. A packaging deficient construct for use in a process according to claim 27, which expresses a viral gag-pol gene and a selectable marker wherein a start codon of the selectable marker is spaced from a stop codon of the viral gag-pol gene by a distance which ensures that said selectable marker protein is expressed from the corresponding mRNA as a result of translation reinitiation.
29. A packaging deficient construct for use in a process according to claim 27, which expresses a viral env gene and a selectable marker gene; wherein a start codon of the selectable marker is spaced from a stop codon of the viral env gene by a distance which ensures that said selectable marker protein is

expressed from the corresponding mRNA as a result of translation reinitiation.

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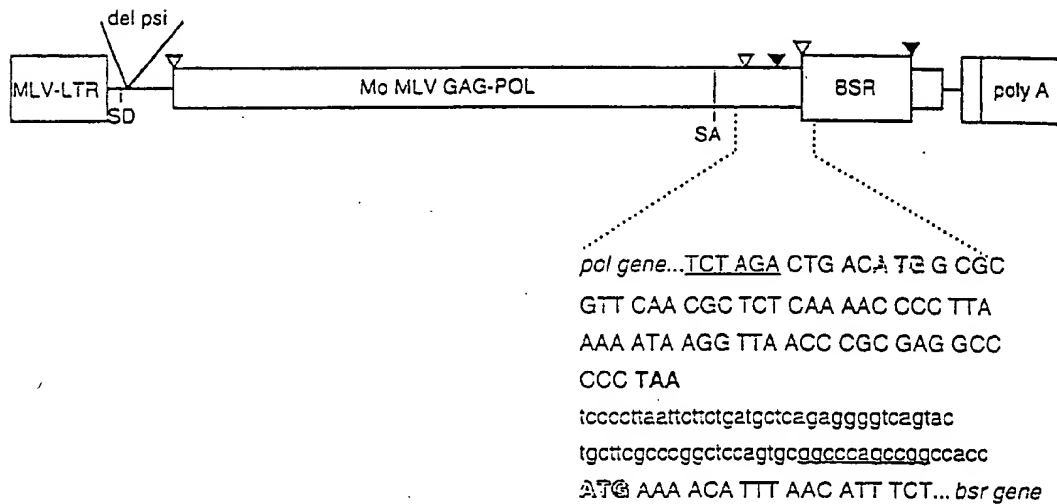


Figure 1. Schematic structure of CeB expression vector

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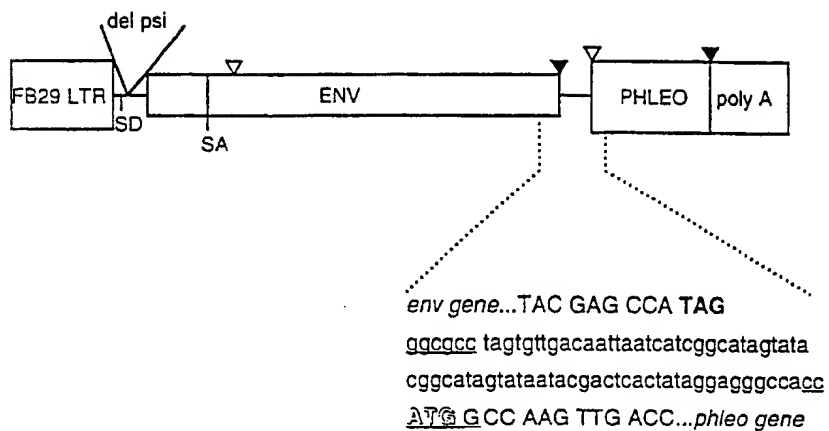


Figure 2. Schematic structure of FBdelPAS expression vector

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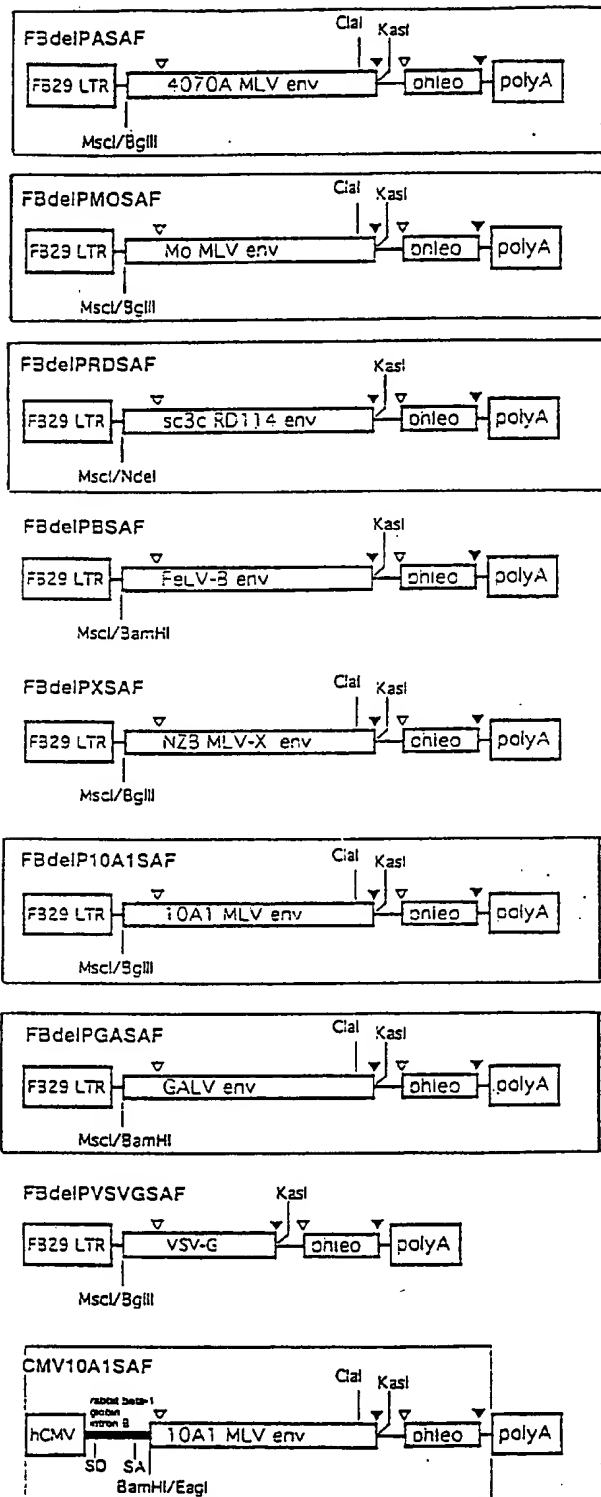


Figure 3. Schematic structure of env expression vectors

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 CTTTGTCAACCTTGCTCAATTCCTTCTCCCCCTCCGATCCTAAGACTGATTTACAAGCCC 240
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Fig. 4

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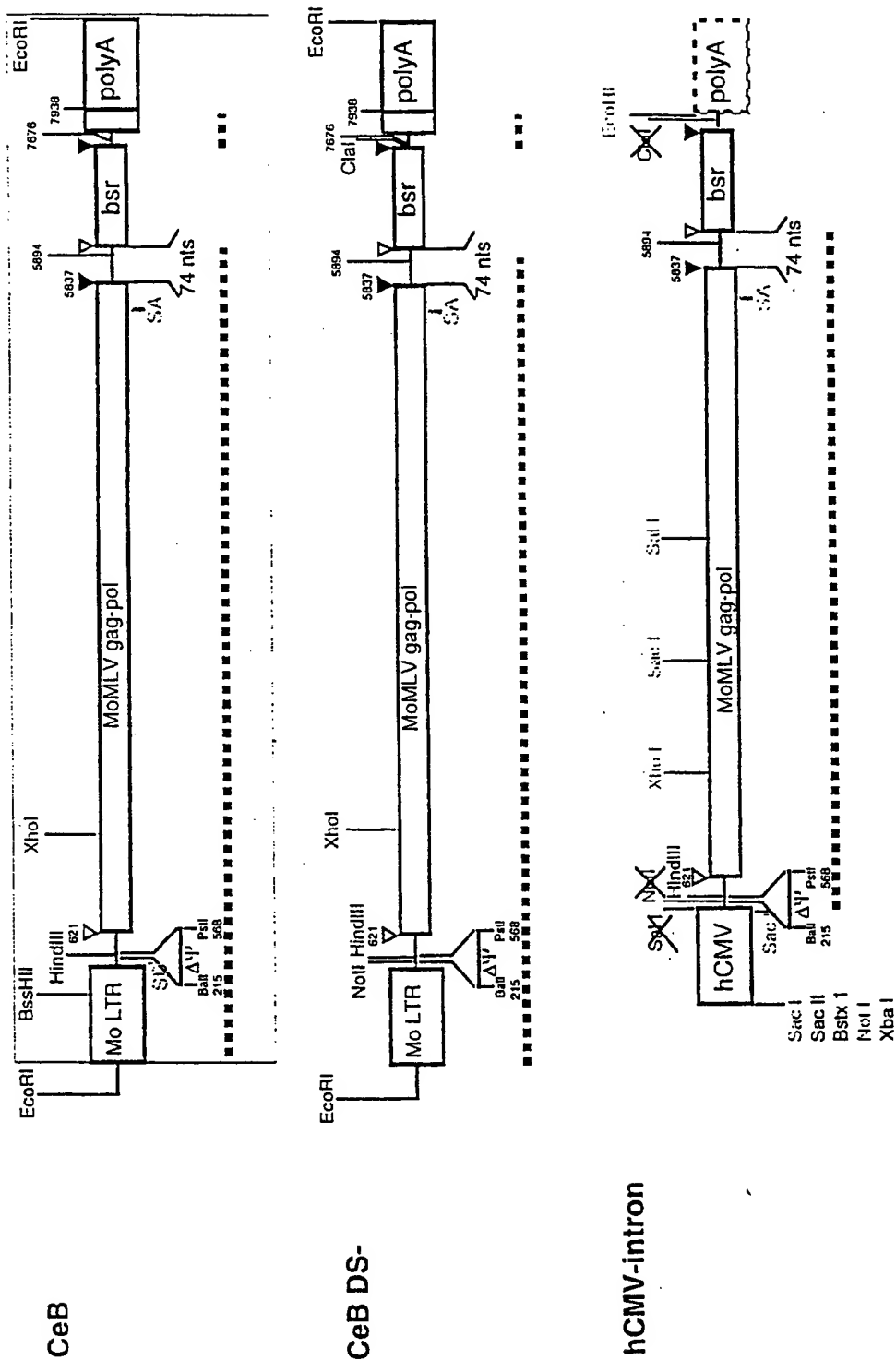


Figure 5. Genetic structure of gag-pol constructs (page 1/3)

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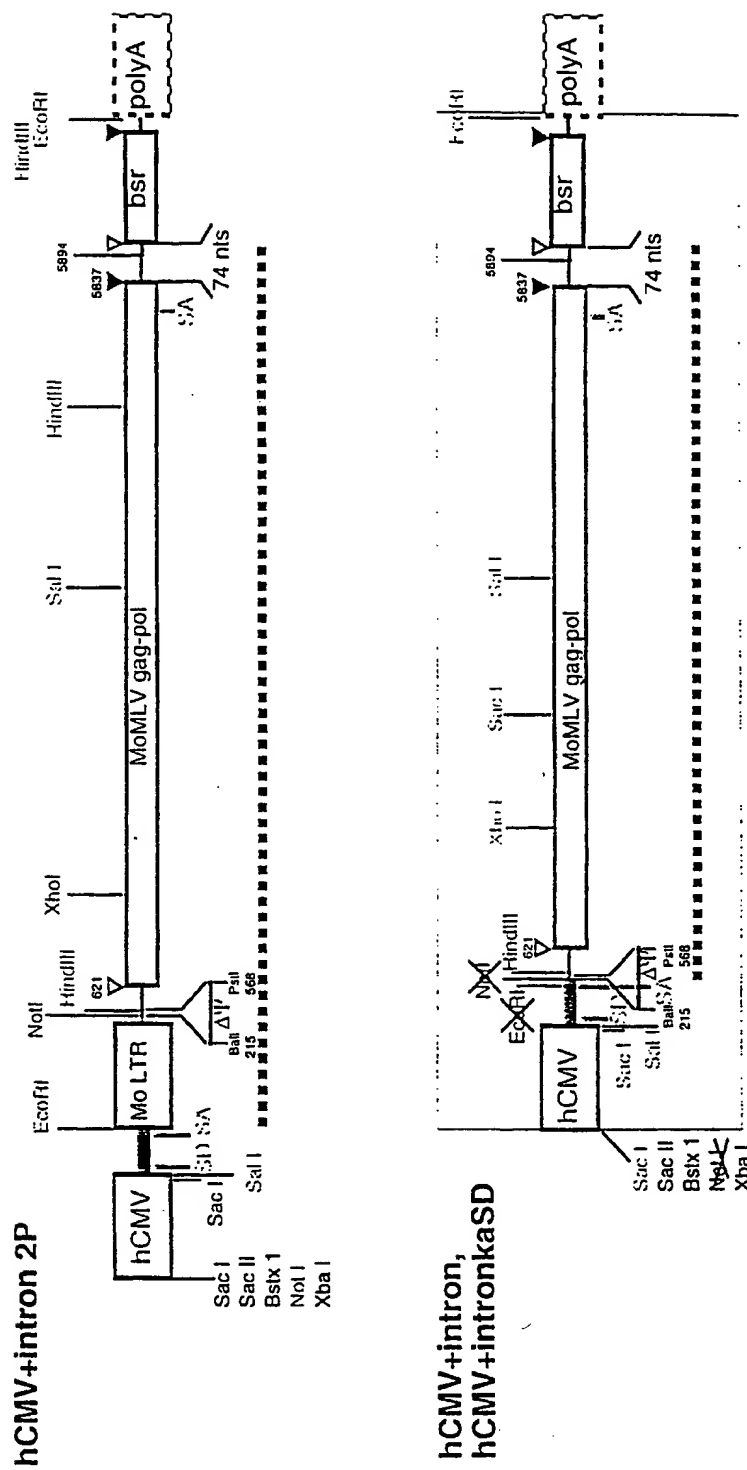
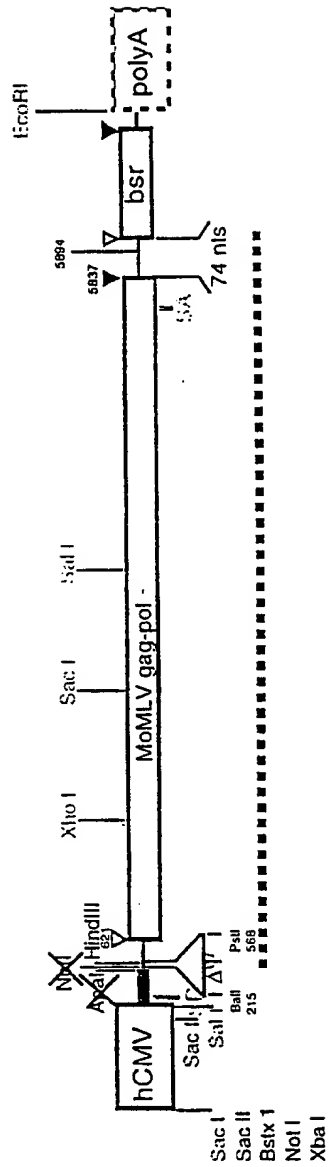


Figure 5. Genetic structure of gag-pol constructs (page 2/3)

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hCMV+SD intron



hCMV+leader

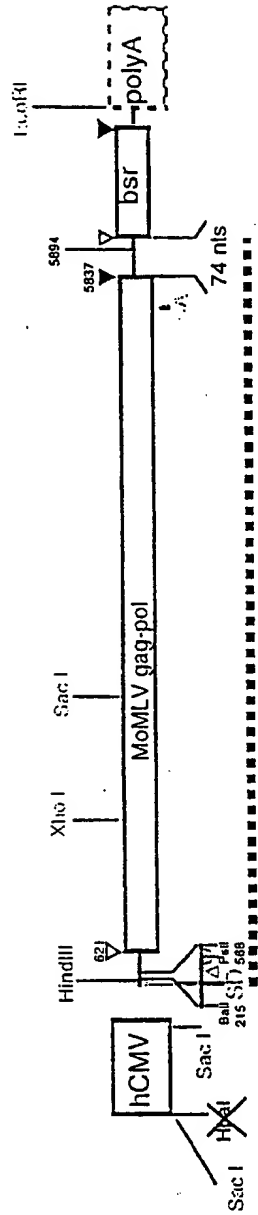


Figure 5. Genetic structure of gag-pol constructs (page 3/3)

Figure 6. CeB Sequence

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1

AATGAAAGAC	CCCACCTGTA	GGTTTGGCAA	GCTAGCTTAA	GTAACGCCAT	TTTGCAAGGC	60
ATGGAAAAAT	ACATAACTGA	GAATAGAGAA	GTTTCAGATCA	AGGTCAGGAA	CAGATGGAAC	120
AGCTGAATAT	GGGCCAAACA	GGATATCTGT	GGTAAGCAGT	TCCTGCCCCG	GCTCAGGGCC	180
AAGAACAGAT	GGAACAGCTG	AATATGGGCC	AAACAGGATA	TCTGTGGTAA	GCAGTTCCTG	240
CCCCGGCTCA	GGGCCAAGAA	CAGATGGTCC	CCAGATGCGG	TCCAGCCCTC	AGCAGTTTCT	300
AGAGAACCAT	CAGATGTTTC	CAGGGTGCCG	CAAGGACCTG	AAATGACCCT	GTGCCTTATT	360
TGAACATAAC	AATCAGTTTC	CTTCTCGCTT	CTGTTTCGCG	GCTTCTGCTC	CCCGAGCTCA	420
ATAAAAGAGC	CCACAACCCC	TCACTCGGGG	CGCCAGTCCT	CCGATTGACT	GAGTCGCCCC	480
GGTACCCGTG	TATCCAATAA	ACCCTCTTGC	AGTTGCATCC	GACTTGTGGT	CTCGCTGTTC	540
CTTGGGAGGG	TCTCTCTGTA	GTGATTGACT	ACCCGTGAGC	GGGGGTCTTT	CATTTGGGGG	600
CTCGTCCGGG	ATCCGGAGAC	CCCTGCCCCG	GGACCACCGA	CCCACCACCG	GGAGGTAAGC	660
TGGAAGCTTC	TGCAGCATCG	TTCTGTGTTG	TCTCTGTCTG	ACTGTGTTTC	TGTATTTGTC	720
TGAGAATATG	GGCCAGACTG	TTACCACTCC	CTTAAGTTTG	ACCTTAGGTC	ACTGGAAAGA	780
TGTCGAGCGG	ATCGCTCACA	ACCAGTCGGT	AGATGTCAAG	AAGAGACGTT	GGTTTACCTT	840
CTGCTCTGCA	GAATGGCCAA	CCTTTAACGT	CGGATGGCCG	CGAGACGGCA	CCTTTAACC	900
AGACCTCATC	ACCCAGGTTA	AGATCAAGGT	CTTTTCACCT	GGCCCGCATG	GACACCCAGA	960
CCAGGTCCCC	TACATCGTGA	CCTGGGAAGC	CTTGGCTTTT	GACCCCCCTC	CCTGGGTCAA	1020
GCCCTTTGTA	CACCTTAAGC	CTCCGCCTCC	TCTTCTCCCA	TCCGCCCGCT	CTCTCCCCCT	1080
TGAACCTCCT	CGTTGACCCC	CGCCTCGATC	CTCCCTTTAT	CCAGCCCTCA	CTCCTTCTCT	1140
AGGCGCCAAA	CCTAAACCTC	AAGTTCTTTC	TGACAGTGGG	GGGCCGCTCA	TGCACCTACT	1200
TACAGAAGAC	CCCCCGCCTT	ATAGGACCCC	AAGACACCCC	CCTTCCGACA	GGGACGGAAA	1260
TGGTGGAGAA	GCGACCCCTG	CGGGAGAGGC	ACCGGACCCC	TCCCCAATGG	CATCTCGCCT	1320
ACGTGGGAGA	CGGGAGCCCC	CTGTGGCCGA	CTCCACTACC	TCCGAGGCAT	TCCCCCTCCG	1380
CGCAGGAGGA	AACGGACAGC	TTCAATACTG	GCCGTCTCTC	TCTTCTGACC	TTTACAACCTG	1440
GAAAAAATAAT	AACCTTCTTT	TTTCTGAAGA	TCCAGGTAAA	CTGACAGCTC	TGATCGAGTC	1500
TGTTCTCATC	ACCCATCAGC	CCACCTGGGA	CGACTGTCAG	CAGCTGTTGG	GGACTCTGCT	1560
GACCGGAGAA	GAAAAACAAC	GGGTGCTCTT	AGAGGCTAGA	AAGGCGGTGC	GGGGCGATGA	1620
TGGGCGCCCC	ACTCAACTGC	CCAATGAAGT	CGATGCGCGT	TTTCCCTTCG	AGCGCCGAGA	1680
CTGGGATTAC	ACCACCCAGG	CAGGTAGGAA	CCACCTAGTC	CACATCGGCC	AGTTGCTCCT	1740
AGCGGGTCTC	CAAAACGCGG	GCAGAAGCCC	CACCAATTTG	GCCAAGGTAA	AAGGAATAAC	1800
ACAAGGGCCC	AATGAGTCTC	CCTCGGCCCT	CCTAGAGAGA	CTTAAGGAAG	CCTATCGCAG	1860
GTACACTCCT	TATGACCCCT	AGGACCCAGG	GCAAGAAACT	AATGTGTCTA	TGCTTTTCAT	1920
TTGGCAGTCT	GCCCCAGACA	TTGGGAGAAA	GTTAGAGAGG	TTAGAAGATT	TAAAAAACAA	1980
GACGCTTGGA	GATTTGGTTA	GAGAGGCAGA	AAAGATCTTT	AATAAACGAG	AAACCCCGGA	2040
AGAAAGAGAG	GAACGTATCA	GGAGAGAAAC	AGAGAGAAAC	GAAGAACGCC	GTAGGACAGA	2100
GGATGAGCAG	AAAGAGAAAG	AAAGAGATCG	TAGGAGACAT	AGAGAGATGA	GCAAGCTATT	2160
GGCCACTGTC	GTTAGTGGAG	AGAAACAGGA	TAGACAGGGA	GGAGAACGAA	GGAGGTCCCA	2220
ACTCGATCGC	GACCACTGTG	CCTACTGCAA	AGAAAAGGGG	CACCTGGGCTA	AAGATTGTCC	2280
CAAGAAACCA	CGAGACCTCT	GGGGACCAAG	ACCCACAGCC	TCCCTCCTGA	CCCTAGATGA	2340
CTAGGGAGGT	CAGGGTCAGG	AGCCCCCCCC	TGAACCCAGG	ATAACCTCTA	AAGTCGGGGG	2400
GCAACCCGTC	ACCTTCTCTG	TAGATACTGG	GGCCCAACAC	TCCGTGTCTA	CCCAAAATCC	2460
TGGACCCCTA	AGTGATAAGT	CTGCCTGGGT	CCAAGGGGCT	ACTGGAGGAA	AGCGGTATCG	2520
CTGGACCACG	GATCGCAAGG	TACATCTAGC	TACCGGTAAG	GTCACCCACT	CTTTCCTCCA	2580
TGTACCAGAC	TGTCCTCTATC	CTCTGTAGG	AAGAGATTTG	CTGACTAAAC	TAAAAGCCCA	2640
AATCCACTTT	GAGGGATCAG	GAGCTCAGGT	TATGGGACCA	ATGGGGCAGC	CCCTGCAAGT	2700
GTTGACCCTA	AATATAGAAG	ATGAGCATCG	GCTACATGAG	ACCTCAAAAG	AGCCAGATGT	2760
TTCTCTAGGG	TCCACATGGC	TGTCTGATTT	TCCTCAGGCC	TGGCGGGA	CCGGGGGCAT	2820
GGGACTGGCA	GTTCCGCAAG	CTCCTCTGAT	CATACCTCTG	AAAGCAACCT	CTACCCCGCT	2880
GTCCATAAAA	CAATACCCCA	TGTCACAAGA	AGCCAGACTG	GGGATCAAGC	CCCACATACA	2940
GAGACTGTTG	GACCAGGGAA	TACTGGTACC	CTGCCAGTCC	CCCTGGAACA	CGCCCTTGCT	3000
ACCCGTTAAG	AAACCAGGGA	CTAATGATTA	TAGGCCTGTC	CAGGATCTGA	GAGAAAGTCAA	3060
CAAGCGGGTG	GAAGACATCC	ACCCACCCGT	GCCCAACCCCT	TACAACCTCT	TGAGCGGGCT	3120
CCACCCGTCC	CACCACTGGT	ACACTGTGCT	TGATTTAAAG	GATGCCTTTT	TCTGCCTGAG	3180
ACTCCACCCC	ACCAGTCAGC	CTCTCTTCGC	CTTTGAGTGG	AGAGATCCAG	AGATGGGAAT	3240
CTCAGGACAA	TTGACCTGGA	CCAGACTCCC	ACAGGGTTTC	AAAAACAGTC	CCACCTGTT	3300
TGATGAGGCA	CTGCACAGAG	ACCTAGCAGA	CTTCCGGATC	CAGCACCCAG	ACTTGATCCT	3360
GCTACAGTAC	GTGGATGACT	TACTGTGGC	CGCCACTTCT	GAGCTAGACT	GCCAACAAGG	3420
TACTCGGGCC	CTGTTACAAA	CCCTAGGGAA	CCTCGGGTAT	CGGGCCTCGG	CCAAGAAAGC	3480
CCAAATTTGC	CAGAAACAGG	TCAAGTATCT	GGGGTATCTT	CTAAAAGAGG	GTCAGAGATG	3540
GCTGACTGAG	GCCAGAAAAG	AGACTGTGAT	GGGGCAGCCT	ACTCCGAAGA	CCCTCGACA	3600
ACTAAGGGAG	TTCTTAGGGA	CGGCAGGCTT	CTGTGCGCTC	TGGATCCCTG	GGTTTGCGA	3660
AATGGCAGCC	CCCTTGTAACC	CTCTACCCAA	AACGGGGACT	CTGTTTAATT	GGGGCCGAGA	3720
CCAACAAAAG	GCCTATCAAG	AAATCAAGCA	AGCTCTTCTA	ACTGCCCCAG	CCCTGGGGTT	3780
GCCAGATTTG	ACTAAGCCCT	TTGAACTCTT	TGTCGACGAG	AAGCAGGGCT	ACGCCAAAGG	3840
TGTCCTAACG	CAAAAACCTG	GACCTTGGCG	TCGGCCGGTG	GCCTACCTGT	CCAAAAGCT	3900
AGACCCAGTA	GCAGCTGGGT	GGCCCCCTTG	CCTACGGATG	GTAGCAGCCA	TTGCCGTACT	3960
GACAAAGGAT	GCAGGCAAGC	TAACCATGGG	ACAGCCACTA	GTCATTCTGG	CCCCCATG	4020
AGTAGAGGCA	CTAGTCAAAC	AACCCCCCGA	CCGCTGGCTT	TCCAACGCCC	GGATGACTCA	4080

Figure 6. CeB Sequence

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CTATCAGGCC	TTGCTTTTGG	ACACGGACCG	GGTCCAGTTC	GGACCGGTGG	TAGCCCTGAA	4140
CCCCGCTACG	CTGCTCCCAC	TGCTTGAGGA	AGGGCTGCAA	CACAAC TGCC	TTGATATCCT	4200
GGCCGAAGCC	CACGGAACCC	GACCCGACCT	AACGGACAG	CCGCTCCAG	ACGCCGACCA	4260
CACCTGGTAC	ACGGATGGAA	GCAGTCTCTT	ACAAGAGGGA	CAGCGTAAGG	CGGCAGCTGC	4320
GGTGACCACC	GAGACCGAGG	TAATCTGGGC	TAAAGCCCTG	CCAGCCGGA	CATCCGCTCA	4380
CGGGGCTGAA	CTGATAGCAC	TCACCCAGGC	CCTAAGATG	GCAGAAGGTA	AGAAGCTAAA	4440
TGTTTATACT	GATAGCCGTT	ATGCTTTTGC	TACTGCCCCT	ATCCATGGAG	AAATATACAG	4500
AAGGCGTGGG	TTGCTCACAT	CAGAAGGCAA	AGAGATCAAA	AATAAAGACG	AGATCTTGGC	4560
CCTACTAAAA	GCCCTCTTTC	TGCCCAAAAG	ACTTAGCATA	ATCCATTGTC	CAGGACATCA	4620
AAAGGGACAC	AGCCCGAGG	CTAGAGGCAA	CCGGATGGCT	GACCAAGCGG	CCCGAAAGGC	4680
AGCCATCACA	GAGACTCCAG	ACACCTCTAC	CCTCCTCATA	GAAAATTCAT	CACCCCTACAC	4740
CTCAGAACAT	TTTCATTACA	CAGTGACTGA	TATAAAGGAC	CTAACCAAGT	TGGGGGCCAT	4800
TTATGATAAA	ACAAAGAAGT	ATTGGGTCTA	CCAAGGAAAA	CCTGTGATGC	CTGACCAGTT	4860
TACTTTTGAA	TTATTAGACT	TTCTTCATCA	GCTGACTCAC	CTCAGCTTCT	CAAAAATGAA	4920
GGCTCTCCTA	GAGAGAAGCC	ACAGTCCCTA	CTACATGCTG	AACCGGGATC	GAACACTCAA	4980
AAATATCACT	GAGACCTGCA	AAGCTTGTGC	ACAAGTCAAC	GCCAGCAAGT	CTGCCGTTAA	5040
ACAGGGAAT	AGGGTCCGCG	GGCATCGGCC	CGGCATCAT	TGGGAGATCG	ATTTCACCGA	5100
GATAAAGCCC	GGATTGTATG	GCTATAAATA	TCTTCTAGTT	TTTATAGATA	CCTTTTCTGG	5160
CTGGATAGAA	GCCTTCCCAA	CCAAGAAAGA	AACCGCCAAG	GTGTAACCA	AGAAGCTACT	5220
AGAGGAGATC	TTCCCCAGGT	TCGGCATGCC	TCAGGTATTG	GGAAGTGACA	ATGGGCCTGC	5280
CTTCGTCTCC	AAGGTGAGTG	AGACAGTGGC	CGATCTGTTG	GGGATTGATT	GGAAATTACA	5340
TTGTGCATAC	AGACCCCAAA	GCTCAGGCCA	GGTAGAAAGA	ATGAATAGAA	CCATCAAGGA	5400
GACTTTAACT	AAATTAACCC	TTGCAACTGG	CTCTAGAGAC	TGGGTGCTCC	TACTCCCTTT	5460
AGCCCTGTAC	CGAGCCCGCA	ACACGCCGGG	CCCCCATGGC	CTCACCCCAT	ATGAGATCTT	5520
ATATGGGGCA	CCCCCGCCCC	TTGTAAACTT	CCCTGACCCCT	GACATGACAA	GAGTTACTAA	5580
CAGCCCTCT	CTCCAAGCTC	ACTTACAGGC	TCTCTACTTA	GTCCAGCAGC	AAGTCTGGAG	5640
ACCTCTGGCG	GCAGCCTACC	AAGAACAAC	GGACCGACCG	GTGGTACCTC	ACCCTTACCG	5700
AGTCGGCGAC	ACAGTGTGGG	TCCGCCGACA	CCAGACTAAG	AACCTAGAAC	CTCGCTGGAA	5760
AGGACCTTAC	ACAGTCTGCG	TGACCACCCC	CACCGCCCTC	AAAGTAGACG	GCATCGCAGC	5820
TTGGATACAC	GCCGCCACAG	TGAAGGCTGC	CGACCCCGGG	GGTGGACCAT	CCTCTAGACT	5880
GACATGGCGT	GTCTAACCGT	CTCAAAACCC	CTTAAATAA	AGGTTAACCC	GCGAGGCCCC	5940
CTAATCCCT	TAATCTCTCT	GATGCTCAGA	GGGGTCAGTA	CTGCTTCGCC	CGGCTCCAGT	6000
GCGGCCCAGC	CGGCCACCAT	GAAAACATTT	AACATTTCTC	AACAAGATCT	AGAATTAGTA	6060
GAAGTAGCGA	CAGAGAAGAT	TACAATGCTT	TATGAGGATA	ATAAACATCA	TGTGGGAGCG	6120
GCAATTCGTA	CGAAAACAGG	AGAAATCATT	TCGGCAGTAC	ATATTGAAGC	GTATATAGGA	6180
CGAGTAACTG	TTTGTGCAGA	AGCCATTGCG	ATTGGTAGTG	CAGTTTCGAA	TGGACAAAAG	6240
GATTTTGACA	CGATTGTAGC	TGTTAGACAC	CCTTATTCTG	ACGAAGTAGA	TAGAAGTATT	6300
CGAGTGGTAA	GTCTTGTGAG	TATGTGTAGG	GAGTTGATTT	CAGACTATGC	ACCAGATTGT	6360
TTTGTGTTAA	TAGAAATGAA	TGGCAAGTTA	GTCAAAACTA	CGATTGAAGA	ACTCATTTCCA	6420
CTCAATATA	CCCGAAATTA	AAAGTTTAC	CACCAAGCTT	ATCGATTAGT	CCAAATTGTT	6480
AAAGACAGGA	TATCAGTGGT	CCAGGCTCTA	GTTTTGACTC	AACAATATCA	CCAGCTGAAG	6540
CCTATAGAGT	ACGAGCCATA	GATAAAATAA	AAGATTTTAT	TTAGTCTCCA	GAAAAGGGG	6600
GGAATGAAAG	ACCCACCTG	TAGGTTTGGC	AAGCTAGCTT	AAGTAACGCC	ATTTTGCAAG	6660
GCATGGAAAA	ATACATAACT	GAGAATAGAG	AAGTTCAGAT	CAAGGTCAGG	AACAGATGGA	6720
ACAGTCGAGA	ACTTGTTTAT	TGCAGCTTAT	AATGGTTACA	AATAAAGCAA	TAGCATCACA	6780
AATTTACAA	ATAAAGCATT	TTTTTCACTG	CATTCTAGTT	GTGGTTTGTC	CAAACCTCATC	6840
AATGTATCTT	ATCATGTCTG	GATCCCCAGG	AAGCTCCTCT	GTGTCCCTCAT	AAACCCTAAC	6900
CTCCTCTACT	TGAGAGGACA	TTCCAATCAT	AGGCTGCCCA	TCCACCCTCT	GTGTCTCTCT	6960
GTTAATTAGG	TCACTTAACA	AAAAGGAAAT	TGGGTAGGGG	TTTTTCACAG	ACCGCTTTCT	7020
AAGGGTAATT	TTAAAATATC	TGGGAAGTCC	CTTCCACTGC	TGTGTCCAG	AAGTGTGGT	7080
AAACAGCCCA	CAAATGTCAA	CAGCAGAAAC	ATACAAGCTG	TCAGCTTTGC	ACAAGGGCCC	7140
AACACCCCTG	TCATCAAGAA	GCACCTGTGGT	TGCTGTGTTA	GTAATGTGCA	AAACAGGAGG	7200
CACATTTTCC	CCACCTGTGT	AGGTTCCAAA	ATATCTAGTG	TTTTCATTTT	TACTTGGATC	7260
AGGAACCCAG	CACCTCCACTG	GATAAGCATT	ATCCTTATCC	AAAACAGCCT	TGTGGTCAGT	7320
GTTTCATCTG	TGACTGTCAA	CTGTAGCATT	TTTTGGGGTT	ACAGTTTGAG	CAGGATATTT	7380
GGTCCTGTAG	TTTGCTAACA	CACCCGTCAG	CTCCAAAGGT	TCCCCACCAA	CAGCAAAAAA	7440
ATGAAAAATT	GACCCCTGAA	TGGGTTTTC	AGCACCATT	TCATGAGTTT	TTTGTGTCCC	7500
TGAATGCAAG	TTTAACATAG	CAGTTACCCC	AATAACCTCA	GTTTAAACAG	TAACAGCTTC	7560
CCACATCAAA	ATATTTCAC	AGGTTAAGTC	CTEATTTAAA	TTAGGCAAG	GAATTC	7616

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Figure 7. hCMV+intron Sequence

1

AGATCTCCCG	ATCCCTATG	GTCGACTCTC	AGTACAATCT	GCTCTGATGC	CGCATAGTTA	60
AGCCAGTATC	TGCTCCCTGC	TTGTGTGTTG	GAGGTCGCTG	AGTAGTGCGC	GAGCAAAATT	120
TAAGCTACAA	CAAGGCAAGG	CTTGACCGAC	AATTGCATGA	AGAATCTGCT	TAGGGTTAGG	180
CGTTTTGCGC	TGCTTCGCGA	TGTACGGGCC	AGATATACGC	GTTGACATTG	ATTATTGACT	240
AGTTATTAAT	AGTAATCAAT	TACGGGGTCA	TTAGTTCATA	GCCCATATAT	GGAGTCCGCG	300
GTTACATAAC	TTACGGTAAA	TGGCCCGCCT	GGCTGACCGC	CCAACGACCC	CCGCCCATTG	360
ACGTCAATAA	TGACGTATGT	TCCCATAGTA	ACGCCAATAG	GGACTTTCCA	TTGACGTCAA	420
TGGGTGGACT	ATTTACGGTA	AACTGCCCAC	TTGGCAGTAC	ATCAAGTGTA	TCATATGCCA	480
AGTACGCCCC	CTATTGACGT	CAATGACGGT	AAATGGCCCC	CCTGGCATTA	TGCCCAGTAC	540
ATGACCTTAT	GGGACTTTCC	TACTTGGCAG	TACATCTACG	TATTAGTCAT	CGCTATTACC	600
ATGGTGATGC	GGTTTTGGGA	GTACATCAAT	GGGCGTGGAT	AGCGGTTTGA	CTCACGGGGA	660
TTTCCAAGTC	TCCACCCCAT	TGACGTCAAT	GGGAGTTTGT	TTTGGCACCA	AAATCAACGG	720
GACTTTCCTA	AATGTCGTAA	CAACTCCGCC	CCATTGACGC	AAATGGGCGG	TAGGCGGTGA	780
CGGTGGGAGG	TCATATAAAG	CAGAGCTCTC	TGGCTAACTA	GAGAACCAC	TGCTTAACTG	840
GCTTATCGAA	ATGTCGACTG	AGAACTTCAG	GGTGAGTTTG	GGGACCCTTG	ATTGTTCTTT	900
CTTTTTCGCT	ATGTGTAAT	TCATGTTATA	TGGAGGGGGC	AAAGTTTTC	GGGTGTTGTT	960
TAGAATGGGA	AGATGTCCCT	TGTATCACCA	TGGACCCTCA	TGATAATTTT	GTTTCTTTCA	1020
CTTTCTACTC	TCTTATGACA	CATTGTCTCC	TCTTATTTTC	TTTTCATTTT	CTGTAACCTT	1080
TTCGTTAAAC	TTTAGCTTGC	ATTTGTAACG	AATTTTAA	TTCACTTTGT	TTTATTTGTC	1140
AGATTGTAAG	TACTTTCTCT	AATCACTTTT	TTTTCAAGGC	AATCAGGGTA	TATTATATTG	1200
TACTTACAGA	CAGTTTTAGA	GAACAATTGT	TATAATTAAA	TGATAAGGTA	GAATATTCT	1260
GCATATAAAT	TCTGGCTGGC	GTGGAATAT	TCTTATTTGGT	AGAAACAAC	ACATCCTGGT	1320
CATCATCCTG	CCTTTCTCTT	TATGGTTACA	ATGATATACA	CTGTTTGAGA	TGAGGATAAA	1380
ATACTCTGAG	TCCAAACCGG	GCCCTCTG	TAACCATGTT	CATGCCCTCT	TCTTTTCTCT	1440
ACAGCTCCTG	GGCAACGTGC	TGGTTGTTGT	GCTGTCTCAT	CATTTTGGCA	AGAATTGGCC	1500
GCAAGCTTCT	GCAGCATCGT	TCTGTGTTGT	CTCTGTCTGA	CTGTGTTTCT	GTATTTGTCT	1560
GAGAATATGG	GCCAGACTGT	TACCACTCCC	TTAAGTTTGA	CCTTAGGTCA	CTGGAAGAT	1620
GTCGAGCGGA	TCCCTACAA	CCAGTCGGTA	GATGTCAAGA	AGAGACGTTG	GGTACCTTC	1680
TGCTCTGCAG	AATGGCCAAC	CTTTAACGTC	GGATGGCCGC	GAGACGGCAC	CTTTAACCGA	1740
GACCTCATCA	CCCAGGTTAA	GATCAAGGTC	TTTTCACTG	GCCCGCATGG	ACACCCAGAC	1800
CAGGTCCCTC	ACATCGTGAC	CTGGAAGGCC	TTGGCTTTTG	ACCCCCCTCC	CTGGGTCAAG	1860
CCCTTTGTAC	ACCCTAAGCC	TCCGCTCCT	CTTCTCCAT	CCGCCCCGTC	TCTCCCCCTT	1920
GAACCTCCTC	GTTCGACCCC	GCCTCGATCC	TCCCTTTATC	CAGCCCTCAC	TCCTTCTCTA	1980
GGCGCCAAAC	CTAAACCTCA	AGTTCTTTCT	GACAGTGGGG	GGCCGCTCAT	CGACCTACTT	2040
ACAGAAGACC	CCCCGCTTA	TAGGGACCCA	AGACCACCCC	CTTCCGACAG	GGACGGAAAT	2100
GGTGGAGAAG	CGACCCCTGC	GGGAGAGGCA	CCGGACCCCT	CCCCAATGGC	ATCTCGCCTA	2160
CGTGGGAGAC	GGGAGCCCCC	TGTGGCCGAC	TCCACTACCT	CGCAGGCATT	CCCCCTCCGC	2220
GCAGGAGGAA	ACCGACAGCT	TCAAATACTGG	CCGTTCTCCT	CTTCTGACCT	TTACAACCTG	2280
AAAAATAATA	ACCCTTCTTT	TTCTGAAGAT	CCAGGTAAAC	TGACAGCTCT	GATCGAGTCT	2340
GTTCTCATCA	CCCATCAGCC	CACTTGGGAC	GACTGTCAGC	AGCTGTTGGG	GACTCTGCTG	2400
ACCGGAGAAG	AAAAACAACG	GGTGCTCTTA	GAGGCTAGAA	AGGCGGTGCG	GGGCGATGAT	2460
GGGCGCCCCA	CTCAACTGCC	CAATGAAGTC	GATGCCGCTT	TTCCCCCGA	GCGCCAGAC	2520
TGGGATTACA	CCACCCAGGC	AGGTAGGAAC	CACCTAGTCC	ACTATCGCCA	GTTGCTCCTA	2580
GCGGGTCTCC	AAAACGCGGG	CAGAAGCCCC	ACCAATTTGG	CCAAGGTAAA	AGGAATAACA	2640
CAAGGGCCCA	CTGAGTCTCC	CTCGGCTTCC	CTAGAGAGAC	TTAAGGAAGC	CTATCGCAGG	2700
TACACTCCTT	ATGACCCTGA	GGACCCAGGG	CAAGAACTA	ATGTGTCTAT	GTCTTTTCATT	2760
TGGCAGTCTG	CCCCAGACAT	TGGGAGAAAG	TTAGAGAGGT	TAGAAGATTT	AAAAAACAA	2820
ACGCTTGGAG	ATTGTTGTTAG	AGAGGCAGAA	AAGATCTTTA	ATAAACGAGA	AACCCCGGAA	2880
GAAAGAGAGG	AACGTATCAG	GAGAGAAACA	GAGGAAAAAG	AAGAACGCCG	TAGGACAGAG	2940
GATGAGCAGA	AAGAGAAAGA	AAGAGATCGT	AGGAGACATA	GAGAGATGAG	CAAGCTATTG	3000
GCCACTGTCT	TTAGTGGACA	GAAACAGGAT	AGACAGGGAG	GAGAACGAAG	GAGGTCCCAA	3060
CTCGATCGCG	ACCAGTGTGC	CTACTGCAAA	GAAAAGGGGC	ACTGGGCTAA	AGATTGTCCC	3120
AAGAAACCA	GAGGACCTCG	GGGACCAAGA	CCCCAGACCT	CCCTCCTGAC	CCTAGATGAC	3180
TAGGGAGGTC	AGGGTCAGGA	GCCCCCCCC	GAACCCAGGA	TAACCTCAA	AGTCGGGGGG	3240
CAACCCGTCA	CCTTCCTGGT	AGATACTGGG	GCCCAACACT	CCGTGCTGAC	CCAAAATCCT	3300
GGACCCCTAA	GTGATAAGTC	TGCCTGGGTC	CAAGGGGCTA	CTGGAGGAAA	GCGGTATCGC	3360
TGGACCAACG	ATCGCAAAGT	ACATCTAGCT	ACCGGTAAGG	TCACCCACTC	TTTCTCTCAT	3420
GTACCAAGCT	GTCCCTATCC	TCTGTAGGA	AGAGATTGTC	TGACTAAACT	AAAAGCCCAA	3480
ATCCACTTTG	AGGGATCAGG	AGCTCAGGTT	ATGGGACCAA	TGGGGCAGCC	CCTGCAAGTG	3540
TTGACCCCTA	ATATAGAAGA	TGAGCATCGG	CTACATGAGA	CCTCAAAAGA	GCCAGATGTT	3600
TCTCTAGGGT	CCACATGGCT	GTCTGATTTT	CCTCAGGCCT	GGGCGGAAC	CGGGGGCATG	3660
GGACTGGCAG	TTCCGCAAGC	TCCTCTGATC	ATACCTCTGA	AAGCAACCTC	TACCCCGCTG	3720
TCCATAAAAC	AATACCCCAT	GTCACAAGAA	GCCAGACTGG	GGATCAAGCC	CCACATACAG	3780
AGACTGTTGG	ACCAGGGAAT	ACTGGTACCC	TGCCAGTCCC	CCTGGAACAC	GCCCCCTGTA	3840
CCCGTTAAGA	AACCAAGGAC	TAATGATTAT	AGGCCTGTCC	AGGATCTGAG	AGAAGTCAAC	3900
AAGCGGGTGG	AAGACATCCA	CCCCACCGTG	CCCAACCCCT	ACAACCTCTT	GAGCGGGCTC	3960
CCACCGTCCC	ACCAGTGGTA	CACTGTGCTT	GATTTAAAGG	ATGCCTTTT	CTGCCTGAGA	4020
CTCCACCCCA	CCAGTCAGCC	TCTCTTCGCC	TTTGAGTGGA	GAGATCCAGA	GATGGGAATC	4080

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2

Figure 7. hCMV+intron Sequence

TCAGGACAAT	TGACCTGGAC	CAGACTCCCA	CAGGGTTTCA	AAAACAGTCC	CACCCTGTTT	4140
GATGAGGCAC	TGCACAGAGA	CCTAGCAGAC	TTCCGGATCC	AGCACCCAGA	CTTGATCCTG	4200
CTACAGTACG	TGGATGACTT	ACTGCTGGCC	GCCACTTCTG	AGCTAGACTG	CCAACAAGGT	4260
ACTCGGGCCC	TGTTACAAAC	CCTAGGGAAC	CTCGGGTATC	GGGCCTCGGC	CAAGAAAGCC	4320
CAAATTTGCC	AGAAACAGGT	CAAGTATCTG	GGGTATCTTC	TAAAAGAGGG	TCAGAGATGG	4380
CTGACTGAGG	CCAGAAAAGA	GACTGTGATG	GGGCAGCCTA	CTCCGAAGAC	CCCTCGACAA	4440
CTAAGGGAGT	TCCTAGGGAC	GGCAGGCTTC	TGTCGCCTCT	GGATCCCTGG	GTTTGCAGAA	4500
ATGGCAGCCC	CCTTGTAACC	TCTCACCAAA	ACGGGGACTC	TGTTTAATTG	GGGCCAGAC	4560
CAACAAAAGG	CCTATCAAGA	AATCAAGCAA	GCTCTTCTAA	CTGCCCCAGC	CCTGGGGTTG	4620
CCAGATTTGA	CTAAGCCCTT	TGAATCTTTT	GTCGACGAGA	AGCAGGGCTA	CGCCAAAGGT	4680
GTCCCTAACG	AAAAACTGGG	ACCTTGGCGT	CGGCCGGTGG	CCTACCTGTC	CAAAAAGCTA	4740
GACCCAGTAG	CAGCTGGGTG	GCCCCCTTGC	CTACGGATGG	TAGCAGCCAT	TGCCGTAAGT	4800
ACAAAGGATG	CAGGCAAGCT	AACCATGGGA	CAGCCACTAG	TCATTCTGGC	CCCCCATGCA	4860
GTAGAGGCAC	TAGTCAAACA	ACCCCCGAC	CGCTGGCTTT	CCAACGCCCG	GATGACTCAC	4920
TATCAGGCCT	TGCTTTTGGG	CACGGACCGG	GTCCAGTTTC	GACCGGTGGT	AGCCCTGAAC	4980
CCGGCTACGC	TGCTCCCACT	GCCTGAGGAA	GGGCTGCAAC	ACAACCTGCT	TGATATCCTG	5040
GCCGAAGCCC	ACGGAACCCG	ACCCGACCTA	ACGGACCAGC	CGCTCCGAGA	CGCCGACCAC	5100
ACCTGGTACA	CGGATGGAAG	CAGTCTCTTA	CAAGAGGGAC	AGCGTAAGCG	GGGAGCTGCG	5160
GTGACCACCG	AGACCGAGGT	AATCTGGGCT	AAAGCCCTGC	CAGCCGGGAC	ATCCGCTCAG	5220
CGGGCTGAAC	TGATAGCACT	CACCCAGGCC	CTAAAGATGG	CAGAAGGTAA	GAAGCTAAAT	5280
GTTTATACTG	ATAGCCGTTA	TGCTTTTGCT	ACTGCCCATG	TCCATGGAGA	AATATACAGA	5340
AGGCGTGGGT	TGCTCACATC	AGAAGGCAAA	GAGATCAAAA	ATAAAGACGA	GATCTTGGCC	5400
CTACTAAAAG	CCCTCTTTCT	GCCCAAAAGA	CTTAGCATAA	TCCATTGTCC	AGGACATCAA	5460
AAGGGACACA	GCGCCGAGGC	TAGAGGCAAC	CGGATGGCTG	ACCAAGCGGC	CCGAAAGGCA	5520
GCCATCACAG	AGACTCCAGA	CACCTCTACC	CTCCTCATAG	AAAATTCATC	ACCCTACACC	5580
TCAGAACATT	TTCATTACAC	AGTGACTGAT	ATAAAGGACC	TAACCAAGTT	GGGGGCCATT	5640
TATGATAAAA	CAAAGAAGTA	TTGGGTCTAC	CAAGGAAAAC	CTGTGATGCC	TGACCAGTTT	5700
ACTTTTGAAT	TATTAGACTT	TCTTCATCAG	TCTGACTCACC	TCAGCTTCTC	AAAAATGAAG	5760
GCCTCTCTAG	AGAGAAGCCA	CAGTCCCTAC	TACATGCTGA	ACCGGGATCG	AACACTCAAA	5820
AATATCACTG	AGACCTGCAA	AGCTTGTGCA	CAAGTCAACG	CCAGCAAGTC	TGCCGTTAAA	5880
CAGGGAACTA	GGGTCCGCGG	GCATCGGCCC	GGCACTCATT	GGGAGATCGA	TTTCACCGAG	5940
ATAAAGCCCG	GATTGTATGG	CTATAAATAT	CTTCTAGTTT	TTATAGATAC	CTTTTCTGGC	6000
TGGATAGAAG	CCTTCCCAAC	CAAGAAAGAA	ACCGCCAAGG	TCGTAACCAA	GAAGCTACTA	6060
GAGGAGATCT	TCCCCAGGTT	CGGCATGCCT	CAGGTATTGG	GAAGTACCAA	TGGGCCTGCC	6120
TTCGTCTCCA	AGGTGAGTCA	GACAGTGGCC	GATCTGTTGG	GGATTGATTG	GAAATTACAT	6180
TGTGCATACA	GACCCCAAAG	CTCAGGCCAG	GTAGAAAGAA	TGAATAGAAC	CATCAAGGAG	6240
ACTTTAACTA	AATTAACGCT	TGCAACTGGC	TCTAGAGACT	GGGTGCTCCT	ACTCCCTTTA	6300
GCCCTGTACC	GAGCCCGCAA	CACGCCGGGC	CCCCATGGCC	TCACCCCATG	TGAGATCTTA	6360
TATGGGGCAC	CCCCGCCCTT	TGTAAACTTC	CCTGACCCTG	ACATGACAAG	AGTTACTAAC	6420
AGCCCTCTCT	TCCAAGCTCA	CTTACAGGCT	CTCTACTTAG	TCCAGCACGA	AGTCTGGAGA	6480
CCTCTGGCGG	CAGCCTACCA	AGAACAACCT	GACCGACCGG	TGGTACCTCA	CCCTTACCGA	6540
GTCCGCGACA	CAGTGTGGGT	CCGCCGACAC	CAGACTAAGA	ACCTAGAACC	TCGCTGGAAA	6600
GGACCTTACA	CAGTCTGTCT	GACCACCCCC	ACCGCCCTCA	AAGTAGACGG	CATCGCAGCT	6660
TGGATACACG	CCGCCCACGT	GAAGGCTGCC	GACCCCGGGG	GTGGACCATC	CTCTAGACTG	6720
ACATGGCGCG	TTCAACGCTC	TCAAAACCCC	TTAAAAATAA	GGTTAACCCG	CGAGGCCCCC	6780
TAATCCCCCT	AATTCTTCTG	ATGCTCAGAG	GGGTCAGTAC	TGCTTCGCCC	GGCTCCAGTG	6840
CGGCCAGACC	GGCCACCATG	AAAACATTTA	ACATTTCTCA	ACAAGATCTA	GAATTAGTAG	6900
AAGTAGCGAC	AGAGAAGATT	ACAATGCTTT	ATGAGGATAA	TAAACATCAT	GTGGGAGCGG	6960
CAATTGCTAC	GAAAACAGGA	GAAATCATTT	CGGCAGTACA	TATTGAAGCG	TATATAGGAC	7020
GAGTAACTGT	TTGTGCAGAA	GCCATTGCGA	TTGGTAGTGC	AGTTTTCGAAT	GGACAAAAGG	7080
ATTTTGACAC	GATTGTAGCT	GTTAGACACC	CTTATTCTGA	CGAAGTAGAT	AGAAGTATTC	7140
GAGTGGTAAG	TCCTTGTGGT	ATGTGTAGGG	AGTTGATTTT	AGACTATGCA	CCAGATTGTT	7200
TTGTGTTAAT	AGAAATGAAT	GGCAAGTTAG	TCAAAACTAC	GATTGAAGAA	CTCATTCCAC	7260
TCAAATATAC	CCGAAATTAA	AAGTTTATAC	ACCAAGCTTA	TCGAATTC		7308

Figure 8. hCMV+intronkaSD Sequence

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AGATCTCCCG	ATCCCTTATG	GTCGACTCTC	AGTACAATCT	GCTCTGATGC	CGCATAGTTA	60
AGCCAGTATC	TGCTCCCCTG	TTGTGTGTG	GAGGTGCGTG	AGTAGTGCGC	GAGCAAAATT	120
TAAGCTACAA	CAAGGCCAAG	CTTGACCGAC	AAATTGCATGA	AGAATCTGCT	TAGGGTTAGG	180
CGTTTTGCGC	TGCTTCGCGA	TGTACGGGCC	AGATATACGC	GTTGACATTG	ATTATTGACT	240
AGTTATTAAT	AGTAATCAAT	TACGGGGTCA	TTAGTTTCATA	GCCCCATATAT	GGAGTTCGCG	300
GTTACATAAC	TTACGGTAAA	TGGCCCCGCT	GGCTGACCGC	CCAACGACCC	CCGCCCATTG	360
ACGTCAATAA	TGACGTATGT	TCCCATAGTA	ACGCCAATAG	GGACTTTCCA	TTGACGTCAA	420
TGGGTGGACT	ATTTACGGTA	AACTGCCCCAC	TTGGCAGTAC	ATCAAGTGTA	TCATATGCCA	480
AGTACGCCCT	CTATTGACGT	CAATGACGGT	AAATGGCCCCG	CCTGGCATTAT	TGCCCAGTAC	540
ATGACCTTAT	GGGACTTTCC	TACTTGCCAG	TACATCTACG	TATTAGTCAT	CGCTATTACC	600
ATGGTGATGC	GGTTTTGGCA	GTACATCAAT	GGGCGTGGAT	AGCGGTTTGA	CTCACGGGGA	660
TTTCCAAGTC	TCCACCCCAT	TGACGTCAAT	GGGAGTTTGT	TTTGGCACCA	AAATCAACGG	720
GACTTTCCAA	AATGTCGTAA	CAACTCCGCC	CCATTGACGC	AAATGGGCGG	TAGGCGTGTA	780
CGGTGGGAGG	TCTATATAAG	CAGAGCTCTC	TGGCTAACTA	GAGAACCAC	TGCTTAACTG	840
GCTTATCGAA	ATGTCGACTG	AGAACTTCAG	GGTGAGTTTG	GGGACCCCTG	ATTGTTCTTT	900
CTTTTTCGCT	ATTGTAAAT	TCATGTTATA	TGGAGGGGGC	AAAGTTTTC	GGGTGTTGTT	960
TAGAAATGGA	AGATGTCCTT	TGTATCACCA	TGGACCCCTCA	TGATAATTTT	GTTTCTTTCA	1020
CTTTCTACTC	TGTTGACAAC	CATTGCTCTC	TCTTATTTTC	TTTTCATTTT	CTGTAACCTT	1080
TTCTGTTAAAC	TTTAGCTTGC	ATTGTGAACG	AAATTTTAAA	TTCATTTTTC	TTTATTGTGC	1140
AGATTGTAAAG	TACTTTCTCT	AATCACTTTT	TTTTCAAGGC	AATCAGGGTA	TATTATATTG	1200
TACTTCAGCA	CAGTTTTAGA	GAACAATTGT	TATAATTAAA	TGATAAGGTA	GAATAATTCT	1260
GCATATAAAT	TCTGGCTGGC	GTGGAAATAT	TCTTATTGGT	AGAAACAAC	ACATCTGGT	1320
CATCATCCTG	CCTTCTCTT	TATGGTTACA	ATGATATACA	CTGTTTGAGA	TGAGGATAAA	1380
ATACTCTGAG	TCCAAACCGG	GCCCTCTGCG	TAACCATGTT	CATGCCCTCT	TCTTTTCTCT	1440
ACAGCTCCTG	GGCAACGTGC	TGGTTGTTGT	GCTGTCTCAT	CATTTTGGCA	AGAATTGGCC	1500
GCAAGCTTCT	GCAGCATCGT	TCTGTGTTGT	CTCTGTCTGA	CTGTGTTTCT	GTATTGTGCT	1560
GAGAATATGG	GCCAGACTGT	TACCACTCCC	TTAAGTTTGA	CCTTAGGTCA	CTGGAAGAT	1620
GTCGAGCGGA	TGCTCACAA	CCAGTCGGTA	GATGTCAAGA	AGAGACGTTG	GGTTACCTTC	1680
TGCTCTGCAG	AATGGCCAAC	CTTAAACGTC	GGATGGCCGC	GAGACGGCAC	CTTTAACCGA	1740
GACCTCATCA	CCAGGTTAA	GATCAAGGTC	TTTTACCTG	GCCCGCATGG	ACACCCAGAC	1800
CAGGTCCCCCT	ACCATCGTAC	CTGGGAAGCC	TTGGCTTTTG	ACCCCCCTCC	CTGGGTCAAG	1860
CCCTTTGTAC	ACCCTAAGCC	TCCGCCTCCT	CTTCTCCAT	CCGCCCGCTC	TCTCCCCCTT	1920
GAACCTCCTC	GTTGACCCCC	GCCTCGATCC	TCCCTTTATC	GAGCCCTCAC	TCCTTCTCTA	1980
GGCGGCCAAAC	CTAAACCTCA	AGTTCTTTCT	GACAGTGGGG	GGCCGCTCAT	CGACCTACTT	2040
ACAGAGAGAC	CCCCGCCTTA	TAGGGACCCA	AGACCACCCC	CTTCCGACAG	GGACGGAAAT	2100
GGTGGAGAAG	CGACCCCTGC	GGGAGAGGCA	CCGGACCCCT	CCCCAATGGC	ATCTCGCCTA	2160
CGTGGGAGAG	GGGAGCCCCC	TGTGGCCGAC	TCCACTACCT	CGCAGGCATT	CCCCCTCCGC	2220
GCAGGAGGAA	ACGGACAGCT	TCAATACTGG	CCGTCTCCT	CTTCTGACCT	TTACAACCTG	2280
AAAAATAATA	ACCCTCTCTT	TTCTGAAGAT	CCAGGTAAAC	TGACAGCTCT	GATCGAGTCT	2340
GTTCTCATCA	CCCATCAGCC	EACCTGGGAC	GACTGTCAGC	AGCTGTTGGG	GACTCTGCTG	2400
ACCGGAGAAG	AAAAACAACG	GGTGTCTTTA	GAGGCTAGAA	AGGCGGTGCG	GGCGCATGAT	2460
GGGCGCCCCA	CTCAACTGCC	CAATGAAGTC	GATGCGGCTT	TTCCCTCGA	GCGCCAGAC	2520
TGGGATTACA	CCACCCAGGC	AGGACGCAAC	CACCTAGTCC	ACTATCGCCA	GTTGCTCCTA	2580
GCGGGTCTCC	AAAACGCCGG	CAGAAGCCCC	ACCAATTTGG	CCAAGGTAAA	AGGAATAACA	2640
CAAGGGCCCC	ATGAGTCTCC	CTCGGCCTTC	CTAGAGAGAC	TTAAGGAAGC	CTATCGCAGG	2700
TACACTCCTT	ATGACCCCTG	GGACCCAGGG	CAAGAAACTA	ATGTGTCTAT	GTCTTTTCATT	2760
TGGCAGTCTG	CCCCAGACAT	TGGGAGAAAG	TTAGAGAGGT	TAGAAGATTT	AAAAACAAG	2820
ACGCTTGGAG	ATTTGGTTAG	AGAGGCAGAA	AAGATCTTTA	ATAAACGAGA	AACCCCGGAA	2880
GAAAGAGAGG	AACGTATCAG	GAGAGAAACA	GAGGAAAAAG	AAGAACGCCG	TAGGACAGAG	2940
GATGAGCAGA	AAGAGAAAGA	AAGAGATCGT	AGGAGACATA	GAGAGATGAG	CAAGCTATTG	3000
GCCACTGTCT	TTAGTTGGACA	GAAACAGGAT	AGACAGGGAG	GAGAACGAAG	GAGGTCCCAA	3060
CTCGATCGCG	ACCAGTGTGC	CTACTGCAAA	GAAAAGGGGC	ACTGGGCTAA	AGATTGTCCC	3120
AAGAAACCAC	GAGGACCTCG	GGGACCAAGA	CCCCAGACCT	CCCTCCTGAC	CCTAGATGAC	3180
TAGGGAGGTC	AGGGTCAGGA	GCCCCCCCCCT	GAACCCAGGA	TAACCTCAA	AGTCGGGGGG	3240
CAACCCGTCA	CCTTCTGGT	AGATACTGGG	GCCCAACACT	CCGTGCTGAC	CCAAAATCCT	3300
GGACCCCTAA	GTGATAAGTC	TGCCTGGGTC	CAAGGGGCTA	CTGGAGGAAA	GCGGTATCGC	3360
TGGACACAGG	ATCGCAAAGT	ACATCTAGCT	ACCGGTAAGG	TCACCACTC	TTTCTCTCAT	3420
GTACCAGACT	GTCCCTATCC	TCTGTTAGGA	AGAGATTTGC	TGACTAAACT	AAAAGCCCAA	3480
ATCCACTTTG	AGGGATCAGG	AGCTCAGGTT	ATGGGACCAA	TGGGGCAGCC	CCTGCAAGTG	3540
TTGACCTTAA	ATATAGAAGA	TGAGCATCGG	CTACATGAGA	CCTCAAAAGA	GCCAGATGTT	3600
TCTCTAGGGT	CCACATGGCT	GTCTGATTTT	CCTCAGGCTT	GGGCGGAAAC	CGGGGGCATG	3660
GGACTGGCAG	TTCGCCAAGC	TCCTCTGATC	ATACCTCTGA	AAGCAACCTC	TACCCCCGTG	3720
TCCATAAAAC	AATACCCCAT	GTCAACAAGAA	GCCAGACTGG	GGATCAAGCC	CCACATACAG	3780
AGACTGTTGG	ACCAGGGGAT	ACTGGTACCC	TGCCAGTCCC	CCTGGAAACAC	GCCCCGTGTA	3840
CCCGTTAAGA	AACCAGGGAC	TAATGATTAT	AGGCCTGTCC	AGGATCTGAG	AGAAGTCAAC	3900
AAGCGGGTGG	AAGACATCCA	CCCCACCGTG	CCCAACCTTT	ACAACCTCTT	GAGCGGGCTC	3960
CCACCGTCCC	ACCAGTGGTA	CACGTGTGCT	GATTTAAAGG	ATGCCTTTTT	CTGCCCTGAGA	4020
CTCCACCCCA	CCAGTCAGCC	TCTCTTCGCC	TTTGAGTGGA	GAGATCCAGA	GATGGGAATC	4080

Figure 8. hCMV+intronkaSD Sequence

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TCAGGACAAT	TGACCTGGAC	CAGACTCCCA	CAGGGTTTCA	AAAACAGTCC	CACCCTGTTT	4140
GATGAGGCAC	TGCACAGAGA	CCTAGCAGAC	TTCCGGATCC	AGCACCCAGA	CTTGATCCTG	4200
CTACAGTACG	TGGATGACTT	ACTGCTGGCC	GCCACTTCTG	AGCTAGACTG	CCAACAAGGT	4260
ACTCGGGCCC	TGTTACAAC	CCTAGGGAAC	CTCGGGTATC	GGGCCTCGGC	CAAGAAAGCC	4320
CAAAATTTGCC	AGAAACAGGT	CAAGTATCTG	GGGTATCTTC	TAAAAGAGGG	TCAGAGATGG	4380
CTGACTGAGG	CCAGAAAAGA	GACTGTGATG	GGGCAGCCTA	CTCCGAAGAC	CCCTCGACAA	4440
CTAAGGGAGT	TCCTAGGGAC	GGCAGGCTTC	TGTCGCCCTC	GGATCCCTGG	GTTTGCAGAA	4500
ATGGCAGCCC	CCTTGTAACC	TCTCACCAAA	ACGGGGACTC	TGTTTAATTG	GGGCCCAGAC	4560
CAACAAAAGG	CCTATCAAGA	AATCAAGCAA	GCTCTTCTAA	CTGCCCCAGC	CCTGGGGTTG	4620
CCAGATTTGA	CTAAGCCCTT	TGAACTCTTT	GTCGACGAGA	AGCAGGGCTA	CGCCAAAGGT	4680
GTCCTAACGC	AAAAACTGGG	ACCTTGGCGT	CGGCCGGTGG	CCTACCTGTC	CAAAAAGCTA	4740
GACCCAGTAG	CAGCTGGGTG	GCCCCCTTGC	CTACGGATGG	TAGCAGCCAT	TGCCGTACTG	4800
ACAAAGGATG	CAGGCAAGCT	AACCATGGGA	CAGCCACTAG	TCATTCTGGC	CCCCCATGCA	4860
GTAGAGGCAC	TAGTCAAACA	ACCCCCGAC	CGCTGGCTTT	CCAACGCCCG	GATGACTCAC	4920
TATCAGGCCCT	TGCTTTTGGA	CACGGACCGG	GTCCAGTTCG	GACCGGTGGT	AGCCCTGAAC	4980
CCGGCTACGC	TGCTCCCACT	GCCTGAGGAA	GGGCTGCAAC	ACAACCTGCC	TGATATCCTG	5040
GCCGAAGCCC	ACGGAAACCG	ACCCGACCTA	ACGGACCAGC	CGCTCCCAGA	CGCCGACCAC	5100
ACCTGGTACA	CGGATGGAAG	CAGTCTCTTA	CAAGAGGGAC	AGCGTAAGGC	GGGAGCTGCG	5160
GTGACCACCG	AGACCGAGGT	AATCTGGGCT	AAAGCCCTGC	CAGCCGGGAC	ATCCGCTCAG	5220
CGGGCTGAAC	TGATAGCACT	CACCCAGGCC	CTAAAGATGG	CAGAAGGTAA	GAAGCTAAAT	5280
GTTTATACTG	ATAGCCGTTA	TGCTTTTGCT	ACTGCCCAT	TCCATGGAGA	AATATACAGA	5340
AGGCGTGGGT	TGCTCACATC	AGAAGGCAAA	GAGATCAAAA	ATAAAGACGA	GATCTTGGCC	5400
CTACTAAAAG	CCCTCTTTCT	GCCCCAAAAG	CTTAGCATAA	TCCATTGTCC	AGGACATCAA	5460
AAGGGACACA	GCGCCGAGGC	TAGAGCAAC	CGGATGGCTG	ACCAAGCGGC	CCGAAAGGCA	5520
GCCATCACAG	AGACTCCAGA	CACCTCTACC	CTCCTCATAG	AAAATTTCATC	ACCCTACACC	5580
TCAGAACATT	TTCATTACAC	AGTGACTGAT	ATAAAGGACC	TAACCAAGTT	GGGGGCCATT	5640
TATGATAAAA	CAAAGAAGTA	TTGGGTCTAC	CAAGGAAAAC	CTGTGATGCC	TGACCAGTTT	5700
ACTTTTGAAT	TATTAGACTT	TCTTCAFCAG	CTGACTCACC	TCAGCTTCTC	AAAAATGAAG	5760
GCTCTCCTAG	AGAGAAGCCA	CAGTCCCTAC	TACATGCTGA	ACCGGGATCG	AACACTCAAA	5820
AATATCACTG	AGACCTGCAA	AGCTTGTGCA	CAAGTCAACG	CCAGCAAGTC	TGCCGTAAAT	5880
CAGGGAACCTA	GAGTCCGCGG	GCATCGGCCC	GGCACTCATT	GGGAGATCGA	TTTCACCGAG	5940
ATAAAGCCCG	GATTGTATGG	CTATAAATAT	CTTCTAGTTT	TTATAGATAC	CTTTTCTGGC	6000
TGGATAGAAG	CCTTCCCAAC	CAAGAAAGAA	ACCGCCAAGG	TCGTAACCAA	GAAGCTACTA	6060
GAGGAGATCT	TCCCCAGGTT	CGGCATGCCCT	CAGGTATTGG	GAACGTACAA	TGGGCCTGCC	6120
TTCTGTCTCA	AGGTGAGTCA	GACAGTGGCC	GATCTGTTGG	GGATTGATTG	GAAATTACAT	6180
TGTGCATACA	GACCCCAAAG	CTCAGGCCAG	GTAGAAAAGAA	TGAATAGAAC	CATCAAGGAG	6240
ACTTTAACTA	AATTAACGCT	TGCAACTGGC	TCTAGAGACT	GGGTGCTCCT	ACTCCCCTTA	6300
GCCCTGTACC	GAGCCCGCAA	CACGCCGGGC	CCCCATGGCC	TCACCCCAT	TGAGATCTTA	6360
TATGGGGCAC	CCCCGCCCTT	TGTAAACTTC	CCTGACCCTG	ACATGACAAG	AGTTACTAAC	6420
AGCCCCCTCT	TCCAAGCTCA	ETTACAGGCT	CTCTACTTAG	TCCAGCACGA	AGTCTGGAGA	6480
CCTCTGGCGG	CAGCCTACCA	AGAACAACCTG	GACCGACCGG	TGGTACCTCA	CCCTTACCGA	6540
GTCCGGCGACA	CAGTGTGGGT	CCGCCGACAC	CAGACTAAGA	ACCTAGAACC	TCGCTGGAAA	6600
GGACCTTACA	CAGTCTGTCT	GACCACCCCC	ACCGCCCTCA	AAGTAGACGG	CATCGCAGCT	6660
TGGATACACG	CCGCCCCACGT	GAAGGCTGCC	GACCCCGGGG	GTGGACCATC	CTCTAGACTG	6720
ACATGGCGCG	TTCAACGCTC	TCAAAACCCC	TTAAAAATAA	GGTTAACCCG	CGAGGCCCCC	6780
TAATCCCCCTT	AATTCCTCTG	ATGCTCAGAG	GGGTCAGTAC	TGCTTCGCCC	GGCTCCAGTG	6840
CGGCCCCAGCC	GGCCACCATG	AAAACATTTA	ACATTTCTCA	ACAAGATCTA	GAATTAGTAG	6900
AAGTAGCGAC	AGAGAAGATT	ACAATGCTTT	ATGAGGATAA	TAAACATCAT	GTGGGAGCGG	6960
CAATTCGTAC	GAAAACAGGA	GAAATCATTT	CGGCAGTACA	TATTGAAGCG	TATATAGGAC	7020
GAGTAACGTG	TTGTGCAGAA	GCCATTGCGA	TTGGTAGTGC	AGTTTCGAAT	GGACAAAAGG	7080
ATTTTGACAC	GATTGTAGCT	GTTAGACACC	CTTATTCTGA	CGAAGTAGAT	AGAAGTATTC	7140
GAGTGGTAAG	TCCTTGTGGT	ATGTGTAGGG	AGTTGATTTC	AGACTATGCA	CCAGATTGTT	7200
TTGTGTTAAT	AGAAATGAAT	GGCAAGTTAG	TCAAACTAC	GATTGAAGAA	CTCATTCAC	7260
TCAAAATATAC	CCGAAATTAA	AAGTTTAC	ACCAAGCTTA	TCAATTTC		7308

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Figure 9. FBdelPASAF Sequence

CATATGCGGT	GTGAAATACC	GCACAGATGC	GTAAGGAGAA	AATACCGCAT	CAGGCGCCAT	60
TCGCCATTCA	GGCTGCGCAA	CTGTTGGGAA	GGGCGATCGG	TGCGGGCCTC	TTCGCTATTA	120
CGCCAGCTGG	CGAAAGGGGG	ATGTGCTGCA	AGGCGATTAA	GTTGGGTAAC	GCCAGGGTTT	180
TCCCAATCAC	GACGTTGTAA	AACGACGGCC	AGTGAATTCC	GATTAGTTCA	ATTTGTATAA	240
GACAGGATCT	CAGTAGTCCA	GGCTTTAGTC	CTGACTCAAC	AATACCACCA	GCTAAAACCA	300
CTAGAATACG	AGCCACAATA	AATAAAAGAT	TTTATTTAGT	TTCCAGAAAA	AGGGGGGAAT	360
GAAAGACCCC	ACCAAATTGC	TTAGCCTGAT	AGCCGCAGTA	ACGCCATTTT	GCAAGGCATG	420
GAAAAATACC	AAACCAAGAA	TAGAGAAAGT	CAGATCAAGG	GCGGGTACAC	GAAAACAGCT	480
AACGTTGGGC	CAAACAGGAT	ATCTGCGGTG	AGCAGTTTCG	GCCCCGGCCC	GGGGCCAAGA	540
ACAGATGGTG	ACGCGGGTTC	GGCCCCGGCC	CGGGGCCAAG	AACAGATGGT	CCCCAGATAT	600
AGCCCAACCC	TACAGCAGTT	CTTAAGACCC	ATCAGATGTT	TCCAGGCTCC	CCCAAGGACC	660
TGAAATGACC	CTGTGCCTTA	TTTGAATTAA	CCAATCAGCC	TGCTTCTCGC	TTCTGTTTCG	720
GCGCTTCTGC	TTCCCGAGCT	CTATAAAGA	GCTCACAACC	CCTCACTCGG	CGCGCCAGTC	780
CTCCGATAGA	CTGAGTCGCC	CGGGTACCCG	TGTATCCAAT	AAATCCTCTT	GCTGTTGCAT	840
CCGACTCGTG	GTCTCGCTGT	TCCTTGGGAG	GGTCTCCTCA	GAGTGATTGA	CTACCCGTCT	900
CGGGGGTCTT	TCATTTGGGG	GCTCGTCCGG	GATCTGGAGA	CCCCTGCCCA	GGGACCACCG	960
ACCCACCACC	GGGAGGTAAG	CTGGCCAAGA	TCCTATATGG	GGCACCCCCG	CCCCTTGTAA	1020
ACTTCCCTGA	CCCTGACATG	ACCAGAGTTA	CTAACAGCCC	CTCTCTCCAA	GCTCACTTAC	1080
AGGCTCTCTA	CTTAGTCCAG	CACGAAGTTT	GGAGACCACT	GGCGGCAGCT	TACCAAGAAC	1140
AACTGGACCG	GCCGGTGGTG	CCTCACCCCT	ACCGGGTCGG	CGACACAGTG	TGGGTCCGCC	1200
GACATCAAAAC	CAAGAACCTA	GAACCTCGCT	GGAAAGGACC	TTACACAGTC	CTGCTGACCA	1260
CCCCCACC GC	CCTCAAAGTA	GACGGTATCG	CAGCTTGGAT	ACACGCAGCC	CACGTAAAGG	1320
CGGCCGACAC	CGAGAGTGGG	CCATCCTCTG	GACGGACATG	GCGCGTTCAA	CGCTCTCAAA	1380
ACCCCTCTAA	GATAAGATTA	ACCCGTGGAA	GCCCTTAATA	GTCATGGGAG	TCCTGTTAGG	1440
AGTAGGGATG	GCAGAGAGCC	CCCATCAGGT	CTTTAATGTA	ACCTGGAGAG	TCACCAACCT	1500
GATGACTGGG	CGTACCGCCA	ATGCCACCTC	CCTCCTGGGA	ACTGTACAAG	ATGCCTTCCC	1560
AAAATTATAT	TTTGATCTAT	GTGATCTGGT	CGGAGAGGAG	TGGGACCCCT	CAGACCAGGA	1620
ACCGTATGTC	GGGTATGGCT	GCAAGTACCC	CGCAGGGAGA	CAGCGGACCC	GGACTTTTGA	1680
CTTTTACGTG	TGCCCTGGGC	ATACCGTAAA	GTCCGGGTGT	GGGGGACCAG	GAGAGGGCTA	1740
CTGTGGTAAA	TGGGGGTGTG	AAACCACCGG	ACAGGCTTAC	TGGAAGCCCA	CATCATCGTG	1800
GGACCTAATC	TCCCTTAAGC	GCGGTAACAC	CCCCTGGGAC	ACGGGATGCT	CTAAAGTTGC	1860
CTGTGGCCCC	TGCTCAGACC	TCTCCAAAGT	ATCCAATTCC	TTCCAAGGGG	CTACTCGAGG	1920
GGGCAGATGC	AACCCTCTAG	TCCTAGAATT	CACTGATGCA	GGAAAAAAGG	CTAACTGGGA	1980
CGGGCCCCAA	TCGTGGGGAC	TGAGACTGTA	CCGGACAGGA	ACAGATCCTA	TTACCATGTT	2040
CTCCTCTGAC	CGGCAGCTCC	TTAATGTGGG	ACCCCGAGTC	CCCATAGGGC	CCAACCCAGT	2100
ATTACCCGAC	CAAAGACTCC	CTTCTCACC	AATAGAGATT	GTACCGGCTC	CACAGCCACC	2160
TAGCCCCCTC	AATACAGTT	ACCCCCCTTC	CACTACCAGT	ACACCCCTCA	CCTCCCCTAC	2220
AAGTCCAAGT	GTCCACAGC	CACCCCCAGG	AACTGGAGAT	AGACTACTAG	CTCTAGTCAA	2280
AGGAGCCTAT	CAGGCCTTA	ACCTACCAA	TCCCGACAAG	ACCCAAGAAT	GTTGGCTGTG	2340
CTTAGTGTCG	GGACCTCCTT	ATTACGAAGG	AGTAGCGGTC	GTGGGCATT	ATACCAATCA	2400
TTCCACCGCT	CCGCCCAACT	GTACGGCCAC	TTCCCAACAT	AAGCTTACCC	TATCTGAAGT	2460
GACAGGACAG	GGCCTATGCA	TGGGGGCAGT	ACCTAAAAC	CACCAAGGCT	TATGTAACAC	2520
CACCCAAAGC	GCCGGCTCAG	GATCCTACTA	CCTTGCAGCA	CCCCCCGAA	CAATGTGGGC	2580
TTGCAGCACT	GGATTGACTC	CCTGCTTGTC	CACCACGGTG	CTCAATCTAA	CCACAGATTA	2640
TTGTGTATTA	GGTTGAATCT	GGCCAGAGT	AAATTTACCAC	TCCCCGATT	ATATGTATGG	2700
TCAGCTTGAA	CAGCGTACCA	AATATAAAAG	AGAGCCAGTA	TCATTGACCC	TGGCCCTTCT	2760
ACTAGGAGGA	TTAACCATGG	GAGGGATTGC	AGCTGGAATA	GGGACGGGGA	CCACTGCCTT	2820
AATTAAGAAC	CAGCAGTTTG	AGCAGCTTCA	TGCCGCTATC	CAGACAGACC	TCAACGAAGT	2880
CGAAAAGTCA	ATTACCAACC	TAGAAAAGTC	ACTGACCTCG	TTGTCTGAAG	TAGTCTTACA	2940
GAACCGCAGA	GGCCTAGATT	TGCTATTCTT	AAAGGAGGGA	GGTCTCTGCG	CAGCCCTAAA	3000
AGAAGAATGT	TGTTTTTATG	CAGACCACAC	GGGGCTAGTG	AGAGACAGCA	TGGCCAAATT	3060
AAGAGAAAGG	CTTAATCAGA	GACAAAAACT	ATTTGAGACA	GGCCAAGGAT	GGTTCTGAAG	3120
GCTGTTTAAT	AGATCCCCCT	GGTTTACCAC	CTTAATCTCC	ACCATCATGG	GACCTCTAAT	3180
AGTACTCTTA	CTGATCTTAC	TCTTTGGACC	TTGCATTCTC	AATCGATTAG	TTCAATTTGT	3240
TAAAGACAGG	ATCTCAGTAG	TCCAGGCTTT	AGTCCCTGACT	CAACAATACC	ACCAGCTAAA	3300
GCCTATAGAG	TACGAGCCAT	AGGGCGCCTA	GTGTTGACAA	TTAATCATCG	GCATAGTATA	3360
CGGCATAGTA	TAATACGACT	CACATATAGGA	GGGCCACCAT	GGCCAAGTTG	ACCAGTGCCG	3420
TTCCGTTATT	CACCCGCGCG	GACGTCGCGC	GAGCGGTCGA	GTTCTGGACC	GACCGGCTCG	3480
GGTTCTCCCG	GGACTTCGTG	GAGGACGACT	TCGCCGGTGT	GGTCCGGGAC	GACGTGACCC	3540
TGTTTCATCAG	CGCGGTCCAG	GACCAAGTGG	TGCCGGACAA	CACCCCTGGC	TGGGTGTGGG	3600
TGCGCGGCCCT	GGACGAGCTG	TACGCCGAGT	GGTCCGAGGT	CGTCTCCACG	AACTTCCGGG	3660
ACGCCTCCGG	GCCGGCCATG	ACCGAGATCG	CGGAGCAGCC	GTGGGGGCGG	GAGTTCTGCCC	3720
TGCGCGACCC	GGCCGGCAAC	TGCGTGCACT	TCGTGGCCGA	GGAGCAGGAC	TGANNNNCGG	3780
ACCGGTCGAC	TTGTTAACTT	GTTTATTGCA	GCTTATAATG	GTTACAAATA	AAGCAATAGC	3840
ATCACAAAAT	TACAAATAA	AGCATTTTTT	TCACGTGATT	CTAGTTGTGG	TTTGTCCTAAA	3900
CTCATCAATG	TATCTTATCA	TGTCTGGATC	CAGATCTGGG	CCCATGCGGC	CGCGGATCGA	3960
TNNNNACATG	TGAGCAAAAG	GCCAGCAAAA	GGCCAGGAAC	CGTAAAAAAG	CCGCGTTGCT	4020
GGCGTTTTTC	CATAGGCTCC	GCCCCCTGGA	CGAGCATCAC	AAAAATCGAC	GCTCAAGTCA	4080

Figure 9. FBdelPASAF Sequence

2

GAGGTGGCGA	AACCCGACAG	GACTATAAAG	ATACCAGGCG	TTTCCCCCTG	GAAGCTCCCT	4140
CGTGCGCTCT	CCTGTTCCGA	CCCTGCCGCT	TACCGGATAC	CTGTCCGCCT	TTCTCCCTTC	4200
GGGAAGCGTG	GCGCTTTCTC	AATGCTCAGC	CTGTAGGTAT	CTCAGTTCGG	TGTAGGTCGT	4260
TCGCTCCAAG	CTGGGCTGTG	TGCACGAACC	CCCCGTTCAG	CCCGACCGCT	GCGCCTTATC	4320
CGGTAACTAT	CGTCTTGAGT	CCAACCCGGT	AAGACACGAC	TTATCGCCAC	TGGCAGCAGC	4380
CACCTGGTAAC	AGGATTAGCA	GAGCGAGGTA	TGTAGGCGGT	GCTACAGAGT	TCTTGAAGTG	4440
GTGGCCTAAC	TACGGCTACA	CTAGAAGGAC	AGTATTTGGT	ATCTGCGCTC	TGCTGAAGCC	4500
AGTTACCTTC	GGAAAAAGAG	TTGGTAGCTC	TTGATCCGGC	AAACAAACCA	CCGCTGGTAG	4560
CGGTGGTTTT	TTTGTTTTGA	AGCAGCAGAT	TACGCGCAGA	AAAAAAGGAT	CTCAAGAAGA	4620
TCCTTTGATC	TTTCTTACGG	GGTCTGACGC	TCAGTGGAAAC	GAAAACTCAC	GTTAAGGGAT	4680
TTTGGTCATG	AGATTATCAA	AAAGGATCTT	CACCTAGATC	CTTTTAAATT	AAAAATGAAG	4740
TTTTAAATCA	ATCTAAAGTA	TATATGAGTA	AACTTGGTCT	GACAGTTACC	AATGCTTAAT	4800
CAGTGAGGCA	CCTATCTCAG	CGATCTGTCT	ATTCGTTCA	TCCATAGTTG	CCTGACTCCC	4860
CGTCGTGTAG	ATAACTACGA	TACGGGAGGG	CTTACCATCT	GGCCCCAGTG	CTGCAATGAT	4920
ACCGCGAGAC	CCACGCTCAC	CGGCTCCAGA	TTTATCAGCA	ATAAACACAGC	CAGCCGGAAG	4980
GGCCGAGCGC	AGAAGTGGTC	CTGCAACTTT	ATCCGCCCTCC	ATCCAGTCTA	TTAATTGTTG	5040
CCGGGAAGCT	AGAGTAAGTA	GTTTCGCCAGT	TAATAGTTTG	CGCAACGTTG	TTGCCATTGC	5100
TACAGGCATC	GTGGTGTCAC	GCTCGTCGTT	TGGTATGGCT	TCATTCAGCT	CCGGTTCCCA	5160
ACGATCAAGG	CGAGTTACAT	GATCCCCCAT	GTTGTGCAAA	AAAGCGGTTA	GCTCCTTCGG	5220
TCCTCCGATC	GTTGTCAGAA	GTAAGTTGGC	CGCAGTGTTA	TCACTCATGG	TTATGGCAGC	5280
ACTGCATAAT	TCTCTTACTG	TCATGCCATC	CGTAAGATGC	TTTTCTGTGA	CTGGTGAGTA	5340
CTCAACCAAG	TCATTCTGAG	AATAGTGTAT	GCGGCGACCG	AGTTGCTCTT	GCCCCGCGTC	5400
AATACGGGAT	AATACCGCGC	CACATAGCAG	AACTTTAAAA	GTGCTCATCA	TTGGAAAACG	5460
TTCTTCGGGG	CGAAAACTCT	CAAGGATCTT	ACCGCTGTTG	AGATCCAGTT	CGATGTAACC	5520
CACCTCGTGCA	CCCAACTGAT	CTTCAGCATC	TTTTACTTTC	ACCAGCGTTT	CTGGGTGAGC	5580
AAAAACAGGA	AGGCAAAATG	CCGCAAAAAA	GGGAATRAAG	GCGACACGGA	AATGTTGAAT	5640
ACTCATACTC	TTCTTTTTTC	AATATTATTG	AAGCATTAT	CAGGGTTATT	GTCTCATGAG	5700
CGGATACATA	TTTGAATGTA	TTTAGAAAAA	TAAACAAATA	GGGGTTCCGC	GCACATTTC	5760
CCGAAAAGTG	CCACCTGACG	TCTAAGAAAC	CATTATTATC	ATGACATTAA	CCTATAAAAA	5820
TAGGCGTATC	ACGAGGCCCT	TTCGTCTCGC	GCGTTTCGGT	GATGACGGTG	AAAACCTCTG	5880
ACACATGCAG	CTCCCGGAGA	CGGTCACAGC	TTGTCTGTAA	GCGGATGCCG	GGAGCAGACA	5940
AGCCCGTCAG	GGCGCGTCAG	CGGGTGTGG	CGGGTGTGG	GGCTGGCTTA	ACTATGCGGC	6000
ATCAGAGCAG	ATTGTACTGA	GAGTGCAC				6028

Figure 10. FBdelPMOSAF Sequence

1

CATATGCGGT	GTGAAATACC	GCACAGATGC	GTAAGGAGAA	AATACCGCAT	CAGGCGCCAT	60
TCGCCATTCA	GGCTGCGCAA	CTGTTGGGAA	GGGCGATCGG	TGCGGGCCCTC	TTCGCTATTA	120
CGCCAGCTGG	CGAAAGGGGG	ATGTGCTGCA	AGGCGATTAA	GTTGGGTAAC	GCCAGGGTTT	180
TCCCAGTCAC	GACGTTGTAA	AACGACGGCC	AGTGAATTCC	GATTAGTTCA	ATTTGTTAAA	240
GACAGGATCT	CAGTAGTCCA	GGCTTTAGTC	CTGACTCAAC	AATACCACCA	GCTAAAACCA	300
CTAGAATACG	AGCCACAATA	AATAAAAGAT	TTTATTTAGT	TTCCAGAAAA	AGGGGGGAAT	360
GAAAGACCCC	ACCAAATTGC	TTAGCCTGAT	AGCCGCAGTA	ACGCCATTTT	GCAAGGCATG	420
GAAAAATACC	AAACCAAGAA	TAGAGAAGTT	CAGATCAAGG	GCGGGTACAC	GAAAACAGCT	480
AACGTTGGGC	CAACAGGAT	ATCTGCCGTG	AGCATGTTTCG	GCCCCGGCCC	GGGGCCAAAGA	540
ACAGATGGTC	ACCGCGGTTT	GGCCCCGGCC	CGGGGCCAAG	AACAGATGGT	CCCCAGATAT	600
GGCCCCAACC	TCAGCAGTTT	CTTAAGACCC	ATCAGATGTT	TCCAGGCTCC	CCCAAGGACC	660
TGAAATGACC	CTGTGCCTTA	TTTGAATTAA	CCAATCAGCC	TGCTTCTCGC	TTCTGTTCCG	720
GCGCTTCTGC	TTCCCGAGCT	GCTATAAAGA	GCTACAACCC	CCTCACTCGG	CGCGCCAGTC	780
CTCCGATAGA	CTGAGTCGCC	CGGGTACCCG	TGTATCCAAT	AAATCCTCTT	GCTGTTGCAT	840
CCGACTCGTG	GTCTCGCTGT	TCCTTGGGAG	GGTCTCCTCA	GAGTGATTGA	CTACCCGCTT	900
CGGGGTCTTT	TCATTTGGGG	GCTCGTCCGG	GATCTGGAGA	CCCCTGCCCA	GGGACCACCG	960
ACCCACCACC	GGGAGGTAAG	CTGGCCAAGA	TCTTATATGG	GGCACCCTCG	CCCCTTGTA	1020
ACTTCCCTGA	CCCTGACATG	ACAAGAGTTA	CTAACAGCCC	CTCTCTCCAA	GCTCACTTAC	1080
AGGCTCTCTA	CTTAGTCCAG	CACGAAGTCT	GGAGACCTCT	GGCGGCAGCC	TACCAAGAAC	1140
AACCTGACCG	ACCGTGGTA	CCTCACCTTT	ACCGAGTCGG	CGACACAGTG	TGGGTCCGCG	1200
GACACCAGAC	TAAGAACCTA	GAACCTCGCT	GGAAAGGACC	TTACACAGTC	CTGCTGACCA	1260
CCCCCACC	CCTCAAAGTA	GACGGCATCG	CAGCTTGGAT	ACACGCGGCC	CACGTGAAGG	1320
CTGCCGACCC	CGGGGGTGGA	CCATCCTCTA	GACTGACATG	GCGCGTTCAA	CGCTCTCAA	1380
ACCGCTTAAA	ACATTAAGTTA	ACCCGCGAGG	CCCCCTAATC	CCCTTAATTTC	TTCTGATGCT	1440
CAGAGGGGTC	AGTACTGCTT	CGCCCCGGCTC	CAGTCTCAT	CAAGTCTATA	ATATCACCTG	1500
GGAGGTAACC	AATGGAGATC	GGGAGACGGT	ATGGGCAACT	TCTGGCAACC	ACCCTCTGTG	1560
GACCTGGTGG	CCTGACCTTA	CCCCAGATT	ATGTATGTTA	GCCCACCATG	GACCATCTTA	1620
TTGGGGGGCTA	GAATATCAAT	CCCCTTTTTC	TTCTCCCCCG	GGGCCCCCTT	GTTGCTCAGG	1680
GGGCAGCAGC	CCAGGCTGTT	CCAGAGACTG	CGAAGAACCT	TTAACCTCCC	TCACCCCTCG	1740
GTGCAACACT	GCCTGGAACA	GACTCAAGCT	AGACCAGACA	ACTCATAAAT	CAAAATGAGGG	1800
ATTTTATGTT	TGCCCCGGGC	CCCACCGCCC	CCAGGAATCC	AAGTCATGTG	GGGGTCCAGA	1860
CTCCTTCTAC	TGTGCTTATT	GGGGCTGTGA	GACAAACGGT	AGAGCTTACT	GGAAGCCCTC	1920
CTCATCATGG	GATTTTCATCA	CAGTAAACAA	CAATCTCACC	TCTGACCAGG	CTGTCCAGGT	1980
ATGCAAAAGAT	AATAAGTGGT	GCAACCCCTT	AGTTATTCGG	TTTACAGACG	CCGGGAGACG	2040
GGTTACTTCC	TGGACCACAG	GACATTACTG	GGGCTTACGT	TTGTATGTCT	CCGGACAAGA	2100
TCCAGGGGTT	ACATTTGGGA	TCCGACTCAG	ATACCAAAT	CTAGGACCCC	GCGTCCCAAT	2160
AGGGCCAAAC	CCCGTTCTGG	CAGACCAACA	GCCACTCTCC	AAGCCCAAAC	CTGTTAAGTC	2220
GCCTTCAATC	ACCAAACCAAC	CCAGTGGGAC	TCCTCTCTCC	CCTACCCAAC	TTCCACCGGC	2280
GGGAACGGAA	AATAGGCTGC	TAAACTTAGT	AGACGGAGCC	TACCAAGCCC	TCAACCTCAC	2340
CAGTCTGTAC	AAAACCCAAG	AGTGCTGGTT	GTGTCTAGTA	GCGGGACCCC	CCTACTACGA	2400
AGGGGTTGGC	GTCTTGGGTA	CCTACTCCAA	CCATACCTCT	GCTCCAGCCA	ACTGCTCCGT	2460
GGCCTCCCAA	CACAAGTTGA	CCCTGTCCGA	AGTGACCGGA	CAGGGACTCT	GCATAGGAGC	2520
AGTTCCCAAA	ACACATCAGG	CCCTATGTAA	TACCACCCAG	ACAAGCAGTC	GAGGGTCCCTA	2580
TTACTTAGTT	CCCCCTACAG	GTACCATGTG	GGCTTGTAGT	ACCGGGCTTA	CTCCATGCAT	2640
CTCCACCAAC	ATACTGAACC	TTACCACTGA	TTATTGTGTT	CTTGTCTGAAC	TCTGGCCAAG	2700
AGTCACCTAT	CATTCCCCCA	GCTATGTTTA	CGGCCTGTTT	GAGAGATCCA	ACCGACACAA	2760
AAGAGAACCG	GTGTGCTTAA	CCCTGGCCCT	ATTATTGGGT	GGACTAACCA	TGGGGGGAAT	2820
TGCCGTGGGA	ATAGGAACAG	GGACTACTGC	TCTAATGGCC	ACTCAGCAAT	TCCAGCAGCT	2880
CCAAGCCGCA	GTACAGGATG	ATCTCAGGGA	GGTTGAAAAA	TCAATCTCTA	ACCTAGAAAA	2940
GTCTCTCACT	TCCCTGTCTG	AAGTTGTCCT	ACAGAATCGA	AGGGGCCCTAG	ACTTGTTATT	3000
TCTAAAAGAA	GGAGGGCTGT	GTGCTGCTCT	AAAAGAAGAA	TGTTGCTTCT	ATGCGGACCA	3060
CACAGGACTA	GTGAGAGACA	GCATGGCCAA	ATTGAGAGAG	AGGCTTAATC	AGAGACAGAA	3120
ACTGTTTGTAG	TCAACTCAAG	GATGGTTTGA	GGGACTGTTT	AACAGATCCC	CTTGTTTATC	3180
CACCTTGATA	TCTACCATT	TGGGACCCCT	CATTGTACTC	CTAATGATTT	TGCTCTTCGG	3240
ACCTGCAATT	CTTAATCGAT	TAGTTCAATT	TGTTAAAGAC	AGGATCTCAG	TAGTCCAGGC	3300
TTAGTCTCTG	ACTCAACAAT	ACCACCAGCT	AAAGCCTATA	GAGTACGAGC	CATAGGGCGC	3360
CTAGTGTGTA	CAATTAATCA	TCGGCATAGT	ATACGCATA	GTATAATACG	ACTCACTATA	3420
GGAGGGCCAC	CATGGCCAA	TTGACCAGTG	CCGTTCCGGT	GCTCACCAGC	CGCGACGTCG	3480
CCGGAGCCGT	CGAGTTCTGG	ACCGACCGGC	TCCGGTTCTC	CCGGGACTTC	GTGGAGGACG	3540
ACTTCGCCGG	TGTGGTCCGG	GACGACGTGA	CCCTGTTTCT	CAGCGCGGTC	CAGGACCAGG	3600
TGGTGCCGGA	CAACACCTGT	GCCTGGGTGT	GGGTGCGCGG	CCTGGACGAG	CTGTACGCCG	3660
AGTGGTCGGA	GGTGTGTCTC	ACGAACTTCC	GGGACGCTC	CGGGCCGGCC	ATGACCGAGA	3720
TCGGCGAGCA	GCCGTGGGGG	CGGGAGTTCG	CCCTGCGCGA	CCCGCCGGCC	AACCTGCGTG	3780
ACTTCGTGGC	CGAGGAGCAG	GACTGANNNN	CGGACCGGTC	GACTTGTTAA	CTTGTTTATT	3840
GCAGCTTATA	ATTGTTACAA	ATAAAGCAAT	AGCATCACAA	ATTTACACAA	TAAAGCATTT	3900
TTTTCACTGC	ATGCTAGTTG	TGGTTTGTCC	AAACTCATCA	ATGTATCTTA	TCATGTCTGG	3960
ATCCAGATCT	GGGCCCATGC	GGCCGCGGAT	CGATNNNNAC	ATGTGAGCAA	AAGGCCAGCA	4020
AAAGGCCAGG	AACCGTAAAA	AGGCCGCGTT	GCTGGCGTTT	TTCCATAGGC	TCCGCCCCCC	4080

Figure 10. FBdelPMOSAF Sequence

TGACGAGCAT	CACAAAAATC	GACGCTCAAG	TCAGAGGTGG	CGAAACCCGA	CAGGACTATA	4140
AAGATAACAG	GCGTTTCCCC	CTGGAAGCTC	CCTCGTGC GC	TCTCCTGTTC	CGACCCTGCC	4200
GCTTACCGGA	TACCTGTCCG	CCTTCTCCCC	TTCGGGAAGC	GTGGCGCTTT	CTCAATGCTC	4260
ACGCTGTAGG	TATCTCAGTT	CGGTGTAGGT	CGTTCGCTCC	AAGCTGGGCT	GTGTGCACGA	4320
ACCCCCGTT	CAGCCCGACC	GCTGCGCCTT	ATCCGGTAAC	TATCGTCTTG	AGTCCAACCC	4380
GGTAAGACAC	GACTTATCGC	CACTGGCAGC	AGCCACTGGT	AACAGGATTA	GCAGAGCGAG	4440
GTATGTAGGC	GGTGCTACAG	AGTTCTTGAA	GTGGTGGCCT	AACACGGCT	ACACTAGAAG	4500
GACAGTATTT	GGTATCTGCG	CTCTGCTGAA	GCCAGTTACC	TTCGGA AAAA	GAGTTGGTAG	4560
CTCTTGATCC	GGCAAAACAA	CCACCGCTGG	TAGCGGTGGT	TTTTTTGTTC	GCAAGCAGCA	4620
GATTACGCGC	AGAAAAAAG	GATCTCAAGA	AGATCCTTTG	ATCTTTTCTA	CGGGGTCTGA	4680
CGCTCAGTGG	AACGAAAACT	CACGTTAAGG	GATTTTGGTC	ATGAGATTAT	CAAAAAGGAT	4740
CTTCACCTAG	ATCCTTTTAA	ATTAAAAATG	AAGTTTTTAA	TCAATCTAAA	GTATATATGA	4800
GTAACCTTGG	TCTGACAGTT	ACCAATGCTT	AATCAGTGAG	GCACCTATCT	CAGCGATCTG	4860
TCTATTTCTG	TCATCCATAG	TTGCCTGACT	CCCCGTCTGT	TAGATAACTA	CGATACGGGA	4920
GGGCTTACCA	TCTGGCCCCA	GTGCTGCAAT	GATACCCGCA	GACCCACGCT	CACCGGCTCC	4980
AGATTATCA	GCAATAAACC	AGCCAGCCGG	AAGGGCCGAG	CGCAGAAGTG	GTCTTGCAAC	5040
TTTATCCGCC	TCCATCCAGT	CTATTAATTG	TTGCCGGGAA	GCTAGAGTAA	GTAGTTCGCC	5100
AGTTAATAGT	TTGCGCAACG	TTGTTGCCAT	TGCTACAGGC	ATCGTGGTGT	CACGCTCGTC	5160
GTTTGGTATG	GCTTCATTCA	GCTCCGGTTC	CCAACGATCA	AGGCGAGTTA	CATGATCCCC	5220
CATGTTGTGC	AAAAAAGCGG	TTAGCTCCTT	CGGTCTCTCG	ATCGTTGTCA	GAAGTAAGTT	5280
GGCCGCGAGT	TTATCACTCA	TGGTTATGGC	AGCACTGCAT	AATTCCTCTA	CTGTCTATGCC	5340
ATCCGTAAGA	TGCTTTTCTG	TGACTGGTGA	GTA CTCAACC	AAGTCATTCT	GAGAATAGTG	5400
TATGCGGCGA	CCGAGTTGCT	CTTGCCCGGC	GTCAATACGG	GATAATACCG	CGCCACATAG	5460
CAGAACTTTA	AAAGTGCTCA	TCATTGGA AA	ACGTTCTTCG	GGGCGAAAAC	TCTCAAGGAT	5520
CTTACCGCTG	TTGAGATCCA	GTTCGATGTA	ACCCACTCGT	GCACCCAAC	GATCTTCAGC	5580
ATCTTTTACT	TTCAACAGCG	TTTCTGGGTG	AGCAAAAACA	GGAAGGCAAA	ATGCCGCAAA	5640
AAAGGGAATA	AGGGCGACAC	GGAAATGTTG	AATACTCATA	CTCTTCCTTT	TTCAATATTA	5700
TTGAAGCATT	TATCAGGGTT	ATTGTCTCAT	GAGCGGATAC	ATATTTGAAT	GTATTTAGAA	5760
AAATAAACAA	ATAGGGGTTT	CGCGCACATT	TCCCCGAAAA	GTGCCACCTG	ACGTCTAAGA	5820
AACCATTATT	ATCATGACAT	TAACCTATAA	AAATAGGCGT	ATCACGAGGC	CCTTTCGTCT	5880
CGCGCGTTTC	GGTGATGACG	GTGAAAACCT	CTGACACATG	CAGCTCCCGG	AGACGGTCAC	5940
AGCTTGCTCTG	TAAGCGGATG	CCGGGAGCAG	ACAAGCCCCG	CAGGGCGCGT	CAGCGGGTGT	6000
TGGCGGGTGT	CGGGGCTGGC	TTAACTATGC	GGCATCAGAG	CAGATTGTAC	TGAGAGTGCA	6060
C						6061

Figure 11. FBdelPGASAF Sequence

1

CATATGCGGT	GTGAAATACC	GCACAGATGC	GTAAGGAGAA	AATACCGCAT	CAGGCGCCAT	60
TCGCCATTCA	GGCTGCGCAA	CTGTTGGGAA	GGCGATCGG	TGCGGGCCTC	TTCGCTATTA	120
CGCCAGCTGG	CGAAAGGGGG	ATGTGCTGCA	AGGCGATTAA	GTTGGGTAAC	GCCAGGGTTT	180
TCCCAGTCAC	GACGTTGTAA	AACGACGGCC	AGTGAATTCC	GATTAGTTCA	ATTTGTTAAA	240
GACGAGTCT	CAGTAGTCCA	GGCTTTAGTC	CTGACTCAAC	AATACCACCA	GCTAAAACCA	300
CTAGAATACG	AGCCACAATA	AATAAAGAT	TTTATTTAGT	TTCCAGAAAA	AGGGGGGAAT	360
GAAAGACCCC	ACCAAAATGC	TTAGCCTGAT	AGCCGCAGTA	ACGCCATTTT	GCAAGGCATG	420
GAAAAATACC	AAACCAAGAA	TAGAGAAGTT	CAGATCAAGG	GCGGGTACAC	GAAAACAGCT	480
AACGTTGGGC	CAAAACAGGAT	ATCTGCGGTG	AGCAGTTTCG	GCCCCGGCCC	GGGGCCAAGA	540
ACAGATGGTC	ACCGCGGTTC	GGCCCCGGCC	CGGGGCCAAG	AACAGATGGT	CCCCAGATAT	600
GGCCCAACCC	TCAGCAGTTT	CTTAAGACCC	ATCAGATGTT	TCCAGGCTCC	CCCAAGGACC	660
TGAAATGACC	CTGTGCCTTA	TTTGAATTAA	CCAATCAGCC	TGCTTCTCGC	TTCTGTTCGC	720
GCGCTTCTGC	TTCCCGAGCT	CTATAAAGA	GCTCACAACC	CCTCACTCGG	CGCGCCAGTC	780
CTCCGATAGA	CTGAGTCGCC	CGGGTACCCG	TGTATCCAAT	AAATCCTCTT	GCTGTTGCAT	840
CCGACTCGTG	GTCTCGCTGT	TCCTTGGGAG	GGTCTCCTCA	GAGTGAATTA	CTACCCGCTC	900
CGGGGGTCTT	TCATTTGGGG	GCTCGTCCGG	GATCTGGAGA	CCCTGCCCCA	GGGACCACCG	960
ACCCACCACC	GGGAGGTAAG	CTGGCCAAGA	TCCCTAAGGT	ACTCGGGTCA	GACAATGGCC	1020
CGGCCTTTGT	TGCTCAGGTA	AGTCAGGGAC	TGGCCACTCA	ACTGGGGATA	AATTGGAAGT	1080
TACATTGTGC	GCTATAGACC	CAGAGCTCAG	GTCAAGGTAGA	AAGAATGAAC	AGAACAATTA	1140
AAGAGACCTT	GACCAAAATTA	GCCTTAGAGA	CCGGTGGAAA	AGACTGGGTG	ACCCCTCCTT	1200
CCTTAGCGCT	GCTTAGGGCC	AGGAATACCC	CTGGCCGGTT	TGGTTTAACT	CCTTATGAAA	1260
TTCTCTATGG	AGGACCACCC	CCCATACTTG	AGTCTGGAGA	AACCTTGGGT	CCCGATGATA	1320
GATTTTCTCC	TGCTCTATTT	ACTCACTTAA	AGGCTTTAGA	AATTGTAAGG	ACCCAATCTT	1380
GGGACCAGAT	CAAAGAGGTG	TATAAGCCTG	GTACCGTAAC	AATCCCTCAC	CCGTTCAGG	1440
TCGGGGATCA	AGTGCTTGTC	AGACGCCATC	GACCCAGCAG	CCTTGAGCCT	CGGTGGAAAG	1500
GCCCATACCT	GGTGTGTGTC	ACTACCCCGA	CCGCGGTAAA	AGTCGATGGT	ATTGCTGCCT	1560
GGGTCCATGC	TTCTCACCTC	AAACCTGCAC	CACCTTCGCG	ACCAGATGAG	TCCTGGGAGC	1620
TGGAAGAAGAC	TGATCATCCT	CTTAAGCTGC	GTATTCGGCG	GCGCGGGGAC	GAGTCTGCAA	1680
AATAAGAACC	CCCACCAGCC	CATGACCCCT	ACTTGGCAGG	TACTGTCCCA	AACTGGAGAC	1740
GTGTCTGGG	ATACAAAGGC	AGTCCAGCCC	CCTTGGACTT	GGTGGCCAC	ACTTAAACCT	1800
GATGTATGTG	CCTTGGCGGC	TAGTCTTGAG	TCCTGGGATA	TCCCGGGAAC	CGATGTCTCG	1860
TCCTCTAAAC	GAGTCAGACC	TCCGGACTCA	GACTATATCG	CCGCTTATAA	GCAATCACC	1920
TGGGGAGCCA	TAGGCTGCAG	CTACCCTCGG	GCTAGGACTA	GAATGGCAAG	CTCTACCTTC	1980
TACGTATGTC	CCCGGGATGG	CCGGACCCCT	TCAGAAGCTA	GAAGGTGCGG	GGGGCTAGAA	2040
TCCTTATACT	GTAAGAAGATG	GGATTGTGAG	ACCACGGGGA	CCGGTTATTG	GCTATCTAAA	2100
TCCTCAAAAG	ACCTCATAAC	TGTAAGATGG	GACCAAAATA	GCGAATGGAC	TCAAAAATTT	2160
CAACAGTGTG	ACCAGACCGG	CTGGTGTAAC	CCCCTTAAAA	TAGATTTTCA	AGACAAAGGA	2220
AAATTATCCA	AGGACTGGAT	AACGGGAAAA	ACCTGGGGAT	TAAGATTCTA	TGTGTCTGGA	2280
CATCCAGGCG	TACAGTTCAC	CATTCCGTTA	AAAATCACCA	ACATGCCAGC	TGTGGCAGTA	2340
GGTCTCTGAC	TCCGCTTTGT	GGAACAAGGA	CCTCCTAGAA	CGTCCCTCGC	TCTCCCACTT	2400
CCTCTTCCCC	CAAGGGAAGC	GCCACCGCCA	TCTCTCCCCG	ACTCTAACTC	CACAGCCCTG	2460
GCGACTAGTG	CACAAACTCC	CACGGTGAGA	AAAACAATTG	TTACCCTAAA	CACCTCCGCT	2520
CCCAACACAG	CGCAGACACT	TTTTGATCTT	GTCCAGGGGG	CCTTCCTAAC	CTTAAATGCT	2580
ACCAACCCAG	GGGCCACTGA	GTCTTGCTGG	CTTTGTTTGG	CCATGGGGCC	CCCTTATTAT	2640
GAAGCAATAG	CCTCATCAGG	AGAGGTGCGC	TACTCCACCG	ACCTTGACCG	GTGCGCTGG	2700
GGGACCCAAG	GAAAGCTCAC	CCTCACTGAG	GTCTCAGGAC	ACGGGTGTGT	CATAGGAAAG	2760
GTGCCCTTTA	CCCATCAGCA	TCTCTGCAAT	CAGACCCCTAT	CCATCAATTC	CTCCGGAGAC	2820
CATCAGTATC	TGCTCCCCTC	CAACCATAGC	TGGTGGGCTT	GCAGCACTGG	CCTCACCCTT	2880
TGCCTCTCCA	CCTCAGTTTT	TAATCAGACT	AGAGATTTCT	GTATCCAGGT	CCAGCTGATT	2940
CCTCGCATCT	ATTACTATCC	TGAAGAAGTT	TTGTTACAGG	CCTATGACAA	TTCTCACCCT	3000
AGGACTAAAA	GAGAGGCTGT	CTCACTTACC	CTAGCTGTTT	TACTGGGGTT	GGGAATCAGG	3060
GCGGGAATAG	GTACTGGTTC	AACTGCCCTTA	ATTAAAGGAC	CTATAGACCT	CCAGCAAGGC	3120
CTGACAAGCC	TCCAGATCGC	CATAGATGCT	GACCTCCGGG	CCCTCCAAGA	CTCAGTCAGC	3180
AAGTTAGAGG	ACTCACTGAC	TTCCCTGTCC	GAGGTAGTGC	TCCAAAATAG	GAGAGGCCCT	3240
GACTTGCTGT	TTCTAAAAGA	AGGTGGCCTC	TGTGCGGGCC	TAAAGGAAGA	GTGCTGTTTT	3300
TACATAGACC	ACTCAGGTGC	AGTACGGGAC	TCCATGAAAA	AACTCAAAGA	AAAACCTGGAT	3360
AAAAGACAGT	TAGAGCGCCA	GAAAAGCCAA	AACTGGTATG	AAGGATGGTT	CAATAACTCC	3420
CCTTGGTTCA	CTACCCTGCT	ATCAACCATC	GCTGGGCCCC	TATTACTCCT	CCTTCTGTTG	3480
CTCATCCTCG	GGCCATGCAT	CATCAATCGA	TTAGTTCAAT	TTGTTAAAGA	CAGGATCTCA	3540
GTAGTCCAGG	CTTTAGTCCT	GACTCAACAA	TACCACCAGC	TAAAGCCTAT	AGAGTACGAG	3600
CCATAGGGCG	CCTAGTGTGG	ACAATTAATC	ATCGGCATAG	TATACGGCAT	AGTATAATAC	3660
GACTCACTAT	AGGAGGGCCA	CCATGGCCAA	GTTGACCAAT	GCCGTTCGGG	TGCTCACC GC	3720
GCGCGACGTC	CCCGGAGCGG	TCGAGTTCTG	GACCGACCGG	CTCGGGTTCT	CCCGGGACTT	3780
CGTGGAGGAC	GACTTCGCGG	GTGTGGTCCG	GGACGACGTG	ACCCGTGTTCA	TCAGCGCGGT	3840
CCAGGACCAG	GTGGTGCCGG	ACAACACCCT	GGCTTGGGTG	TGGGTGCGCG	GCCTGGACGA	3900
GCTGTACGCC	GAGTGGTCGG	AGGTCGTGTC	CACGAACCTC	CGGGACGCCT	CCGGGCGGGC	3960
CATGACCGAG	ATCGGCGAGC	AGCCGTGGGG	GCGGGAGTTC	GCCCTGCGCG	ACCCGCGCGG	4020
CAACTGCGTG	CACCTTCGTGG	CCGAGGAGCA	GGACTGANNN	NCGGACCGGT	CGACTTGTTA	4080

Figure 11. FBdelPGASAF Sequence

2

ACTTGTATTAT	TGCAGCTTAT	AATGGTTACA	AATAAGCAA	TAGCATCACA	AATTTACAA	4140
ATAAAGCATT	TTTTTCACTG	CATTCTAGTT	GTGGTTTGTC	CAAACATCATC	AATGTATCTT	4200
ATCATGTCTG	GATCCAGATC	TGGGCCCATG	CGGCCGCGGA	TCGATNNMNA	CATGTAGCA	4260
AAAGGCCAGC	AAAAGGCCAG	GAACCGTAAA	AAGGCCGCGT	TGCTGGCGTT	TTCCATAGG	4320
CTCCGCCCCC	CTGACGAGCA	TCACAAAAAT	CGACGCTCAA	GTGAGAGGTG	GCGAAACCCG	4380
ACAGGACTAT	AAAGATACCA	GGCGTTTCCC	CCTGGAAGCT	CCCTCGTGCG	CTCTCCTGTT	4440
CCGACCTGTC	CGCTTACCGG	ATACCTGTCC	GCCTTTCTCC	CTTCGGGAAG	CGTGGCGCTT	4500
TCTCAATGCT	CACGCTGTAG	GTATCTCAGT	TCGGTGTAGG	TCGTTGCTGC	CAAGCTGGGC	4560
TGTGTGCAG	AACCCCCCGT	TCAGCCCGAC	CGCTGCGCCT	TATCCGGTAA	CTATCGTCTT	4620
GAGTCCAAAC	CGGTAAGACA	CGACTTATCG	CCACTGGCAG	CAGCCACTGG	TAACAGGATT	4680
AGCAGAGCGA	GGTATGTAGG	CGGTGCTACA	GAGTTCTTGA	AGTGGTGGCC	TAACACGGC	4740
TACACTAGAA	GGACAGTATT	TGGTATCTGC	GCTCTGCTGA	AGCCAGTTAC	CTTCGGAAAA	4800
AGAGTTGGTA	GCTCTTGATC	CGGCAAAACA	ACCACCGCTG	GTAGCGGTGG	TTTTTTTGT	4860
TGCAAGCAGC	AGATTACGCG	CAGAAAAAAA	GGATCTCAAG	AAGATCCTTT	GATCTTTTCT	4920
ACGGGGTCTG	ACGCTCAGTG	GAACGAAAAC	TCACGTTAAG	GGATTTTGGT	CATGAGATTA	4980
TCAAAAAGGA	TCTTCACCTA	GATCCTTTTA	AATTAAAAAT	GAAGTTTAA	ATCAATCTAA	5040
AGTATATATG	AGTAAACTTG	GTCTGACAGT	TACCAATGCT	TAATCAGTGA	GGCACCTATC	5100
TCAGCGATCT	GTCTATTTTCG	TTCATCCATA	GTTGCCTGAC	TCCCCGTCGT	GTAGATAACT	5160
ACGATACGGG	AGGGCTTACC	ATCTGGCCCC	AGTGCTGCAA	TGATACCGCG	AGACCCACGC	5220
TCACCGGCTC	CAGATTTATC	AGCAATAAAC	CAGCCAGCCG	GAAGGGCCGA	GCGCAGAAGT	5280
GGTCTTGCAA	CTTTATCCGC	CTCCATCCAG	TCTATTAATT	GTTGCCGGGA	AGCTAGAGTA	5340
AGTAGTTTCG	CAGTTAATAG	TTTGCGCAAC	GTTGTGCGCA	TTGCTACAGG	CATCGTGGTG	5400
TCACGCTCGT	CGTTTGGTAT	GGCTTCATTC	AGCTCCGGTT	CCCAACGATC	AAGGCGAGTT	5460
ACATGATCCC	CCATGTTGTG	CAAAAAAGCG	GTTAGCTCCT	TCGGTCTCTC	GATCGTTGTC	5520
AGAAGTAAGT	TGGCCGCGAGT	GTTATCACTC	ATGGTTATGG	CAGCACTGCA	TAATTCTCTT	5580
ACTGTCAATG	CATCCGTAAG	ATGCTTTTCT	GTGACTGGTG	AGTACTCAAC	CAAGTCATTC	5640
TGAGAATAGT	GTATGCGGCG	ACCGAGTTGC	TCTTGCCCGG	CGTCAATACG	GGATAATACC	5700
GCGCCACATA	GCAGAACTTT	AAAAGTGCTC	ATCATTGGAA	AACGTTCTTC	GGGGCGAAAA	5760
CTCTCAAGGA	TCTTACCGCT	GTTGAGATCC	AGTTGATGT	AACCCACTCG	TGCACCCAAC	5820
TGATCTTCAG	CATCTTTTAC	TTTCACCAGC	GTTTCTGGGT	GAGCAAAAAC	AGGAAGGCAA	5880
AATGCCGCAA	AAAAGGGAAT	AAGGGCGACA	CGGAAATGTT	GAATACTCAT	ACTCTTCCTT	5940
TTTCAATATT	ATTGAAGCAT	TTATCAGGGT	TATTGTCTCA	TGAGCGGATA	CATATTTGAA	6000
TGTATTTAGA	AAAATAAACA	AATAGGGGTT	CCGCGCACAT	TTCCCCGAAA	AGTGCCACCT	6060
GACGTCTAAG	AAACCATTAT	TATCATGACA	TTAACCTATA	AAAATAGGCG	TATCACGAGG	6120
CCCTTTCTGTC	TCGCGCGTTT	CGGTGATGAC	GGTGAAAACC	TCTGACACAT	GCAGCTCCCC	6180
GAGACGGTCA	CAGCTTGTCT	GTAAGCGGAT	GCCGGGAGCA	GACAAGCCCG	TCAGGGCGCG	6240
TCAGCGGGTG	TTGGCGGGTG	TCGGGGCTGG	CTTAACTATG	CGGCATCAGA	GCAGATTGTA	6300
CTGAGAGTGC	AC					6312

Figure 12. FBdelPRDSAF Sequence

1

CATATGCGGT	GTGAAATACC	GCACAGATGC	GTAAGGAGAA	AATACCGCAT	CAGGCGCCAT	60
TCGCCATTCA	GGCTGCGCAA	CTGTTGGGAA	GGGCGATCGG	TGCGGGCCTC	TTCGCTATTA	120
CGCCAGCTGG	CGAAAGGGGG	ATGTGCTGCA	AGGCCATTAA	GTTGGGTAAC	GCCAGGGTTT	180
TCCCAGTCAC	GACGTTGTAA	AACGACGGCC	AGTGAATTCC	GATTAGTTCA	ATTGTGTTAA	240
GACAGGATCT	CAGTAGTCCA	GGCTTTAGTC	CTGACTCAAC	AATACCACCA	GCTAAAACCA	300
CTAGAAATACG	AGCCACAATA	AATAAAAGAT	TTTATTAGT	TTCCAGAAAA	AGGGGGGAAT	360
GAAAGACCCC	ACCAAATTGC	TTAGCCTGAT	AGCCGCAGTA	ACGCCATTTT	GCAAGGCATG	420
GAAAAATACC	AAACCAAGAA	TAGAGAAGTT	CAGATCAAGG	GCGGGTACAC	GAAAACAGCT	480
AACGTTGGGC	CAACACAGGAT	ATCTGCGGTG	AGCAGTTTCG	GCCCCGGCCC	GGGGCCAAGA	540
ACAGATGGTC	ACCGCGGTTT	GGCCCCGGCC	CGGGGCCAAG	AACAGATGGT	CCCCAGATAT	600
GGCCCCAACCC	TCAGCAGTTT	CTTAAGACCC	ATCAGATGTT	TCCAGGCTCC	CCCCAGGACC	660
TGAAATGACC	CTGTGCCTTA	TTTGAATTAA	CCAATCAGCC	TGCTTCTCGC	TCTGTGTCGC	720
GCGCTTCTGC	TTCCCGAGCT	CTATAAAAGA	GCTCACAACC	CCTCACTCGG	CGCGCCAGTC	780
CTCCGATAGA	CTGAGTCGCC	CGGGTACCCG	TGTATCCAAT	AAATCCTCTT	GCTGTTGCAT	840
CCGACTCGTG	TCTCTCGTGT	TCCTTGGGAG	GGTCTCCTCA	GAGTGATTGA	CTACCCGTCT	900
CGGGGGTCTT	TCATTTGGGG	GCTCGTCCGG	GATCTGGAGA	CCCCTGCCCA	GGGACCACCG	960
ACCCACCACC	GGGAGGTAAG	CTGGCCAAGA	TCCCCCGGGC	TGCAGGAATT	TATGAAATCC	1020
TTTATGGGGG	ACCCCCCCTT	TTGTCAACCT	TGCTCAATTC	CTTCTCCCCC	TCCGATCCTA	1080
AGACTGATTT	ACAAGCCCCG	CTAAAAGGGC	TGCAAGGCGT	GCAGGCCCAA	ATCTGGACAC	1140
CCCTGGCCGA	ATTGTACCGG	CCAGGACATC	CACAACTAG	CCACCCATTT	CAGGTGGGAG	1200
ACTCCGTGTA	CGTCCGGCGG	CACCGCTCTC	AAGGATTGGA	GCCTCGTTGG	AAGGGACCTT	1260
ACATCGTCTT	GCTGACCAAG	CCCACCGCCA	TAAAGGTTGA	CGGGATCGCC	GCCTGGATTG	1320
ACGCATCGCA	CGCCAAGGCA	GCCCCAAAAA	CCCCTGGACC	AGAAACTCCC	AAAACCTGGA	1380
AGCTCCGCCG	TTCCGGAGAAC	CCTCTTAAGA	TAAGACTCTC	CCGTGCTCTG	CTGCTAATCC	1440
ACCTTGTCCT	TGTACTAACC	CAAAATGAAA	CTCCCAACAG	GAATGGTCAT	TTTATGTAGC	1500
CTAATAATAG	TTCCGGCCAG	GTTTGACGAC	CCCCGCCAAG	CTATCGCATT	AGTACAAAAA	1560
CAACATGGTA	AACCATGCGA	ATGCAGCGGA	GGGCAGGTAT	CCGAGGCCCC	ACCGAACTCC	1620
ATCCAACAGG	TAACTTGCCG	AGGCAAGACG	GCCTACTTAA	TGACCAACCA	AAAATGAAAA	1680
TGCAGAGTCA	CTCCAATAAT	CTCACCTAGC	GGGGGAGAAC	TCCAGAAGTG	CCCCTGTAAC	1740
ACTTTCCAGG	ACTCGATGCA	CAGTTCTTGT	TATACTGAAT	ACCGGCAATG	CAGGCGAATT	1800
AATAAGACAT	ACTACACGGC	CACCTTGCTT	AAAATACGGT	CTGGGAGCCT	CAACGAGGTA	1860
CAGATATTAC	AAAACCCCAA	TCAGCTCCTA	CAGTCCCCTT	GTAGGGGCTC	TATAAATCAG	1920
CCCGTTTGCT	GGAGTGCCAC	AGCCCCCATC	CATATCTCCG	ATGGTGGAGG	ACCCCTCGAT	1980
ACTAAGAGAG	TGTGGACAGT	CCAAAAAAGG	CTAGAACAAA	TTCATAAGGC	TATGACTCCT	2040
GAACTTCAAT	ACCACCCCTT	AGCCCTGCCC	AAAGTCAGAG	ATGACCTTAG	CCTTGATGCA	2100
CGGACTTTTG	ATATCCTGAA	TACCACCTTT	AGGTTACTCC	AGATGTCCAA	TTTATGCTTT	2160
GCCCAAGATT	GTTGGCTCTG	TTTAAACTA	GGTACCCTTA	CCCCTCTTGC	GATACCCACT	2220
CCCTCTTTAA	CCTACTCCCT	AGCAGACTCC	CTAGCGAATG	CCTCCTGTCA	GATTATACCT	2280
CCCCTCTTGG	TTCAACCGAT	GCAGTTCTCC	AACCTGCTCT	GTTTATCTTC	CCCTTTTCAT	2340
AACGATACGG	AACAAATAGA	CTTAGGTGCA	GTCACCTTTA	CTAAGTCAC	CTCTGTAGCC	2400
AATGTGAGTA	GTCCTTTATG	TGCCCTAAAC	GGGTGAGTCT	TCCTCTGTGG	AAATAACATG	2460
GCATACACCT	ATTTACCCCA	AACTGGGACC	AGACTTTGCG	TCCAAGCCTC	CCTCCTCCCC	2520
GACATTGACA	TTAAGCCGGG	GGATGAGCCA	GTCCCATTC	CTGCCATTGA	TCATTATATA	2580
CATAGACCTA	AACGAGCTGT	ACAGTTTCATC	CCTTTACTAG	CTGGACTGGG	AATCACCGCA	2640
GCATTACCA	CCGGAGCTAC	AGGCCTAGGT	GTCTCCGTCA	CCCAGTATAC	AAAATTATCC	2700
CATCAGTTAA	TATCTGATGT	CCAAGTCTTA	TCCGGTACCA	TACAAGATT	ACAAGACCAG	2760
GTAGACTCGT	TAGCTGAAGT	AGTTCTCCAA	AATAGGAGGG	GACTGGACCT	ACTAACGGCA	2820
GAACAAGGAG	GAATTTGTTT	AGCCTTACAA	GAAAAATGCT	GTTTTTATGC	TAACAAGTCA	2880
GGAATTGTGA	GAAACAAAAT	AAGAACCCTA	CAAGAAGAAT	TACAAAAACG	CAGGGAAGAC	2940
CTGGCAACCA	ACCCTCTCTG	GACCGGGCTG	CAGGGCTTTC	TTCCGTACCT	CCTACCTCTC	3000
CTGGGACCCC	TACTCACCTT	CCTACTCATA	CTAACCATTG	GGCCATGCGT	TTTCAGTCGC	3060
CTCATGGCCT	TCATTAATGA	TAGACTTAAT	GTTGTACATG	CCATGGTGCT	GGCCCAGCAA	3120
TACCAAGCAC	TCAAAGCTGA	GGAAGAAGCT	CAGGATTGAG	GCGCCTAGTG	TTGACAATTA	3180
ATCATCGGCA	TAGTATACGG	CATAGTATAA	TACGACTCAC	TATAGGAGGG	CCACCATGGC	3240
CAAGTTGACC	AGTGCCGTTT	CGGTGCTCAC	CGCGCGCGAC	GTCGCGGGAG	CGGTGCGAGT	3300
CTGGACCGAC	CGGCTCGGGT	TCTCCCGGGA	CTTCGTGGAG	GACGACTTCG	CCGGTGTGGT	3360
CCGGGACGAC	GTGACCCTGT	TCATCAGCGC	GGTCCAGGAC	CAGGTGGTGC	CGGACAACAC	3420
CCTGGCCTGG	GTGTGGGTGC	GCGGCCTGGA	CGAGCTGTAC	GCCGAGTGGT	CGGAGGTCGT	3480
GTCCACGAAC	TTCCGGGAGG	CCTCCGGGCC	GGCCATGACC	GAGATCGGCG	AGCAGCCGTG	3540
GGGGCGGGAG	TTCCGCCCTG	GCGACCCGGC	CGGCAACTGC	GTGCACCTTC	TGGCCGAGGA	3600
GCAGGACTGA	NNNNCGGACC	GGTCGACTTG	TTAACTTGTT	TATTGCAGCT	TATAATGGTT	3660
ACAAATAAAG	CAATAGCATC	ACAAATTTCA	CAAAATAAGC	ATTTTTTTCA	CTGCATTCTA	3720
GTTGTGGTTT	GTCCAAACTC	ATCAATGTAT	CTTATCATGT	CTGGAATCCAG	ATCTGGGCCC	3780
ATGCGGCCGC	GGATCGATNN	NNACATGTGA	GCAAAAGGCC	AGCAAAAGGC	CAGGAACCGT	3840
AAAAAGGCCG	CGTTGCTGGC	GTTTTTCCAT	AGGCTCCGCC	CCCCGACGA	GCATCACAAA	3900
AATCGACGCT	CAAGTCAGAG	GTGGCGAAAC	CCGACAGGAC	TATAAGATA	CCAGGCGTTT	3960
CCCCCTGGAA	GCTCCCTCGT	GCGCTCTCCT	GTTCCGACCC	TGCCGCTTAC	CGGATACCTG	4020
TCCGCCTTTC	TCCCTTCGGG	AAGCGTGGCG	CTTCTCAAT	GCTCAGCTG	TAGGTATCTC	4080

Figure 12. FBdelPRDSAF Sequence

2

AGTTCGGTGT	AGGTCGTTTC	CTCCAAGCTG	GGCTGTGTGC	ACGAACCCCC	CGTTCAGCCC	4140
GACCGCTGCG	CCTTATCCGG	TAACTATCGT	CTTGAGTCCA	ACCCGGTAAG	ACACGACTTA	4200
TCGCCACTGG	CAGCAGCCAC	TGGTAACAGG	ATTAGCAGAG	CGAGGTATGT	AGGCGGTGCT	4260
ACAGAGTTCT	TGAAGTGGTG	GCCTAACTAC	GGCTACACTA	GAAGGACAGT	ATTTGGTATC	4320
TGCGCTCTGC	TGAAGCCAGT	TACCTTCGGA	AAAAGAGTTG	GTAGCTCTTG	ATCCGGCAAA	4380
CAAACCAACG	CTGGTAGCGG	TGGTTTTTTT	GTTTGCAAGC	AGCAGATTAC	GCGCAGAAAA	4440
AAAGGATCTC	AAGAAGATCC	TTTGATCTTT	TCTACGGGGT	CTGACGCTCA	GTGGAACGAA	4500
AACTCACGTT	AAGGGATTTT	GGTCATGAGA	TTATCAAAAA	GGATCTTCAC	CTAGATCCCT	4560
TTAAATTAAA	AATGAAGTTT	TAAATCAATC	TAAAGTATAT	ATGAGTAAAC	TTGGTCTGAC	4620
AGTTACCAAT	GCTTAATCAG	TGAGGCACCT	ATCTCAGCGA	TCTGTCTATT	TCGTTTCATCC	4680
ATAGTTGCCT	GACTCCCCGT	CGTGTAGATA	ACTACGATAC	GGGAGGGCTT	ACCATCTGGC	4740
CCCAGTGCTG	CAATGATACC	GCGAGACCCA	CGCTCACCAG	CTCCAGATTT	ATCAGCAATA	4800
AACCAGCCAG	CCGGAAGGGC	CGAGCGCAGA	AGTGGTCTTG	CAACTTTATC	CGCCTCCATC	4860
CAGTCTATTA	ATTGTGTCCG	GGAAGCTAGA	GTAAGTAGTT	CGCCAGTTAA	TAGTTTGCGC	4920
AACGTTGTGT	CCATTGCTAC	AGGCATCGTG	GTGTACGCT	CGTCGTTTGG	TATGGCTTCA	4980
TTCAGCTCCG	GTTCCCAACG	ATCAAGGCGA	GTTACATGAT	CCCCCATGTT	GTGCAAAAAA	5040
GCGGTTAGCT	CCTTCGGTCC	TCCGATCGTT	GTGAGAAAGT	AGTTGGCCGC	AGTGTTATCA	5100
CTCATGGTTA	TGGCAGCACT	GCATAATTCT	CTTACTGTCA	TGCCATCCGT	AAGATGCTTT	5160
TCTGTGACTG	GTGAGTACTC	AACCAAGTCA	TTCTGAGAAT	AGTGTATGCG	GCGACCGAGT	5220
TGCTCTTGCC	CGGCGTCAAT	ACGGGATAAT	ACCGCGCCAC	ATAGCAGAAC	TTTAAAAGTG	5280
CTCATCATTT	GAAGACGTTT	TTCGGGGCGA	AAACTCTCAA	GGATCTTACC	GCTGTTGAGA	5340
TCCAGTTCGA	TGTAACCCAC	TCGTGCACCC	AACTGATCTT	CAGCATCTTT	TACTTTTACC	5400
AGCGTTTCTG	GGTGAGCAAA	AACAGGAAGG	CAAAATGCCG	CAAAAAAGGG	AATAAGGGCG	5460
ACACGGAAAT	GTTGAATACT	CATACTCTTC	CTTTTTCAT	ATTATTGAAG	CATTTATCAG	5520
GGTTATTGTC	TCATGAGCGG	ATACATATTT	GAATGTATTT	AGAAAAATAA	ACAAATAGGG	5580
GTTCCGCGCA	CATTTCCTCC	AAAAGTGCCA	CCTGACGTCT	AAGAAACCAT	TATTATCATG	5640
ACATTAACCT	ATAAAAAATAG	GCGTATCACG	AGGCCCTTTC	GTCTCGCGCG	TTTCGGTGAT	5700
GACGGTGAAA	ACCTCTGACA	CATGCAGCTC	CCGGAGACGG	TCACAGCTTG	TCTGTAAGCG	5760
GATGCCGGGA	GCAGACAAGC	CCGTACGGGC	GCGTCAGCGG	GTGTTGGCGG	GTGTCGGGGC	5820
TGGCTTAAC	ATGCGGCATC	AGAGCAGATT	GTACTGAGAG	TGCAC		5865

Figure 13. hCMV10A1 Sequence

1

AGATCTCCCG	ATCCCCCTATG	GTCGACTCTC	AGTACAATCT	GCTCTGATGC	CGCATAGTTA	60
AGCCAGTATC	TGCTCCCTGC	TTGTGTGTTG	GAGGTGCTG	AGTAGTGC	GAGCAAAATT	120
TAAGCTACAA	CAAGGCAAGG	CTTGACCGAC	AATGTCATGA	AGAATCTGCT	TAGGGTTAGG	180
CGTTTTGCGC	TGCTTCGCGA	TGTACGGGCC	AGATATACGC	GTTGACATTG	ATTATTGACT	240
AGTTATTAAT	AGTAATCAAT	TACGGGGTCA	TTAGTTCATA	GCCCATATAT	GGAGTTCGCG	300
GTTACATAAC	TTACGGTAAA	TGGCCCGCCT	GGCTGACCGC	CCAACGACCC	CCGCCCATTG	360
ACGTCAATAA	TGACGTATGT	TCCCATAGTA	ACGCCAATAG	GGACTTTCCA	TTGACGTCAA	420
TGGGTGGACT	ATTTACGGTA	AACTGCCCAC	TTGGCAGTAC	ATCAAGTGTA	TCATATGCCA	480
AGTACGCCCT	CTATTGACGT	CAATGACGGT	AAATGGCCCC	CCTGGCATTA	TGCCCAGTAC	540
ATGACCTTAT	GGGACTTTCC	TACTTGGCAG	TACATCTACG	TATTAGTCAT	CGCTATTACC	600
ATGGTGATGC	GGTTTTGGCA	GTACATCAAT	GGGCGTGGAT	AGCGGTTTGA	CTCACGGGGA	660
FTTCCAAGTC	TCCACCCCAT	TGACGTCAAT	GGGAGTTTGT	TTTGGCACCA	AAATCAACGG	720
GACTTTCCAA	AATGTCGTAA	CAACTCCGCC	CCATTGACGC	AAATGGGCGG	TAGGCGTGTA	780
CGGTGGGAGG	TCTATATAAG	CAGAGCTCTC	TGGCTAACTA	GAGAACCAC	TGCTTAACTG	840
GCTTATCGAA	ATGTCGACTG	AGAACTTCAG	GGTGAGTTTG	GGGACCCCTG	ATTGTTCTTT	900
CTTTTTCGCT	ATTGTAAAT	TCATGTTATA	TGGAGGGGGC	AAAGTTTTCA	GGGTGTTGTT	960
TAGAAATGGGA	TACTTTCTCT	TGTATCACCA	TGGACCCCTCA	TGATAATTTT	GTTTCTTTCA	1020
CTTTCTACTC	TGTTGACAAC	CATTGTCTCC	TCTTATTTTC	TTTTCATTTT	CTGTAACCTT	1080
TTGCTTAAAC	TTTAGCTTGC	ATTGTAAACG	AATTTTTAAA	TTCACCTTTG	TTTATTTGTC	1140
AGATTGTAAG	TACTTTCTCT	AATCACTTTT	TTTTCAAGGC	AATCAGGGTA	TATTATATTG	1200
TACTTCAGCA	CAGTTTTAGA	GAACAATTGT	TATAATTAAA	TGATAAGGTA	GAATATTTC	1260
GCATATAAAT	TCTGGCTGGC	GTGGAATAT	TCTTATTGGT	AGAAACAAC	ACATCCTGGT	1320
CATCATCCTG	CCTTTCTCTT	TATGGTTACA	ATGATATACA	CTGTTTGAGA	TGAGGATAAA	1380
ATACTCTGAG	TCCAAACCGG	GCCCCCTGCG	TAACCATGTT	CATGCCTTCT	TCTTTTCTCT	1440
ACAGTCCTCG	GGCAACGTGC	TGGTTGTTGT	GCTGTCTCAT	CATTTTGGCA	AGGATCGGCC	1500
GGACACGAGT	CAGGACCGAC	ATGGAAGGTC	CAGCGTTCTC	AAAACCCCTT	AAAGATAAGA	1560
TTAACCCGTG	GAAGTCCTTA	ATGGTCATGG	GGTCTATTT	AAGAGTAGGG	ATGGCAGAGA	1620
GCCCCATCA	GGTCTTTAAT	GTAACCTGGA	GAGTCACCAA	CCTGATGACT	GGGCGTACCG	1680
CCAATGCCAC	CTCCCTTTTA	GGAACGTGAC	AAGATGCCTT	CCCAAGATTA	TATTTTGATC	1740
TATGTGATCT	GGTCGGAGAA	GAGTGGGACC	CTTCAGACCA	GGAACCATAT	GTCCGGGTATG	1800
GCTGCAAAATA	CCCCGGAGGG	AGAAAGCGGA	CCCGGACTTT	TGACTTTTAC	GTGTGCCCTG	1860
GGCATACCGT	AAAATCGGGG	TGTGGGGGGC	CAAGAGAGGG	CTACTGTGGT	GAATGGGGTT	1920
GTGAAACCAC	CGGACAGGCT	TACTGGAAGC	CCACATCATC	ATGGGACCTA	ATCTCCCTTA	1980
AGCGCGGTAA	CACCCCTTGG	GACACGGGAT	GCTCCAAAT	GGCTGTGGC	CCCTGCTACG	2040
ACCTCTCCAA	AGTATCCAAT	TCCTTCCAAG	GGGCTACTCG	AGGGGGCAGA	TGCAACCCTC	2100
TAGTCTTAGA	ATTCACTGAT	GCAGGAAAAA	AGGCTAATTG	GGACGGGGCC	AAATCGTGGG	2160
GACTGAGACT	GTACCGGACA	GGAACAGATC	CTATTACCAT	GTCTCCCTG	ACCCGCCAGG	2220
TCCTCAATAT	AGGGCCCCGC	ATCCCCATTG	GGCCTAATCC	CGTGATCACT	GGTCAACTAC	2280
CCCCCTCCCG	ACCCGTGCGAG	ATCAGGCTCC	CCAGGCCTCC	TCAGCCTCCT	CCTACAGGCG	2340
CAGCCTCTAT	AGTCCCTGAG	ACTGCCCCAC	CTTCTCAACA	ACCTGGGACG	GGAGACAGGC	2400
TGCTAAACCT	GTGAGAAGGA	GCCTATCAGG	CGCTTAAACCT	CACCAATCCC	GACAAGACCC	2460
AAGAAATGTTG	GCTGTGCTTA	GTGTCCGGAC	CTCCTTATTA	CGAAGGAGTA	GCGGTGCTGG	2520
GCACCTATAC	CAATCATCT	ACCGCCCCGG	CCAGCTGTAC	GGCCACTTCC	CAACATAAGC	2580
TTACCCATATC	TGAAGTGACA	GGACAGGGCC	TATGCATGGG	AGCACTACCT	AAAACCTCAC	2640
AGGCCTTATG	TAACACCACC	CAAAGTGCCG	GCTCAGGATC	CTACTACCTT	GCAGCACCCG	2700
CTGGAACAAT	GTGGGCTTGT	AGCACTGGAT	TGACTCCCTG	CTTGCTCCAC	ACGATGCTCA	2760
ATCTAACCCAC	AGACTATTGT	GTATTAGTTG	AGCTCTGGCC	CAGAATAATT	TACCACTCCC	2820
CCGATTATAT	GTATGGTCAG	CTTGAACAGC	GTACCAATA	TAAGAGGGAG	CCAGTATCGT	2880
TGACCTTGGC	CCTTCTGCTA	GGAGGATTAA	CCATGGGAGG	GATTGCAGCT	GGAATAGGGA	2940
CGGGGACCAC	TGCCCTAATC	AAAACCCAGC	AGTTTGAGCA	GCTTCACGCC	GCTATCCAGA	3000
CAGACCTCAA	CGAAGTCGAA	AAATCAATTA	CCAACCTAGA	AAAGTCACTG	ACCTCGTTGT	3060
CTGAAGTAGT	CCTACAGAAC	CGAAGAGGCC	TAGATTTGCT	CTTCTTAAAA	GAGGGAGGTC	3120
TCTGCGCAGC	CCTAAAAGAA	GAATGTTGTT	TTTATGCAGA	CCACACGGGA	CTAGTGAGAG	3180
ACAGCATGGC	CAAACCTAAG	GAAAGGCTTA	ATCAGAGACA	AAAACCTATT	GAGTCAGGCC	3240
AAGGTTGGTT	CGAAGGGCAG	TTTAATAGAT	CCCCCTGGTT	TACCACCTTA	ATCTCCACCA	3300
TCATGGGACC	TCTAATAGTA	CTCTTACTGA	TCTTACTCTT	TGGACCCTGC	ATTCTCAATC	3360
GATTAGTTCA	ATTTGTTAAA	GACAGGATCT	CAGTAGTCCA	GGCTTTAGTC	CTGACTCAAC	3420
AATACCACCA	GCTAAAGCCT	ATAGAGTACG	AGCCATAGGG	CGCCTAGTGT	TGACAATTAA	3480
TCATCGGCAT	AGTATACGGC	ATAGTATAAT	ACGACTCACT	ATAGGAGGGC	CACCATGGCC	3540
AAGTTGACCA	GTGCCGTTCC	GGTGCTCACC	GCGCGCGACG	TCGCCGGAGC	GGTCGAGTTC	3600
TGGACCGACC	GGCTCGGGTT	CTCCCGGGAC	TTCTGTGGAG	ACGACTTCGC	CGGTGTGGTC	3660
CGGGACGACG	TGACCCTGTT	CATCAGCGCG	GTCCAGGACC	AGGTGGTGCC	GGACAACACC	3720
CTGGCCCTGGG	TGTGGGTGCG	CGGCCTGGAC	GAGCTGTACG	CCGAGTGGTC	GGAGGTCGTG	3780
TCCACGAAC	TCCGGGACGC	CTCCGGGCCG	GCCATGACCG	AGATCGGCGA	GCAGCCGTGG	3840
GGGCGGGAGT	TGCCCCTGCG	CGACCCGGCC	GGCAACTGCG	TGCACTTCGT	GGCCGAGGAG	3900
CAGGACTGAN	NNNCGGACCG	GTCGA				3925

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/GB 96/02061

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C12N15/86 C12N5/10 C12N15/67

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	JOURNAL OF VIROLOGY 69 (7). 1995. 4086-4094. ISSN: 0022-538X, July 1995, XP002023654 LUUKKONEN B G M ET AL: "Efficiency of reinitiation of translation on human immunodeficiency virus type 1 mRNAs is determined by the length of the upstream open reading frame and by intercistronic distance." see the whole document ---	1-29
A	VIROLOGY (1995), 208(1), 215-25 CODEN: VIRLAX; ISSN: 0042-6822, 1 April 1995, XP002023655 HERZOG, ETIENNE ET AL: "Translation of the second gene of peanut clump virus RNA 2 occurs by leaky scanning in vitro" see the whole document ---	1-29
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☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

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Date of the actual completion of the international search

23 January 1997

Date of mailing of the international search report

12.02.97

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax (+31-70) 340-3016

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Hornig, H

INTERNATIONAL SEARCH REPORT

International Application No.

PCT/GB 96/02061

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
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A	J. VIROL. (1993), 67(8), 4886-95 CODEN: JOVIAM; ISSN: 0022-538X, August 1993, XP000616337 FOUILLOT, NATHALIE ET AL: "Translation of the hepatitis B virus P gene by ribosomal scanning as an alternative to internal initiation" see the whole document ---	1-29
A	VIROLOGY, vol. 188, no. 1, May 1992, ACADEMIC PRESS, INC., NEW YORK, US, pages 342-352, XP002023656 C.-G. LIN AND S.J. LO: "Evidence for involvement of a ribosomal leaky scanning mechanism in the translation of the hepatitis B virus Pol gene from the viral pregenome RNA" see the whole document ---	1-29
A	VIROLOGY, vol. 185, no. 2, December 1991, ACADEMIC PRESS, INC., NEW YORK, US, pages 862-866, XP000616129 F.-L. COSSET ET AL.: "Newcastle disease virus (NDV) vaccine based on immunization with avian cells expressing the NDV hemagglutinin-neuraminidase glycoprotein" cited in the application see the whole document ---	1-29
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C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	NATURE (LONDON) (1984), 309(5963), 82-5 CODEN: NATUAS; ISSN: 0028-0836, 3 May 1984, XP002023657 LIU, CHUNG CHENG ET AL: "Initiation of translation at internal AUG codons in mammalian cells" see the whole document ---	1-29
A	WO,A,94 24870 (BIOTRANSPLANT INC ;GEN HOSPITAL CORP (US); LE GUERN CHRISTIAN A (U) 10 November 1994 see the whole document ---	1-29
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